**Figure S1. KGB-1 suppresses *mir-71* expression cell nonautonomously. (A)** Representative images of *cdc-25.1* RNAi-sterilized transgenic animals expressing GFP from the *mir-71* promoter and KGB-1 from tissue-specific promoters, as designated, following a 2-day exposure, beginning at L4, to control (EV) or *vhp-1* RNAi. Pharyngeal yellow fluorescence results from bleed through of a *myo-2p::tdTomato* co-injection marker into the green channel – this area was avoided in signal quantification. Scale bar, 200 µm. **(B)** Quantification of signal intensity in worms such as in A. Lines mark averages ± SE; individual measurements shown in dots, 25-31 worms per group; \*\* p<0.01, \*\*\* p<0.001, t-test.

**Figure S2.** ***mir-71* mediates effects of KGB-1 activation on DAF-16 output in adults.** Representative images of *cdc-25.1* RNAi-sterilized worms of the indicated genetic backgrounds expressing a DAF-16::GFP fusion protein following a two-day exposure, beginning at L4, to control (EV) or *vhp-1* RNAi. Scale bar, 200 µm.

**Figure S3.** ***mir-71* is not required for KGB-1 dependent enhancement of DAF-16 function in larvae. (A)** Arepresentative image of L3 transgenic animals expressing a DAF-16::GFP fusion protein following exposure, from egg stage, to control (EV) or *vhp-1* RNAi. Filled arrowheads mark worms with strong nuclear localization, empty arrowheads mark weak nuclear localization. Scale bar, 100 µm. **(B)** Portion of worms with DAF-16::GFP nuclear localization as in A, with genetic backgrounds and RNAi treatments as designated. Shown are averages and SDs of 3 independent experiments (N=346-718 worms total per group); \* p<0.05, t-test. **(C)** qRT-PCR measurements of *mtl-1* gene expression in L4 larvae of strains with RNAi treatments as designated. Shown are averages ± SDs for 2 independent experiments.