

Table S1 Primers for strain construction and confirmation

Purpose	Primer	Sequence (5' to 3')
<i>cbp80^Δ::hph</i> confirmation	CBP80-NF	CGAGACATGTTATGGGCTACAC
	CBP80-CR	CACATCCACTTTGTCTGACTGG
<i>gfp/yfpc/yfnp-sad-8</i> construction	SAD-8-E	AACACAACCTCTTTCGGGGCGATT
	SAD-8-NGFP1	GCAGCCTGAATGGCGAATGGACGCGCTGATGTCAATGGCGATGGTAGACAG
	SAD-8-NGFP2	CAGGAGCGGGTGC GG GTGCTGGAGCGATGGATTAGATATTGAGATGGACGTGGA
	SAD-8-F	GGTTCGCAAGCGTTGAAAAGACC
	SAD-8-G	ATAGACGGGCGAGGCGTGTGTC
	SAD-8-H	AATCCATCCGTGCGTCACATCA
	SAD-8-I	GTACGACTGTGCAACGGCATAGC
<i>nup120-mCherry</i> construction	NCU02742-CT-LL	GTACGACTGTGCAACGGCATAGC
	NCU02742-CT-LR	CAGCGCCTGCACCAGCTCCTGCCCCGACATGAGGATGTCCCATCGTCACC
	NCU02742-CT-RL	CTCCTTCAATATCAGTTAAACAAGGGCATGTTGGGCTGCTTGATTGTCCT
	NCU02742-CT-RR	CTTGGTGC GG TTTGAGGTTGGT
	NCU02742-CT-NL	AGACAGATAGGTGAAGGCGACAAGG
	NCU02742-CT-NR	AAGTGGAGCAAGCGAGCGAAAG
<i>r^Δ::hph</i> confirmation	RSP-988505F	GCCACCTTTCCAACCCAATAATGC
	RSP-993102R	TGCCTTTTCAGCTGGAGACCAAGAC
<i>sad-8^Δ::hph</i> confirmation	NCU01310-F1	TTGTGTGTGTGTCGGTGATTATTATTG
	NCU01310-R1	CCTTG TAGCCATCCTCTCGCTGAA
<i>yfpc/yfnp-cbp80</i> construction	A-NT4187LF-LP	TGCTTATTCTTGCCTCAAACCTTTTCG
	B-NT4187LF-RP	GCAGCCTGAATGGCGAATGGACGCGCTGTGTGAAACCCGGTAAGTCAAATAAA
	C-NT4187RF-LP	CAGGAGCGGGTGC GG GTGCTGGAGCGATGGCCGACTACGATCGCAGA
	D-NT4187RF-RP	AACAGCGACATCCTCTGGCTTCC
	E-NT4187-Nest-LP	TGTGGGGCATTTGGGTGTTGTAA
	F-NT4187-Nest-RP	CGGCAACAACGTCTGAGGGATG
<i>yfnp-cbp20</i> construction	A-NTCbp20LF-LP	GAGAAGCTCCGGATTTTGGGCATC
	B-NTCbp20LF-RP	GCAGCCTGAATGGCGAATGGACGCGCGGTTACCTGGTAGTGAAGATGGATT
	C-NTCbp20RF-LP	CAGGAGCGGGTGC GG GTGCTGGAGCGATGTTGAACACTCGCCAAAGGAGC
	D-NTCbp20RF-RP	GATGCTGCCCAAGGACAAGTACAC
	E-NTCbp20-Nest-LP	AGTAGCTTGTGTTGAGGTCTGCT
	F-NTCbp20-Nest-RP	GGCATTCCTTTTCCAGAGCTTCCC
<i>yfnp-sms-2</i> construction	SMS-2-E	GTCCACTTGGTGCCATTCCCACT
	SMS-2-NGFP1	GCAGCCTGAATGGCGAATGGACGCGCGGAGGGTGTCAAAACCTCACAA
	SMS-2-NGFP2	CAGGAGCGGGTGC GG GTGCTGGAGCGATGCTGCTCCTGGCTCTCCC
	SMS-2-F	GTGCCATTCTGCTGCTTCCAGTT
	SMS-2-G	CACTTGCCATACCACGCCATGATT
	SMS-2-H	TGCTCAAACCGCCGTAATTGTTG

Primers for DJ-PCR-based fluorescent tagging were designed as previously described (Hammond *et al.* 2011b). The center DJ-PCR fragment for *nup120-mCherry* tagging was obtained from plasmid pEB1.1, which is similar to pTH112.8 except that *mCherry* (Castro-Longoria *et al.* 2010) is present instead of *yfnp*. The center DJ-PCR fragment for *yfnp-cbp20* and *yfnp-sad-8* tagging was obtained from plasmid pNTYN-2, which is similar to pTH1123.1 except that *nat1* (Lichius and Read 2010) is present instead of *hph*.

LITERATURE CITED

- Castro-Longoria, E., M. Ferry, S. Bartnicki-Garcia, J. Hasty, and S. Brody, 2010 Circadian rhythms in *Neurospora crassa*: dynamics of the clock component frequency visualized using a fluorescent reporter. *Fungal Genet. Biol.* 47: 332–341.
- Lichius, A., and N. D. Read, 2010 A versatile set of Lifeact-RFP expression plasmids for live-cell imaging of F-actin in filamentous fungi. *Fungal Genet. Rep.* 57: 8–14.