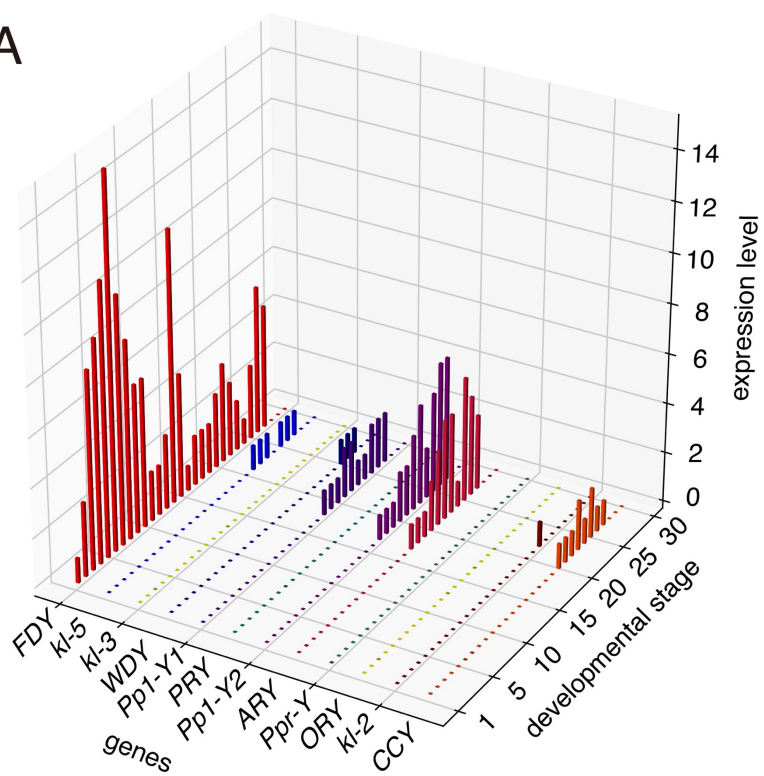


A



B

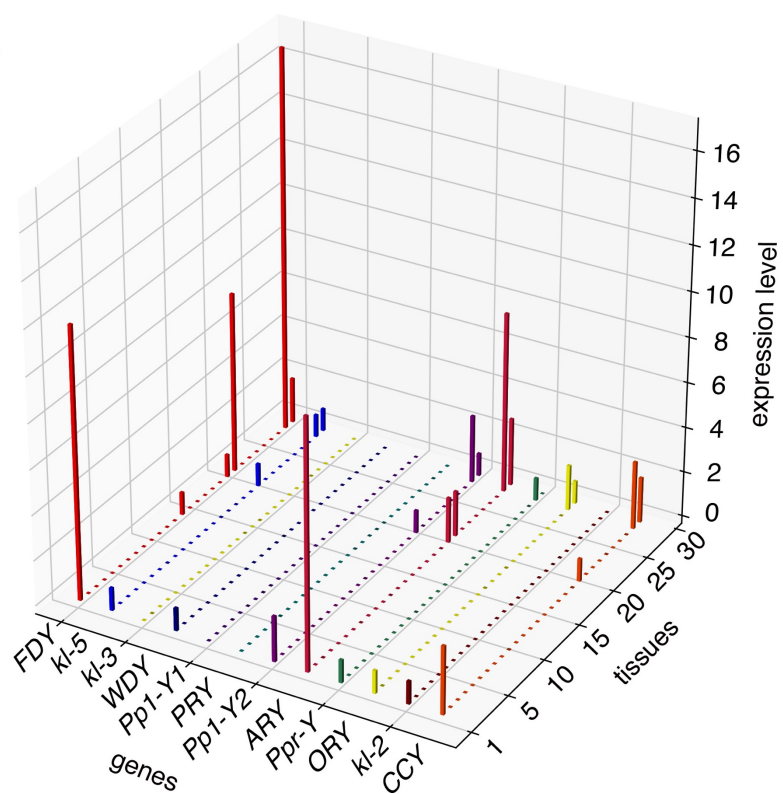


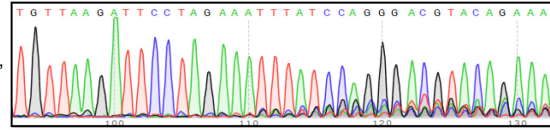
Figure S1 Expression of Y chromosome genes during different developmental stages and in different tissues. (A) Expression of the indicated genes from FlyBase. **X axis**, genes. **Y axis**, numbers from **1-30** indicate different development stages. **1-12** represent different 2 hour windows during embryogenesis (e.g. 1, 0-2 hr; 12, 22-24 hr). **13-18**, different larval stages: 13, L1; 14, L2; 15, L3 12 h; 16, L3 puffstage 1-2; 17, L3 puffstage 3-6; 18, L3 puffstage 7-9. **19-24**, different pupal stages: 19, white prepupae; 20, prepupae 12 hr; 21, pupae 1 day; 22, pupae 2 day; 23, pupae 3 day; 24, pupae 4 day. **25-30**, adults: 25, 1-day old males; 26, 5-day old males; 27, 30-day old males; 28, 1-day old females; 29, 5-day old females; 30, 30-day old females. Z axis, relative expression levels (arbitrary units). (B) Expression of the indicated genes in different tissues from FlyBase. **X axis**, genes. **Y axis**, numbers from **1-30** indicate different tissues. **1**, imaginal disc, larvae L3 wandering. **2**, central nervous system of larvae L3. **3**, central nervous system of pupae P8. **4-12**, head of adults: 4, virgin female 1 day; 5, virgin female 4 day; 6, virgin female 20 day; 7, mated female 1 day; 8, mated female 4 day; 9, mated female 20 day; 10, mated male 1 day; 11, mated male 4 day; 12, mated male 20 day. **13**, salivary gland of larvae L3 wandering. **14**, salivary gland of white prepupae. **15**, digestive system of larvae L3. **16-18**, digestive system of adults: 16, 1-day old; 17, 4-day old; 18, 20-day old. **19**, fat body of larvae L3. **20**, fat body of white prepupae. **21**, fat body of pupae P8. **22**, carcass of larvae L3 wandering. **23-25**, carcass of adults: 23, 1-day old; 24, 4-day old; 25, 20-day old. **26**, ovary of virgin 4-day female. **27**, ovary of mated 4-day female. **28**, testis of mated 4-day male. **29**, accessory gland of mated 4-day male. Z axis, relative expression levels (arbitrary units).

kl-2

Targeting site DNA

5'-TGTTAAGATTTCCTAGAAATTTATCCAGGGACGTACAGAAA-3'

gRNA

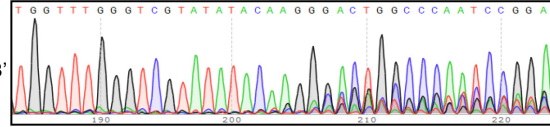


kl-3

Targeting site DNA

5'-TGGTTTGGGTCGTATATACAAGGGACTGGCCCAATCCGGA-3'

gRNA

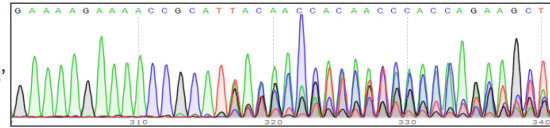


kl-5

Targeting site DNA

5'-GAAAAGAAAACCGCATTATAACGACCTTAACCCAAAAGCT-3'

gRNA

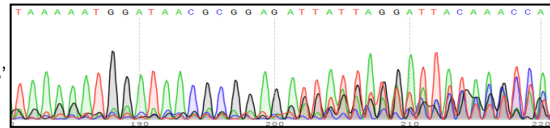


CCY

Targeting site DNA

5'-TAAAAATGGATAACGCGGAGATTAAGAGGATTATAAATCA-3'

gRNA

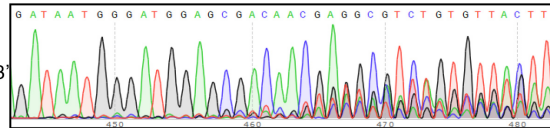


Pp1-Y2

Targeting site DNA

5'-GATAATGGGATGGAGCGACAACGATCGAGGCGTCAGTGTT-3'

gRNA

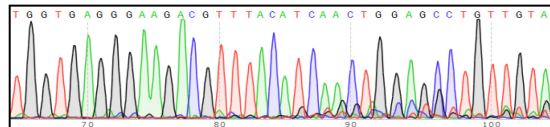


Ppr-Y

Targeting site DNA

5'-TGGTGAGGGAAGACGTTTACATCAACTGGAGCCTGTTGTA-3'

gRNA

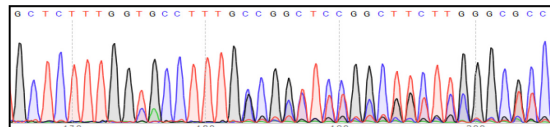


FDY

Targeting site DNA

5'-GCTCTTTGGCACCTTTGGCGCCTTTGCCGGTTCCGGCTTC-3'

gRNA

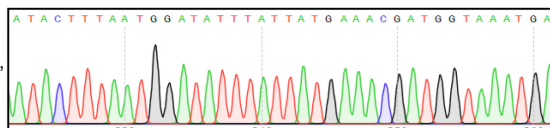


PRY

Targeting site DNA

5'-ATACTTTAATGGATATTTATTATGAAACGATGGTAAATGA-3'

gRNA

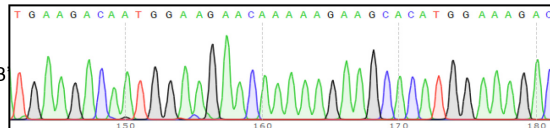


ORY

Targeting site DNA

5'-TGAAGACAATGGAAGAACAAAAAGAAGCACATGGAAAGAC-3'

gRNA



ARY

Targeting site DNA

5'-ATACACAATGTGGAGAAAAGGCTTTACTAATGGCCCCAAA-3'

gRNA

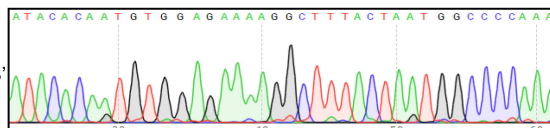


Figure S2 Targeted indel mutations generated by CRISPR/Cas9. The genes are indicated. The underlined regions represent the gRNA sequences. The sequencing chromatograms from F₀ males are shown on the right.

CCY

GGAAGACAGCTTAATAAATG**CGG**CCGATGACGCAATCGATTGGTTACAAGATCTTATGAAAAGAAACATCCTTAACAA
CCAGCTACCTGAATCAGACATGAAGCTATTATCGATAAAAATGGATAACGCGGAGATTAAG**AGG**ATTATAA *w*¹¹⁸

GGAAGACAGCTTAATAAATG**CGG**CCGATGACGCAATCGATTGGTTACAAGATCTTATGAAAAGAAACATCCTTAACAA
CCAGCTACCTGAATCAGACATGAAGCTATTATCGATAAAAATGGATAACGCGGAGATTA -----TAA [-9]

Pp1-Y2

CTATGGTCAGACCCCGATCCAAAGATAATGGGATGGAGCGACAACGATCG**AGG**CGTCAGT *w*¹¹⁸

CTATGGTCAGACCCCGATCC-----GAGGCGTCAGT [-29]

CTATGGTCAGACCCCGATCCAAAGATAATGGGATGGAGCGA-----GGCGTCAGT [-10]

CTATGGTCAGACCCCGATCCAAAGA-----GGCGTCAGT [-26]

Ppr-Y

AAACACGTTTCATCGTGGTGAGGGAAGACGTTTACATCAAC**TGG**AGCCTGTTGTATTGGAA *w*¹¹⁸

AAACACGTTTCATCGTGGTGAGGGAAGACGTTTAC-----TGGAGCCTGTTGTATTGGAA [-6]

AAACACGTTTCATCGTGGTGAGGGAAGACGTT-ACATCAACTGGAGCCTGTTGTATTGGAA [-1]

AAACACGTTTCATCGTGGTGAGGGAAGACGTTTA-----GCCTGTTGTATTGGAA [-11]

AAACACGTTTCATCGTGGTGAGG-----AGCCTGTTGTATTGGAA [-21]

FDY

GTCGCGGCGCCCAAGAAGCCGGAACCGGCAAAGGCGCCAA**AGG**TGCCAAAGAGCAAGTCG *w*¹¹⁸

GTCGCGGCGCCCAAGAAGCCGGAACCGGCAAAGGCGCCAAAG-----AGCAAGTCG [-9]

Figure S3 Targeted indel mutations identified in each of the indicated genes. The top sequences in each panel are the *w*¹¹⁸ DNA sequences, with the PAM sequences highlighted in red, and the gRNA sequences underlined. Shown below are the indels in F₁ males. The missing nucleotides are represented by red dashes. The number of deleted nucleotides is indicated to the right.

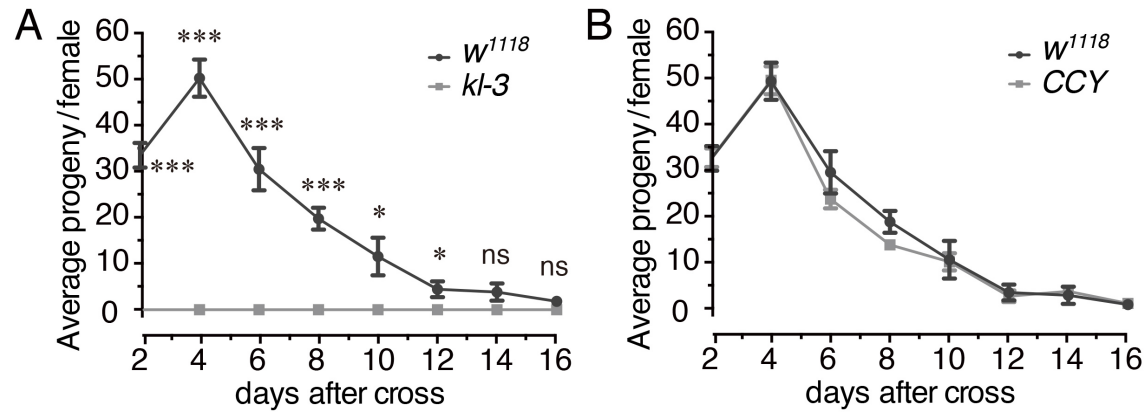


Figure S4 Male fertility tests performed with flies with mutations in Y chromosome genes. (A) *kl-3*. (B) *CCY*. *w¹¹¹⁸* males were used as the control. Means $\pm$ SEMs. Unpaired Student's *t*-tests. *n*=20 crosses each. * *p*<0.05, *** *p*<0.001, ns: not significant.

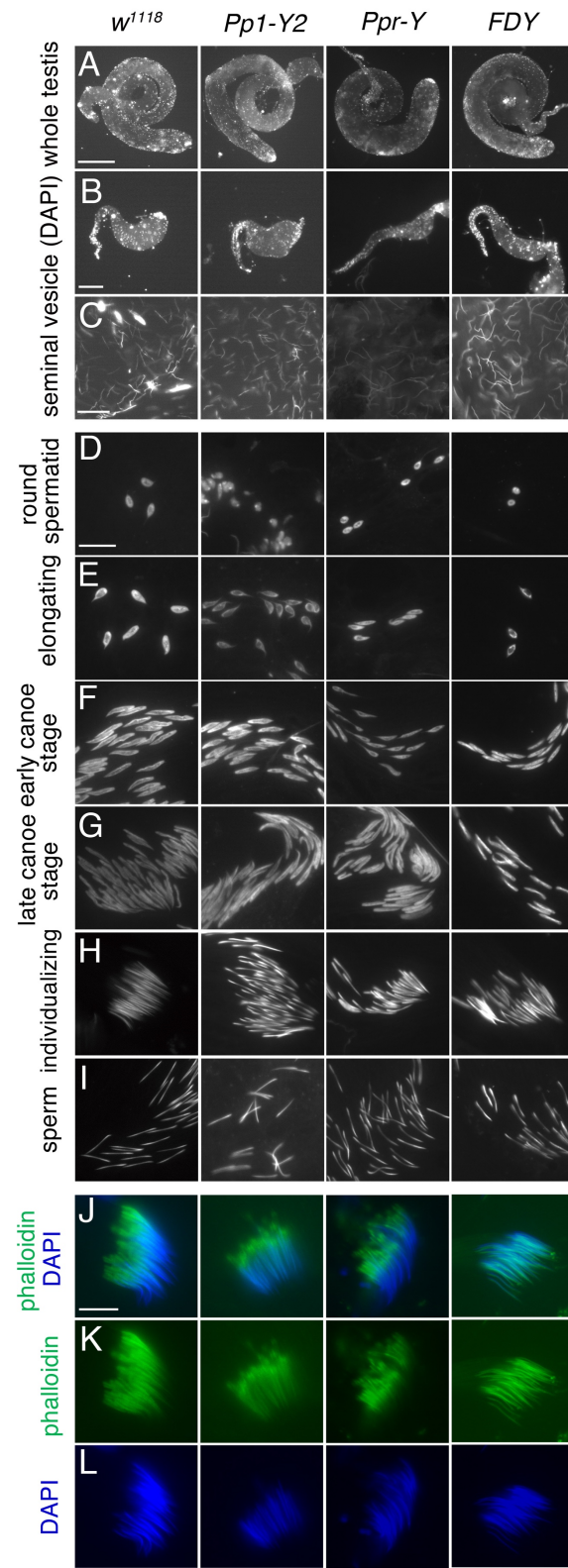


Figure S5 Testing effects of mutations in Y chromosome genes on spermatogenesis. Testes were isolated from mutant males and *w¹¹¹⁸* (control) males and stained with: (A—L) DAPI, and (J—K) and phalloidin. (A-C) Morphology of the testes. (A) Whole testes. (B) Seminal vesicles. (C) Mature sperm in the seminal vesicles. (D—I) Different stages during spermatogenesis. (J—L) Actin and nuclei in individualization complexes (ICs) were stained with phalloidin (green) and DAPI (blue), respectively. Scale bars: (A) 200 μm , (B) 100 μm , (C) 20 μm , (D—L) 10 μm .

Gene	Target site (5' to 3') (PAM is underlined)	oligonucleotide- forward (5' to 3')	oligonucleotide- reverse (5' to 3')	on-target scores	off targets	off-target scores
<i>kl-2</i>	GATTCCTAGAAATT TATCC <u>AGG</u>	TAATACGACTCACTA TAGATTCCTAGAAAT TTATCCAGTTTTAGA GCTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	59.7	3	96.5
<i>kl-3</i>	GGGTCGTATATACA AGGGACT <u>TGG</u>	TAATACGACTCACTA TAGGGTCGTATATAC AAGGGACGTTTTAG AGCTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	55.6	0	99.1
<i>kl-5</i>	GGGTTAAGGTCGT TATAATG <u>CGG</u>	TAATACGACTCACTA TAGGGTTAAGGTCG TTATAATGGTTTTAGA GCTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	65.8	0	49.5
<i>CCY</i>	GGATAACGCGGAG ATTAAG <u>AGG</u>	TAATACGACTCACTA TAGGATAACGCGGA GATTAAGGTTTTAGA GCTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	53.0	3	99.1
<i>Pp1-Y2</i>	GGATGGAGCGACA ACGATCG <u>AGG</u>	TAATACGACTCACTA TAGGATGGAGCGAC AACGATCGGTTTTAG AGCTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	67.7	0	49.3
<i>Ppr-Y</i>	GGGAAGACGTTTA CATCAACT <u>TGG</u>	TAATACGACTCACTA TAGGGAAGACGTTTA CATCAACGTTTTAGA GCTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	41.3	0	99.0
<i>FDY</i>	GGAACCGGCAAAG GCGCCAA <u>AGG</u>	TAATACGACTCACTA TAGGAACCGGCAAA GGCGCCAAGTTTTA GAGCTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	56.1	3	48.9
<i>ORY</i>	GGAAGAACAAAAA GAAGCACAT <u>TGG</u>	TAATACGACTCACTA TAGGAAGAACAAAA AGAAGCACAGTTTTA GAGCTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	63.6	4	81.4
<i>PRY</i>	GGATATTTATTATGA AACGAT <u>TGG</u>	TAATACGACTCACTA TAGGATATTTATTATG AAACGAGTTTTAGAG CTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	64.6	0	48.4
<i>ARY</i>	GGAGAAAAGGCTT TACTAAT <u>TGG</u>	TAATACGACTCACTA TAGGAGAAAAGGCT TTACTAAGTTTTAGA GCTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	55.7	0	95.6

Table S1 Target sites and oligonucleotides used to transcribe gRNAs. The underlined nucleotides are the PAM regions. The sequences of the forward and reverse oligonucleotides used to synthesize the gRNAs are shown. The numbers of off-target sites are indicated. The on-target and off-target scores range from 0-100 (Hsu *et al.* 2013; Doench *et al.* 2016). The on-target score represents the cleavage efficiency of Cas9, with a higher score indicating greater potential effectiveness. The score indicates the probability that a given gRNA will be in top 20% of cleavage activity. The scoring system is not linear, and only

5% of gRNAs receive a score of 60 or higher. The off-target score means the inverse probability of Cas9 having off-target activity. A higher score means the gRNA has less chance to bind to an unintended sequence in the genome.

Gene	Target site (5' to 3') (PAM is underlined)	oligonucleotide-forward (5' to 3')	oligonucleotide-reverse (5' to 3')	off targets
<i>ARY</i>	GAATATATTTCTAACGAC CA <u>AGG</u>	TAATACGACTCACTATAGAAT ATATTTCTAACGACCAGTTTT AGAGCTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	0
	GAAGATAATTCTCAAGT TG <u>CGG</u>	TAATACGACTCACTATAGAA GATAATTCTCAAGTTGGTTT TAGAGCTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	0
	TCTCAAGTTGCGGTGAT ACT <u>TGG</u>	TAATACGACTCACTATATCTC AAGTTGCGGTGATACTGTTT TAGAGCTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	0
	CTTAGATACTTGGCGAG CAAT <u>TGG</u>	TAATACGACTCACTATACTTA GATACTTGGCGAGCAAGTTT TAGAGCTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	0
<i>ORY</i>	GGACAGAAAGAAAGCC AAAA <u>AGG</u>	TAATACGACTCACTATAGGA CAGAAAGAAAGCCAAAAGT TTTAGAGCTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	4
	GAAGATGCTATGAGCAA AAT <u>TGG</u>	TAATACGACTCACTATAGAA GATGCTATGAGCAAAAGTTT TAGAGCTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	0
	AAAACCTAAAGATTTGT TGG <u>GGG</u>	TAATACGACTCACTATAAAAA CTTAAAGATTTGTTGGGTTT TAGAGCTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	0
	TGATGCATCTGATTCTT TCC <u>AGG</u>	TAATACGACTCACTATATGAT GCATCTGATTCTTTCCGTTT TAGAGCTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	0
<i>PRY</i>	AGAGGTACGTCGATTG GAAAT <u>TGG</u>	TAATACGACTCACTATAAGA GGTACGTCGATTGAAAAGT TTTAGAGCTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	2
	GATTCACAGCCTCTACA TTA <u>AGG</u>	TAATACGACTCACTATAGATT CACAGCCTCTACATTAGTTT TAGAGCTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	0
	CCTCGTTCGCCTTACAA AGA <u>GGG</u>	TAATACGACTCACTATACCT CGTTCGCCTTACAAAGAGTT TTAGAGCTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	0
<i>Pp1-Y1</i>	GGAAGATCGATAGAAAA ATGAT <u>TGG</u>	TAATACGACTCACTATAGGA AGATCGATAGAAAAATGAGT TTTAGAGCTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	1
	GGATAGAGGAAAATACT CGGT <u>TGG</u>	TAATACGACTCACTATAGGA TAGAGGAAAATACTCGGGTT TTAGAGCTAGAAATAGC	AAAAAAAGCACCG ACTCGGTGCCAC	1

Table S2 Target sites and oligonucleotides used to transcribe gRNAs to edit *ARY*, *ORY*, *PRY* and *Pp1-Y1*. The underlined nucleotides are the PAM sequences. The sequences of the forward and reverse oligonucleotides used to synthesize the gRNAs are shown. The numbers of off target sites are listed. Note that these gRNAs did not work effectively, so we did not test the fertility of the injected flies.

Gene	Primer name	Primer sequence (5' to 3')
<i>kl-2</i>	<i>kl-2-F</i>	CGAACAAAGCAGGCATTGAAACC
	<i>kl-2-R</i>	GCCCACGACACCGCAATAC
<i>kl-3</i>	<i>kl-3-F</i>	CGATGTCATAGTTGGGATAACTGATG
	<i>kl-3-R</i>	ATTATTATTGTTTACTTACTATATTTGTTGAGCAGCC
<i>kl-5</i>	<i>kl-5-F</i>	CGCGACGATAGACAGCGG
	<i>kl-5-R</i>	GAGAGCAATGCGCTCGTTGC
<i>CCY</i>	<i>CCY-F</i>	CGCGTTTTTGTGCGGATTTAGTG
	<i>CCY-R</i>	GCACTCGCTTCTACACATTTGTGC
<i>Pp1-Y2</i>	<i>PP1-Y2-F</i>	CTCCTCGAGCTTTCCGCACC
	<i>PP1-Y2-R</i>	GAATTCTCCGCAGTAATTTGGTGCAG
<i>Ppr-Y</i>	<i>Ppr-Y-F</i>	CCGAAGTACAGAAGCCCCTTG
	<i>Ppr-Y-R</i>	CCCTTTCCACACTTAAAATGCTTGG
<i>FDY</i>	<i>FDY-F</i>	CCGCTACGAGCTGTTGTTTCATG
	<i>FDY-R</i>	CGAATTCTCCGATCGACTCCTT
<i>PRY</i>	<i>PRY-F</i>	GCGATCATTTTTCTGTATGTGACGACG
	<i>PRY-R</i>	GGCAGCTACTTAACTTCCAATGAGC
<i>ORY</i>	<i>ORY-F</i>	GAGAACGCGACCGGTTAGC
	<i>ORY-R</i>	CCGTTCGGCCATTTGCATTC
<i>ARY</i>	<i>ARY-F</i>	ATGATCCCAGCGGACTTTTTGAC
	<i>ARY-R</i>	GACATGGGCATCAGAATTCATCGC

Table S3 List of primers used to amplify the indicated regions to identify indels by DNA sequencing. F, forward primer. R, reverse primer.

Gene	Primer name	Primer sequence (5' to 3')
<i>kl-2</i>	<i>kl-2</i> -RT-F	GATGTTCTCACTGGTTGCCTTG
	<i>kl-2</i> -RT-R	CACATTGTCTCCCCATTCCATA
<i>kl-3</i>	<i>kl-3</i> -RT-F	GAAGTTCGATGCCTGTTGAACG
	<i>kl-3</i> -RT-R	TGTTGCTGGGCCTAATAAACGA
<i>kl-5</i>	<i>kl-5</i> -RT-F	CCGAACCCATTAGCAAATTTATCA
	<i>kl-5</i> -RT-R	CGTCCATGCAAATAGGAAGAGG
<i>CCY</i>	<i>CCY</i> -RT-F	GGTGCAATCAAGTTATGCAGGA
	<i>CCY</i> -RT-R	CATTAGTTGCTGGTATTGACAATCAT
<i>Pp1-Y2</i>	<i>Pp1-Y2</i> -RT-F	TTTTTCTCCTTCGTGGAAACCA
	<i>Pp1-Y2</i> -RT-R	ACCCTTGTCAGGAACGTCACAT
<i>Ppr-Y</i>	<i>Ppr-Y</i> -RT-F	GAAGTACAGAAGCCCCTTGTGG
	<i>Ppr-Y</i> -RT-R	TGTTCCAATACAACAGGCTCCA
<i>FDY</i>	<i>FDY</i> -RT-F	GCCAAAAACGCTAGTCCAGTGA
	<i>FDY</i> -RT-R	CGAATTCTCCGATCGACTCCTT
<i>WDY</i>	<i>WDY</i> -RT-F	GACCCCCAATGGTTAGGAAATC
	<i>WDY</i> -RT-R	AGATGTCCAGAAGCGCATCATT
<i>PRY</i>	<i>PRY</i> -RT-F	TGCGTTTCATCGTCAAATCTGT
	<i>PRY</i> -RT-R	AACGCTTCTTTTCGCTCACTTG
<i>ORY</i>	<i>ORY</i> -RT-F	ACGTCCCCAAATATTGTCATCG
	<i>ORY</i> -RT-R	TGCGAGGCCTTCACACTTTATT
<i>ARY</i>	<i>ARY</i> -RT-F	TAGATACTTGCGGAGCAATGGA
	<i>ARY</i> -RT-R	ACCAAGAGGTGAAAAGGCTGTC
<i>Pp1-Y1</i>	<i>Pp1-Y1</i> -RT-F	CATCGCTGCTTAGCTGGAAGAT
	<i>Pp1-Y1</i> -RT-R	GCCCAGATGTCTCGAAATAACG

Table S4 List of primers used for RT-PCR. F, forward primer. R, reverse primer.