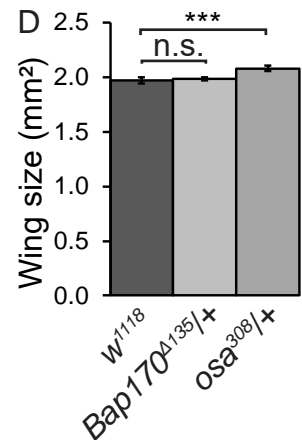
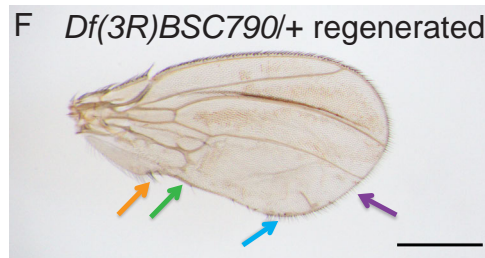
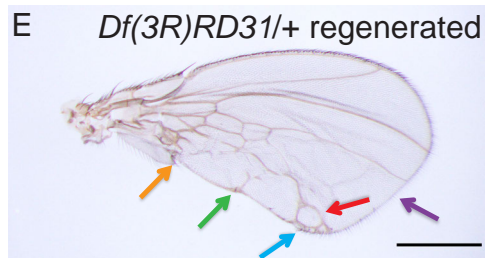
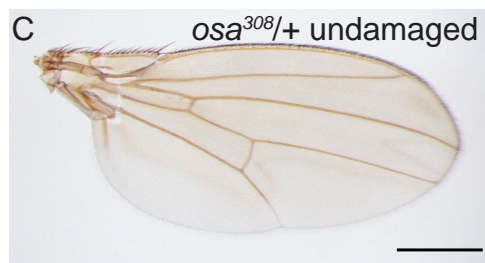
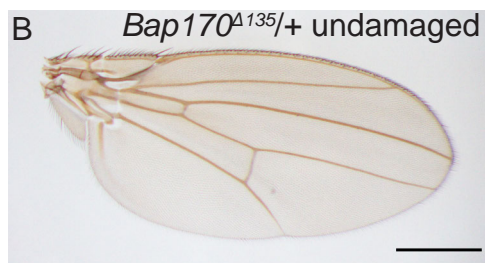
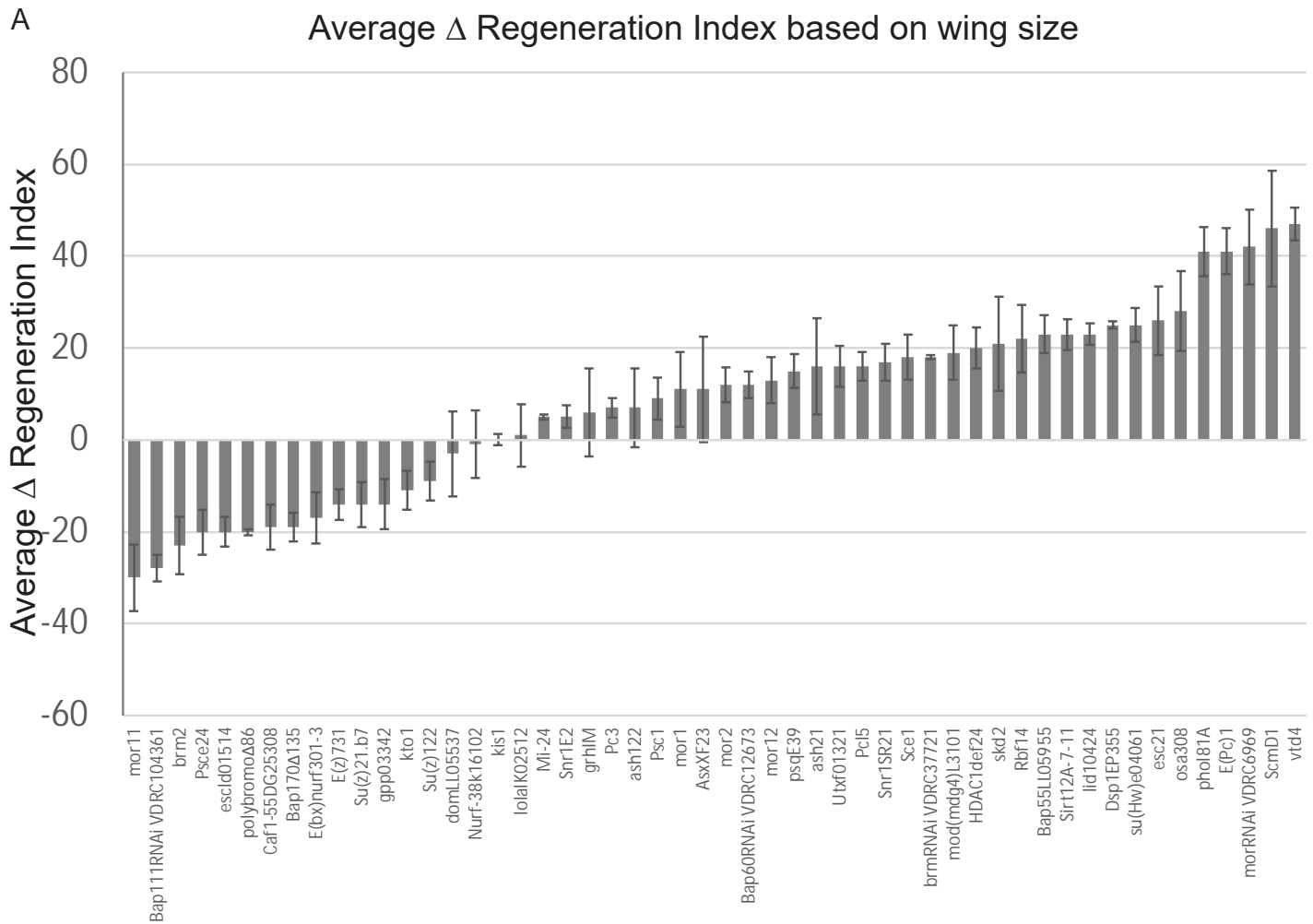


Allele or RNAi	Complex	Regeneration Phenotype	Δ Index	Phenotype Strength
<i>Pc</i> ³	PRC1		7%	
<i>Psc</i> ¹	PRC1		9%	
<i>Psc</i> ^{e24}		Reduced	-20%	**
<i>Sce</i> ¹	PRC1	Enhanced	18%	*
<i>Scm</i> ^{D1}	PRC1	Enhanced	46%	***
<i>E(z)</i> ⁷³¹	PRC2	Reduced	-14%	*
<i>Su(z)</i> ¹² ²	PRC2		-9%	
<i>esc</i> ²¹	PRC2	Enhanced	26%	**
<i>Caf1-55</i> ^{DG25308}	PRC2, NURF	Reduced	-19%	*
<i>esc</i> ^{l01514}	PRC2	Reduced	-20%	**
<i>pho</i> ^{l81A}	PhoRC	Enhanced	41%	***
<i>ash2</i> ¹	COMPASS, COMPASS-like	Enhanced	16%	*
<i>Utx</i> ^{f01321}	COMPASS-like	Enhanced	16%	*
<i>ash1</i> ²²	ASH1		7%	
<i>E(bx)</i> ^{nurf301-3}	NURF	Reduced	-17%	*
<i>Nurf-38</i> ^{k16102}	NURF		-1%	
<i>Mi-2</i> ⁴	NuRD		5%	
<i>brm</i> ²	SWI/SNF (BAP & PBAP)	Reduced	-23%	**
<i>brm</i> ^{RNAi VDRC37721}		Enhanced	18%	*
<i>osa</i> ³⁰⁸	SWI/SNF(BAP)	Enhanced	28%	**
<i>Bap170</i> ^{Δ135}	SWI/SNF(PBAP)	Reduced	-19%	*
<i>polybromo</i> ^{Δ86}	SWI/SNF(PBAP)	Reduced	-20%	**
<i>Snr1</i> ^{E2}	SWI/SNF (BAP & PBAP)		5%	
<i>Snr1</i> ^{SR21}		Enhanced	17%	*
<i>mor</i> ¹	SWI/SNF (BAP & PBAP)	Enhanced	11%	*
<i>mor</i> ²		Enhanced	12%	*
<i>mor</i> ¹¹		Reduced	-30%	***
<i>mor</i> ¹²		Enhanced	13%	*
<i>mor</i> ^{RNAi VDRC6969}		Enhanced	42%	***
<i>Bap55</i> ^{LL05955}	SWI/SNF (BAP & PBAP)	Enhanced	23%	**
<i>Bap60</i> ^{RNAi VDRC12673}	SWI/SNF (BAP & PBAP)	Enhanced	12%	*
<i>Bap111</i> ^{RNAi VDRC104361}	SWI/SNF (BAP & PBAP)	Reduced	-28%	**
<i>psq</i> ^{E39}	GBP	Enhanced	15%	*

<i>Rbf</i> ^{f4}	dREAM	Enhanced	22%	**
<i>Dsp1</i> ^{EP355}		Enhanced	25%	**
<i>grh</i> ^{IM}			6%	
<i>lola</i> ^{K02512}			1%	
<i>Pcl</i> ⁵		Enhanced	16%	*
<i>HDAC1</i> ^{def24}	Sin3A/HDAC complex	Enhanced	20%	**
<i>Sirt1</i> ^{12A-7-11}	SIRT1-LSD1 complex	Enhanced	23%	**
<i>vtd</i> ⁴	cohesin	Enhanced	47%	***
<i>Su(z)2</i> ^{1.b7}		Reduced	-14%	*
<i>gpp</i> ⁰³³⁴²		Reduced	-14%	*
<i>mod(mdg4)</i> ^{L3101}		Enhanced	19%	*
<i>su(Hw)</i> ^{e04061}	gypsy chromatin insulator complex	Enhanced	25%	**
<i>lid</i> ¹⁰⁴²⁴		Enhanced	23%	**
<i>Asx</i> ^{XF23}		Enhanced	11%	*
<i>dom</i> ^{LL05537}	TIP60 complex		-3%	
<i>E(Pc)</i> ¹	TIP60 complex	Enhanced	41%	***
<i>kis</i> ¹			0%	
<i>kto</i> ¹	mediator	Reduced	-11%	*
<i>skd</i> ²	mediator	Enhanced	21%	**

Table S1. Screen of Chromatin Regulators. For each line, the regeneration index was calculated by summing the product of approximate wing size (0%, 25%, 50%, 75% and 100%) and the corresponding percentage of wings for each wing size. The Δ Index, which is the difference between the regeneration indices of the line being tested and the control tested simultaneously, was calculated by subtracting the regeneration index of the control from the regeneration index of the mutant or RNAi line. A cutoff Δ index of 10% was set, over which we considered the regenerative capacity to be affected. Green indicates lines that had a higher regeneration index compared to the control, purple indicates lines that had a lower regeneration index compared to the control. * = Δ Index ≥ 10 or ≤ -10 , ** = Δ Index ≥ 20 or ≤ -20 , *** = Δ Index ≥ 30 or ≤ -30 .



H Quantification of P-to-A transformations

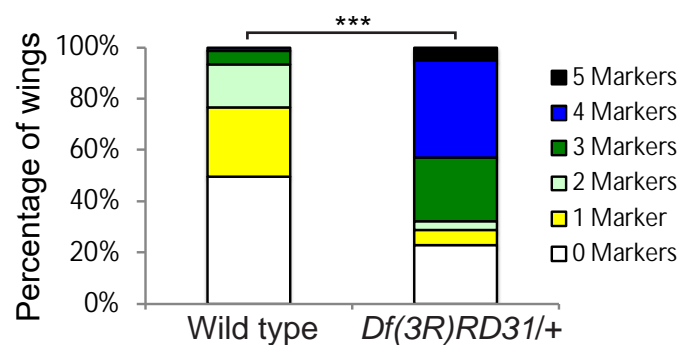


Figure S1. Roles of the SWI/SNF components in regeneration and development

(A) Average Δ regeneration index of chromatin regulator mutants and RNAi lines screen. The Δ regeneration Index is the difference between the regeneration indices of the line being tested and the control tested simultaneously. Δ regeneration index was calculated as described in materials and methods.

(B) Adult *Bap170* ^{Δ 135/+} wing that developed from an undamaged disc

(C) Adult *osa*^{308/+} wing that developed from an undamaged disc

(D) Quantification of adult wing sizes in females that did not have damaged discs. *w*¹¹¹⁸ (n=16), *Bap170* ^{Δ 135/+} (n=16), *osa*^{308/+} (n=16). Results for males were similar.

(E-G) Adult wings after damage and regeneration of the disc in *Df(3R)RD31/+*

(E), *Df(3R)BSC790/+* (F), and *osa*^{00090/+} (G) animals. Arrows show five anterior-specific markers in the posterior compartment: anterior crossveins (red), alula-like costa bristles (orange), margin vein (green), socketed bristles (blue), and change of wing shape with wider distal portion of the wing, similar to the anterior compartment (purple).

(H) Quantification of the number of Posterior-to-Anterior transformation markers in each wing after damage and regeneration of the disc, using wings that were 75% normal size or larger, comparing *Df(3R)RD31/+* wings to wild-type (*w*¹¹¹⁸)

wings, n = 56 wings (*Df(3R)RD31/+*) and 48 wings (*w¹¹¹⁸*), from 3 independent experiments. Chi-square test $p < 0.001$.

Error bars are s.e.m. Scale bars are 500 μ m for all adult wings images. * $p < 0.05$,

** $p < 0.01$, *** $p < 0.001$ Student's *t*-test.

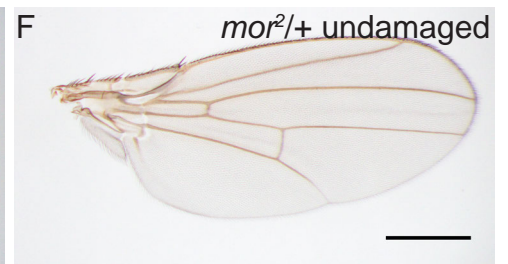
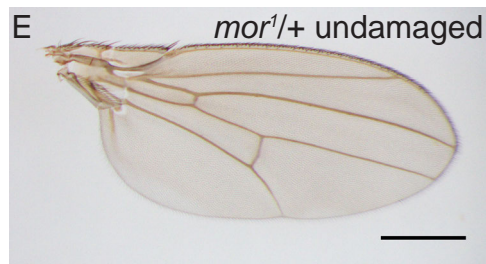
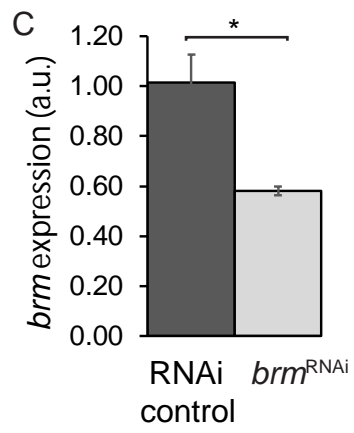
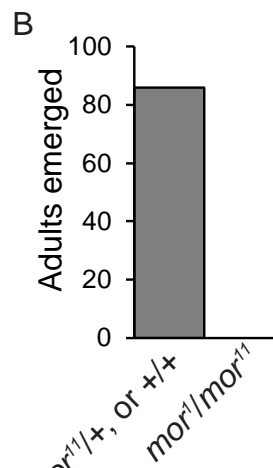
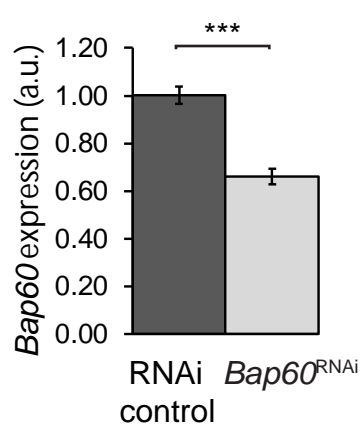


Figure S2. Controls for SWI/SNF mutants and RNAi lines

(A) *Bap60* expression examined by qPCR of *Bap60*^{RNAi} and control undamaged wing discs. The graph shows fold change relative to control discs. RNAi lines were crossed to *w*¹¹¹⁸; +; *rn-GAL4*, *tubGAL80^{ts}/TM6B* and kept at 18°C.

Temperature shift to 30°C at day 7 for 24 hours then back to 18°C. Wing discs of non-Tubby larvae were dissected at 24 hours after shifting back to 18°C.

(B) Complementation test for *mor*¹ and *mor*¹¹ mutants.

(C) *brm* expression examined by qPCR of *brm*^{RNAi} and control undamaged wing discs. The graph shows fold change relative to control discs. RNAi lines were crossed to *w*¹¹¹⁸; +; *rn-GAL4*, *tubGAL80^{ts}/TM6B* and kept at 18°C. Temperature shift to 30°C at day 7 for 24 hours then back to 18°C. Wing discs of non-Tubby larvae were dissected at 24 hours after shifting back to 18°C.

(D-F) Adult wings from undamaged discs: *Bap55*^{LL05955/+} (D), *mor*^{1/+} (E) and *mor*^{2/+} (F).

Error bars are S.E.M.. Scale bars are 500µm for all adult wings images. * p < 0.05, ** p < 0.01, *** p < 0.001 Student's *t*-test for (A) and (C).

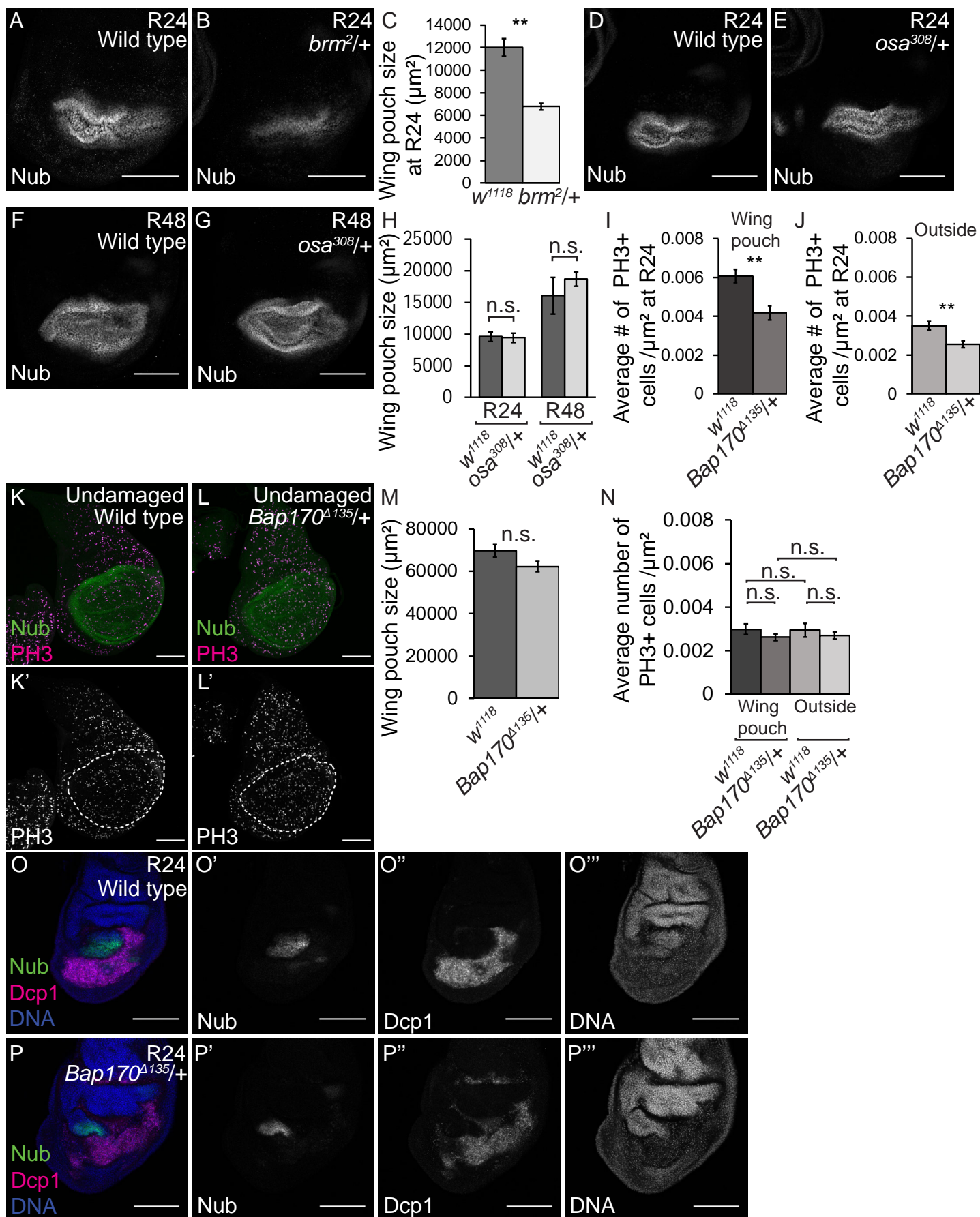


Figure S3. The PBAP complex is required for regenerative growth whereas the BAP complex is not.

(A) Wild-type (w^{1118}) regenerating wing disc at R24 with wing pouch marked by anti-Nubbin immunostaining.

(B) $brm^2/+$ regenerating wing disc at R24 with wing pouch marked by anti-Nubbin immunostaining.

(C) Comparison of regenerating wing pouch size at 24 hours after imaginal disc damage in $brm^2/+$ and wild-type (w^{1118}) animals. n = 11 wing discs ($brm^2/+$) and 10 wing discs (w^{1118}).

(D) Wild-type (w^{1118}) regenerating wing disc at R24 with wing pouch marked by anti-Nubbin immunostaining.

(E) $osa^{308}/+$ regenerating wing disc at R24 with wing pouch marked by anti-Nubbin immunostaining.

(F) Wild-type (w^{1118}) regenerating wing disc at R48 with wing pouch marked by anti-Nubbin immunostaining.

(G) $osa^{308}/+$ regenerating wing disc at R48 with wing pouch marked by anti-Nubbin immunostaining.

(H) Comparison of regenerating wing pouch size at 24 and 48 hours after imaginal disc damage and regeneration in $osa^{308}/+$ and wild-type (w^{1118}) animals.

At R24, n = 26 wing discs (*osa*^{308/+}) and 27 wing discs (*w*¹¹¹⁸). At R48, n = 6 wing discs (*osa*^{308/+}) and 21 wing discs (*w*¹¹¹⁸).

(I) Average number of mitotic cells (marked with PH3 immunostaining) per μm^2 in the regenerating wing primordium at R24 in *Bap170* ^{Δ 135/+} and wild-type (*w*¹¹¹⁸) animals. n = 8 wing discs (*Bap170* ^{Δ 135/+}) and 10 wing discs (*w*¹¹¹⁸).

(J) Average number of mitotic cells (marked with PH3 immunostaining) per μm^2 outside the regenerating wing primordium at R24 in *Bap170* ^{Δ 135/+} and wild-type (*w*¹¹¹⁸) animals. n = 8 wing discs (*Bap170* ^{Δ 135/+}) and 10 wing discs (*w*¹¹¹⁸).

(K-L) Undamaged wild-type (*w*¹¹¹⁸) (K) and *Bap170* ^{Δ 135/+} (L) wing discs at R24 with Nubbin (green) and PH3 (magenta, grayscale in K' and L') immunostaining. Dashed white outline shows the wing primordium labeled with Nubbin.

(M) Comparison of wing pouch size in *Bap170* ^{Δ 135/+} (n=17) and wild-type (*w*¹¹¹⁸)(n=17) animals age-matched to R24 but without tissue damage (maintained at 18°C).

(N) Average number of mitotic cells (marked with PH3 immunostaining) per μm^2 within or outside the undamaged wing pouch in *Bap170* ^{Δ 135/+} (n=17 for the wing pouch and 16 outside the wing pouch) and wild-type (*w*¹¹¹⁸)(n=17 for the wing pouch and 15 outside the wing pouch) animals age-matched to R24 but without tissue damage (maintained at 18°C).

(O) Wild-type (w^{1118}) regenerating wing disc at R24 with Nubbin (green) (O') and cleaved caspase Dcp1 (magenta)(O'') immunostaining marking the debris field, and DNA (blue) was detected with Topro3 here in subsequent panel. (O''').

(P) $bap170^{\Delta 135/+}$ regenerating wing disc at R24 with Nubbin (green)(P') and cleaved caspase Dcp1 (magenta)(P'') immunostaining, and DNA (blue)(P''').

Error bars are S.E.M.. Scale bars are 100 μ m for all wing discs images. * $p < 0.05$, ** $p < 0.01$, Student's t -test.

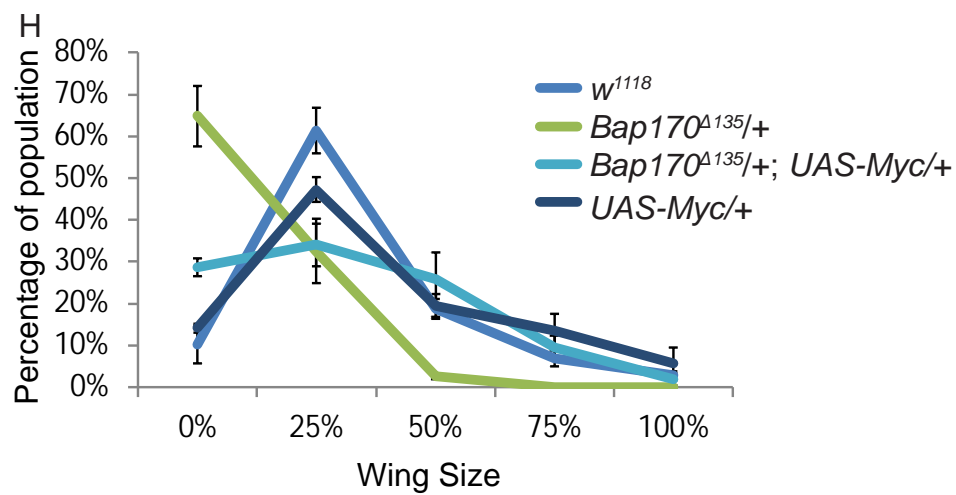
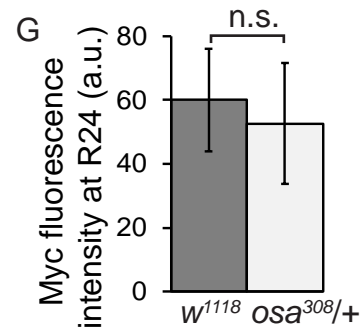
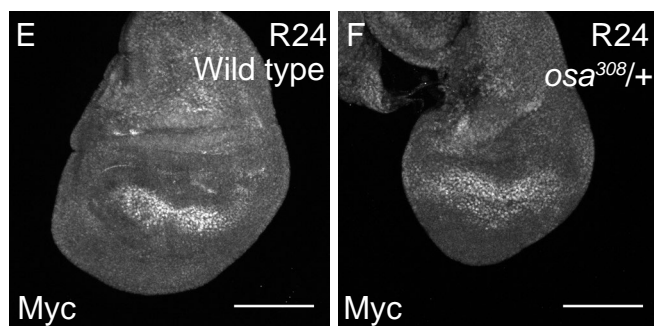
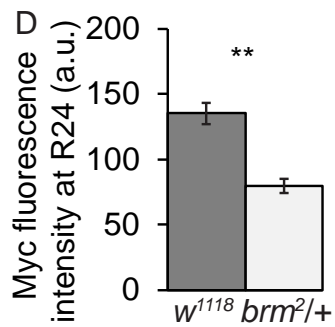
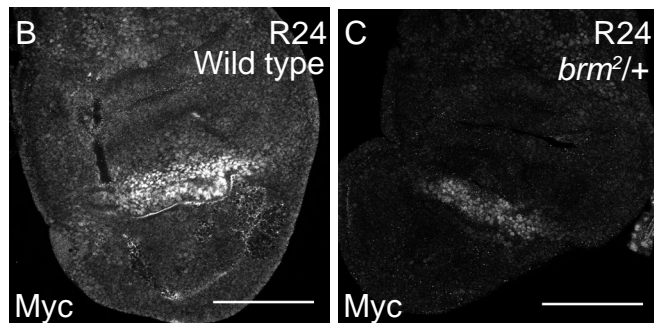
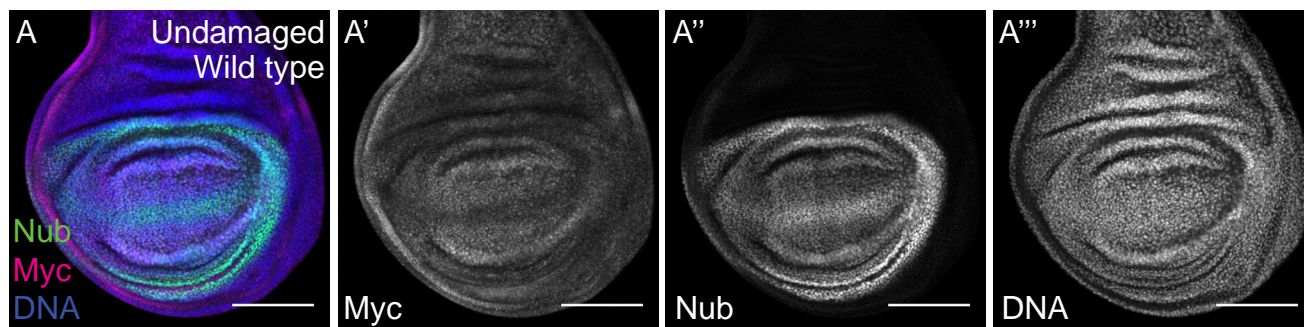


Figure S4. The PBAP complex regulates Myc during regeneration

(A) Wild-type (w^{1118}) undamaged wing disc with Myc (magenta) (A') and Nubbin (green) (A'') immunostaining. DNA (blue) (A''') was detected with Topro3.

(B-C) Wild-type (w^{1118}) (B) and $brm^2/+$ (C) regenerating wing discs at R24 with Myc immunostaining.

(D) Quantification of anti-Myc immunostaining fluorescence intensity in the wing pouch in $brm^2/+$ and wild-type (w^{1118}) regenerating wing discs at R24. $n = 11$ wing discs ($brm^2/+$) and 12 wing discs (w^{1118}).

(E-F) Wild-type (w^{1118}) (E) and $osa^{308}/+$ (F) regenerating wing discs at R24 with Myc immunostaining.

(G) Quantification of anti-Myc immunostaining fluorescence intensity in the wing pouch in $osa^{308}/+$ and wild-type (w^{1118}) regenerating wing discs at R24. $n = 28$ wing discs ($osa^{308}/+$) and 27 wing discs (w^{1118}).

(H) Comparison of the size of adult wings after imaginal disc damage and regeneration in wild-type (w^{1118}), $Bap170^{\Delta135}/+$, $Bap170^{\Delta135}/+; UAS-Myc/+$, and $UAS-Myc/+$ animals. $n = 364$ wings (w^{1118}), 194 wings ($Bap170^{\Delta135}/+$), 194 wings ($Bap170^{\Delta135}/+; UAS-Myc/+$) and 294 wings ($UAS-Myc/+$) from 3 independent experiments. Chi-square test for $Bap170^{\Delta135}/+$ and $Bap170^{\Delta135}/+; UAS-Myc/+$ has $p < 0.001$

Error bars are S.E.M.. Scale bars are 100 μ m for all wing discs images. * $p < 0.05$, ** $p < 0.01$, Student's t -test for (D) and (G).

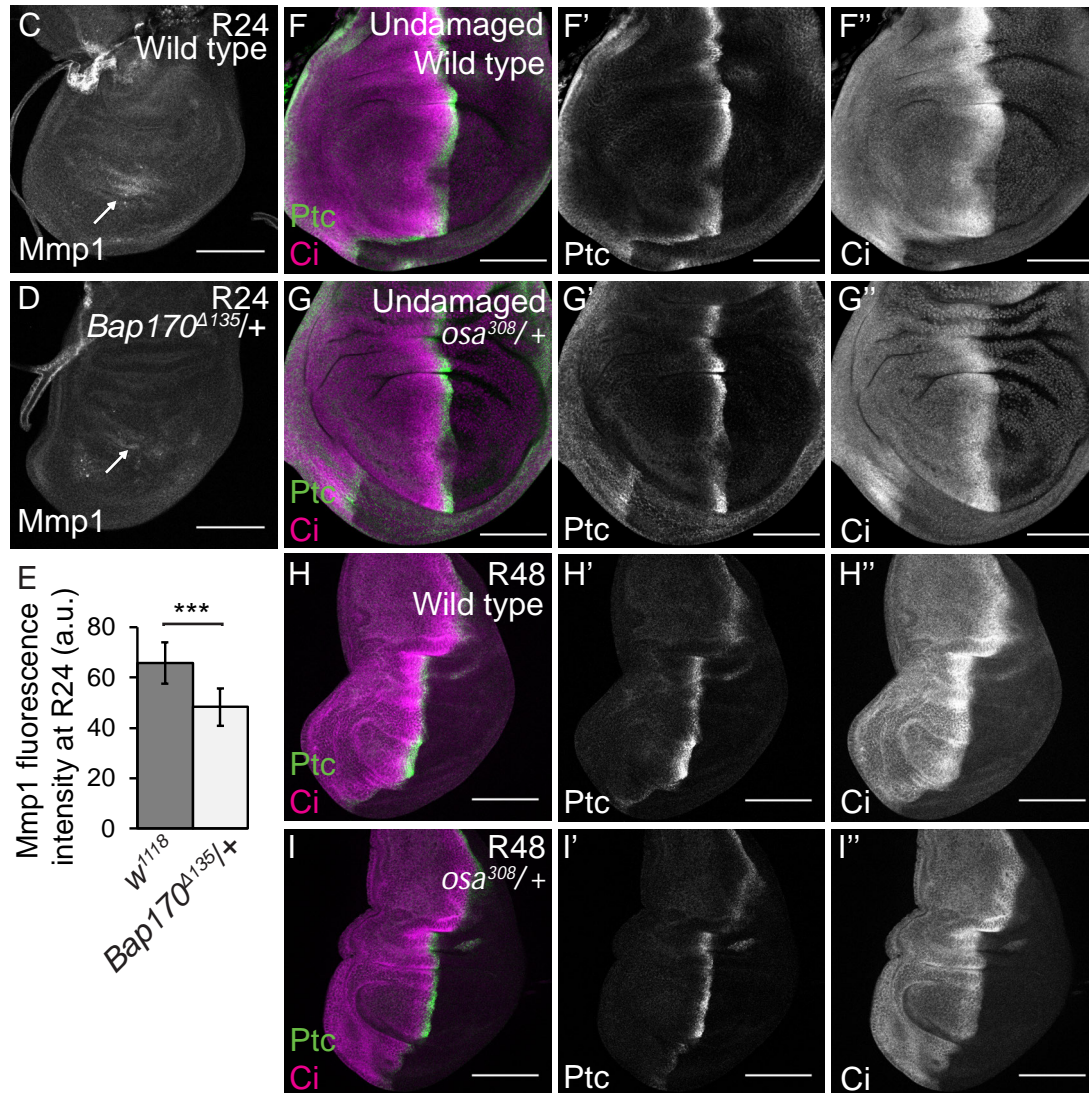
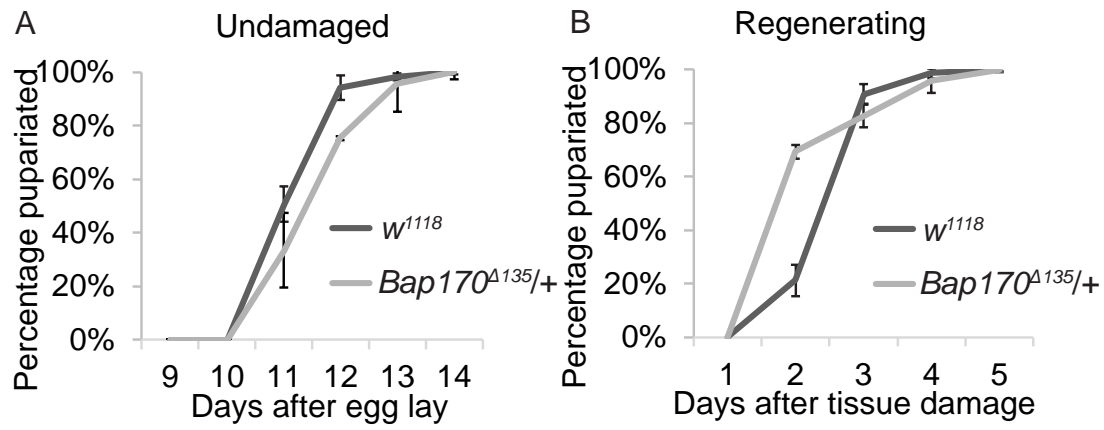


Figure S5. The function of BAP and PBAP in regeneration and development

(A) Pupariation rates of animals during normal development at 18°C. n = 103 pupae (*Bap170^{Δ135/+}*) and 227 pupae (*w¹¹¹⁸*) from 3 independent experiments.

Student's *t*-test not significant.

(B) Pupariation rates of animals after tissue damage (30°C) and regeneration (18°C). n = 117 pupae (*Bap170^{Δ135/+}*) and 231 pupae (*w¹¹¹⁸*) from 3 independent experiments. Because the temperature shift to 30°C in the ablation protocol increases the developmental rate, the pupariation timing of regenerating animals (B) cannot be compared to the undamaged control animals (A). Chi-square test $p < 0.001$.

(C-E) *mmp1* expression examined by immunofluorescence in wild-type (*w¹¹¹⁸*)

(C) and *Bap170^{Δ135/+}* (D) regenerating wing discs at R24. Quantification in (E).

n= 19 (*w¹¹¹⁸*) and 17 (*Bap170^{Δ135/+}*), error bars are S.E.M., $p=0.00041$.

(F) Wild-type (*w¹¹¹⁸*) undamaged wing disc with Ptc (green)(F') and Ci

(magenta)(F'') immunostaining.

(G) *osa^{308/+}* undamaged wing disc with Ptc (green)(G') and Ci (magenta)(G'')

immunostaining.

(H) Wild-type (*w¹¹¹⁸*) regenerating wing disc at R48 with Ptc (green)(H') and Ci

(magenta)(H'') immunostaining.

(l) *osa*³⁰⁸/+ regenerating wing disc at R48 with Ptc (green)(l') and Ci
(magenta)(l'') immunostaining.

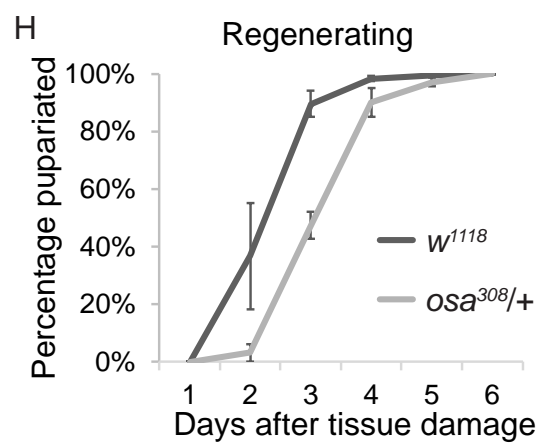
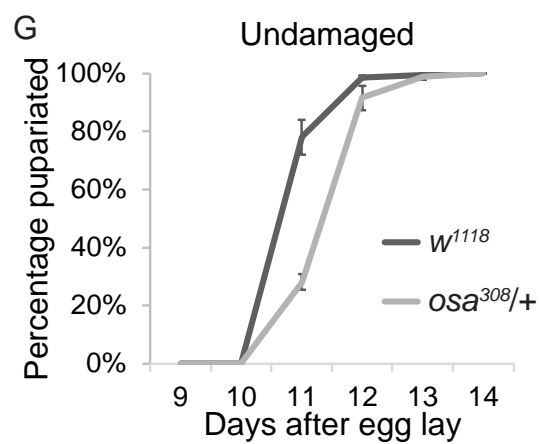
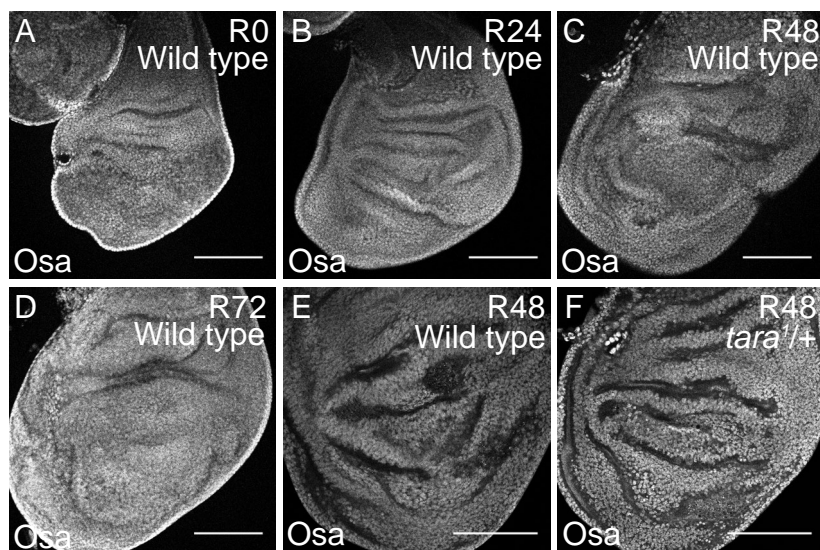


Figure S6. *osa* does not change expression or affect pupariation after tissue damage

(A-D) Wild-type (w^{1118}) regenerating wing discs at 0, 24, 48, and 72 hours after imaginal disc damage with Osa immunostaining.

(E-F) Wild-type (w^{1118}) (E) and *tara*^{1/+} (F) regenerating wing discs at R48 with Osa immunostaining.

(G) Pupariation rates of animals during normal development at 18°C. n = 79 pupae (*osa*^{308/+}) and 173 pupae (w^{1118}) from 3 independent experiments. Chi-square test $p < 0.001$, student's t-test at day 11 $p < 0.01$.

(H) Pupariation rates of animals after tissue damage (30°C) and regeneration (18°C). n = 101 pupae (*osa*^{308/+}) and 155 pupae (w^{1118}) from 3 independent experiments. Chi-square test $p < 0.001$, student's t-test at day 3 $p < 0.01$. Because the temperature shift to 30°C in the ablation protocol increases the developmental rate, the pupariation timing of regenerating animals (H) cannot be compared to the undamaged control animals (G).

Scale bars are 100µm for all wing discs images. Scale bars are 100µm for all wing discs images. Error bars are S.E.M. except where noted.