

**Table S1** Primers used in this study.

**Figure S1** Expression of *ScTHI5* in *C. glabrata* confers a growth advantage during thiamine starvation conditions. Transformation of *C. glabrata* wild-type cells with a plasmid containing the *CgPMU3* promoter (1000 bp) driving expression of *ScTHI5* either with or without a stop codon at the end of the *ScTHI5* ORF. Without a stop codon, a YFP fusion protein is formed, and we confirmed that expression was thiamine starvation regulated by flow cytometry (left). Streaking these two plasmid containing strains with a third strain containing an empty vector on SD-his-thiamine and replica plating twice to dilute the residual thiamine demonstrated that the *CgPMU3p-ScTHI5* plasmids conferred a growth advantage in the absence of thiamine (right).

**Figure S2** The frequency of abundant sequences increases after selection in thiamine starvation conditions. After selection and next generation sequencing (NGS) of the samples, we calculated the abundance of each 11 bp sequence. We then graphed the frequency of the 20 most abundant sequences in each sample relative to the percent of the total sequences. After selection, the majority of sequences are represented by less than 20 sequences.

**Table S2** Raw data from the seven samples that we sequenced. At the bottom of each column, we indicate how many sequencing reads we acquired from the NGS, and the proportion of those sequences that were unique (i.e. only observed as one sequence out of the total). We randomly acquired an additional 2000-5000 sequences to confirm that we had no bias in selecting a relatively small subset of the total NGS run, and noted only slight changes (i.e. sequences may change from the 10<sup>th</sup> most abundant to the 12<sup>th</sup> most abundant). There were no sequences identified that appeared in multiple samples below the top 25 sequences with either dataset. We judged that the data did not change based on sampling error.

**Table S3** List of sequences observed in wild-type selections 1-3 (Figure S2) that were represented in at least two replicates. While we examined the top 30 sequences in the selection samples, none of the sequences that were less abundant than the top 20 led to a fold enrichment >15x, and given that the fold enrichment still resulted in less than 1% of the final proportion, we believe we have examined all of the relevant sequences.

**Figure S3** Alignment of sequences in Table S3 with *C. glabrata* THI promoters and *CgPMU3* promoter. We aligned the sequences highly enriched by selection with the *CgPMU3* 11 bp element, and other *CgTHI* promoter 13 bp elements, asking what nucleotides were likely required for elevated expression. We note that sequences A-D became abundant post selection, and in most cases were not even observed in the preselection sequences (i.e. less than 1/8986), suggesting that the selection enriched for sequences that conferred elevated expression. Sequences A and G have a lot of similarity to *CgPET18* and sequence E matches *CgPMU3* in 8 of 11 nucleotides, as the surrounding nucleotides confer the GA in front of the CGTACA.

**Figure S4** Scanning mutagenesis of the 13 bp THI UAS in the *CgPET18* promoter suggests

the xxCCGTxxAxxTG nucleotides are critical for expression. Single mutations were introduced into the full-length *CgPET18* promoter, replacing the native nucleotide with an A (or C if an A was in that position) and measuring fluorescence in thiamine starvation conditions.

**Figure S5** Schematic of *S. cerevisiae* promoters and location of mutations and deletions made in this study. Presented is 300 bp of the *ScTHI5* promoter and 200 bp of the *ScTHI20* promoter with both strands shown, as well as the first 15 bp of the ORF. Based on MEME analysis (located under the sequences), we identified regions (boxed within the sequence) that shared similarity to the THI UAS identified in *C. glabrata*. We replaced the entire boxed region in *ScTHI5p* and in *ScTHI20p* with the sequences shown above them, and did not observe a significant decline in expression of the promoter (Figure 7A). Additionally, we replaced the *ScTHI20* region with 13 Ts and did not see a significant decline in expression (data not shown). Realignment of these regions with the THI UAS consensus suggested numerous regions that were similar to the THI UAS (highlighted in yellow). Regions in green indicate putative Thi2 binding sites based on Nosaka, 2006. When we remove regions of the promoter overlapping with the Thi2 binding site (highlighted in grey), we observe a large decline in expression of the promoters (Figure 7B and 7C).

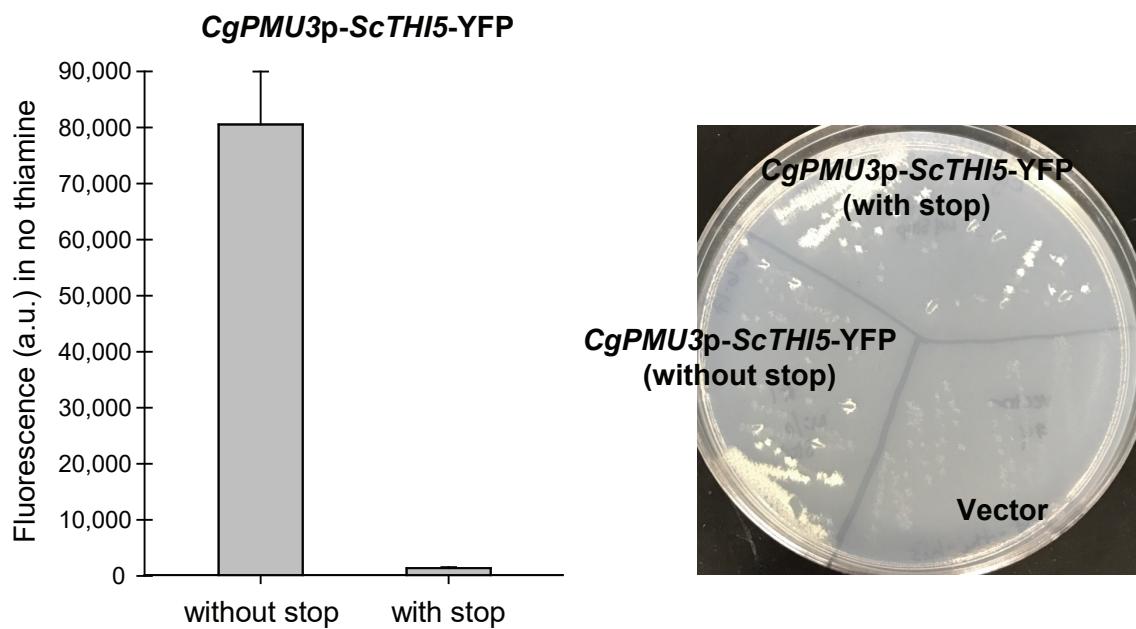
**Figure S6** Expression of *S. cerevisiae* THI promoters is dependent on Pdc2, Thi2, and Thi3. While deleting *ScPDC2* and *ScTHI3* reduces almost all expression of the *ScTHI4* promoter, deletion of *THI2* allows for regulation, albeit with lower expression.

**Table S1** Primers used in this study.

<i>CgPET18 p-YFP-pRS313</i>	-1000 bp	01718	ggtagccggccgcgtctagaactatggatccAGAGATTACTACAACGTTTG	use with 01720
	-400 bp	01719	ggtagccggccgcgtctagaactatggatccTTAGCTGATTCGGAG	use with 01720
	-320 bp	02457	ccgcgggtggccgcgtctagaactatggatccTTAGCTGATTCGGGT	use with 01720
	-318 bp	02458	ccgcgggtggccgcgtctagaactatggatccGATATGGTCCGGTGT	use with 01720
	-314 bp	02459	ccgcgggtggccgcgtctagaactatggatccCTAGATGGTCCGGTAA	use with 01720
	-312 bp	02460	ccgcgggtggccgcgtctagaactatggatccCTAGATGGTCCGGTAA	use with 01720
	-300 bp	01844	ggtagccggccgcgtctagaactatggatccCTAGATGGTCCGGTGT	use with 01720
	-200 bp	01843	ggtagccggccgcgtctagaactatggatccTATTTGTAATAAAGATGTT	use with 01720
	-100 bp	01842	ggtagccggccgcgtctagaactatggatccGAATGAGGTATAAATAGGC	use with 01720
at ATG with homology to YFP (3')		01720	accatgtataatttttaccccttagacatTTTTAACCATTTGGTTA	
<i>PMU3 UASΔ</i>		02599	ACAGCCATACGCCCTTAATggACTATATTGTTGAATAAAC	
		02600	TTTATTACAAAATATTGTCATAAAGCCCGTAGTGTG	use with 01720
<i>THI UAS::PacI</i>		02497	GAGCTAAATAAGGATATTaaacccGTTGATTAAGTAAAGTTATG	use with 01720
		02498	CATAACTTATTCAATTACCGGTTaaataaTATCCTACTTATTAGCTC	use with 01720
<i>THI UAS::PMU3 UAS</i>		02668	ATGGATATCATGAGATGGacgtacaacgAATAAGTTGCACTCG	use with 01720
		02669	TGCAATACTTATGtgtaacgtcCAATGATAATCCATTAT	use with 01720
mutate -305 bp T --> A		02605	ATAAACTGGATATCAGATTGatccgttaatgAATAAGTTATGCATCTC	use with 01720
		02606	GATGCTATACTTATGcatttacaggatCAAATCTGATTCACCTTAT	use with 01720
mutate -304 bp T --> A		02607	ATAAACTGGATATCAGATTGatccgttaatgAATAAGTTATGCATCTC	use with 01720
		02608	GATGCTATACTTATGcatttacaggatCAAATCTGATTCACCTTAT	use with 01720
mutate -303 bp C --> A		02609	ATAAACTGGATATCAGATTGatccgttaatgAATAAGTTATGCATCTC	use with 01720
		02610	GATGCTATACTTATGcatttacaggatCAAATCTGATTCACCTTAT	use with 01720
mutate -302 bp C --> A		02611	ATAAACTGGATATCAGATTGatccgttaatgAATAAGTTATGCATCTC	use with 01720
		02612	GATGCTATACTTATGcatttacaggatCAAATCTGATTCACCTTAT	use with 01720
mutate -301 bp G --> A		02613	ATAAACTGGATATCAGATTGttccatgtaaatgAATAAGTTATGCATCTC	use with 01720
		02614	GATGCTATACTTATGttccatgtaaatgAATAAGTTATGCATCTC	use with 01720
mutate -300 bp T --> A		02615	AAAGTGGATATCAGATTGttccgtaaatgAATAAGTTATGCATCTC	use with 01720
		02616	GAGATGCTATACTTATGttccgtaaatgAATAAGTTATGCATCTC	use with 01720
mutate -299 bp G --> A		02617	AAAGTGGATATCAGATTGttccgtataatgAATAAGTTATGCATCTC	use with 01720
		02618	GAGATGCTATACTTATGttccgtataatgAATAAGTTATGCATCTC	use with 01720
mutate -298 bp T --> A		02619	AAAGTGGATATCAGATTGttccgtaaatgAATAAGTTATGCATCTC	use with 01720
		02620	CGAGATGCTATACTTATGttccgtaaatgAATAAGTTATGCATCTC	use with 01720
mutate -297 bp A --> C		02621	AAAGTGGATATCAGATTGttccgtaaatgAATAAGTTATGCATCTC	use with 01720
		02622	CGAGATGCTATACTTATGttccgtaaatgAATAAGTTATGCATCTC	use with 01720
mutate -296 bp A --> C		02623	AAAGTGGATATCAGATTGttccgtaaatgAATAAGTTATGCATCTC	use with 01720
		02624	CGAGATGCTATACTTATGttccgtaaatgAATAAGTTATGCATCTC	use with 01720
mutate -295 bp A --> C		02625	AAAGTGGATATCAGATTGttccgtaaatgAATAAGTTATGCATCTC	use with 01720
		02626	CGAGATGCTATACTTATGttccgtaaatgAATAAGTTATGCATCTC	use with 01720
mutate -294 bp T --> A		02627	AAAGTGGATATCAGATTGttccgtaaatgAATAAGTTATGCATCTC	use with 01720
		02628	CGAGATGCTATACTTATGttccgtaaatgAATAAGTTATGCATCTC	use with 01720
mutate -293 bp G --> A		02629	AAAGTGGATATCAGATTGttccgtaaatgAATAAGTTATGCATCTC	use with 01720
		02630	CGAGATGCTATACTTATGttccgtaaatgAATAAGTTATGCATCTC	use with 01720
<i>CgTH10 p-YFP-pRS313</i>	-1000 bp	01588	GCTGGCCGGCCGCTCTAGACTAGTGATCTACTCATCTTAAACCA	use with 1587
	-900 bp	01589	GGTGGCCGGCCGCTCTAGACTAGTGATCTCGAAGATCAGACAAAT	use with 1587
	-800 bp	01590	GGTGGCCGGCCGCTCTAGACTAGTGATCTCGAAGATCAGACAAAT	use with 1587
	-700 bp	01591	GGTGGCCGGCCGCTCTAGACTAGTGATCTCGAAGATCAGACAAAT	use with 1587
	-600 bp	01592	GGTGGCCGGCCGCTCTAGACTAGTGATCTCGAAGATCAGACAAAT	use with 1587
	-580 bp	02316	ACCGCGTggccggccgcgtctagaactatggatccGGAGTTCCCTCTA	use with 1587
	-575 bp	02404	accgcgtggccggccgcgtctagaactatggatccGGAGTTCCCTCTAAGTGA	use with 1587
	-570 bp	02405	accgcgtggccggccgcgtctagaactatggatccGGAGTTCCCTCTAAGTGA	use with 1587
	-565 bp	02406	accgcgtggccggccgcgtctagaactatggatccGGAGTTCCCTCTAAGTGA	use with 1587
	-564 bp	02502	ccgcgggtggccggccgcgtctagaactatggatccGGAGTTCCCTCTAAGTGA	use with 1587
	-563 bp	02503	ccgcgggtggccggccgcgtctagaactatggatccGGAGTTCCCTCTAAGTGA	use with 1587
	-562 bp	02504	ccgcgggtggccggccgcgtctagaactatggatccGGAGTTCCCTCTAAGTGA	use with 1587
	-561 bp	02505	ccgcgggtggccggccgcgtctagaactatggatccGGAGTTCCCTCTAAGTGA	use with 1587
	-560 bp	02317	ACCGCGTggccggccgcgtctagaactatggatccGGAGTTCCCTCTAAGTGA	use with 1587
	-540 bp	02318	ACCGCGTggccggccgcgtctagaactatggatccGGAGTTCCCTCTAAGTGA	use with 1587
	-520 bp	02319	ACCGCGTggccggccgcgtctagaactatggatccGGAGTTCCCTCTAAGTGA	use with 1587
	-500 bp	01593	GGTGGCCGGCCGCTCTAGACTAGTGATCTCGATGGCCGAGTTGGTAA	use with 1587
	-400 bp	01594	GGTGGCCGGCCGCTCTAGACTAGTGATCTCGATGGCCGAGTTAAAT	use with 1587
	-300 bp	01595	GGTGGCCGGCCGCTCTAGACTAGTGATCTCGATGGCCGAGCAT	use with 1587
	-200 bp	01596	GGTGGCCGGCCGCTCTAGACTAGTGATCTCGATGGCCGAGTTAAATTT	use with 1587
	-100 bp	01597	GGTGGCCGGCCGCTCTAGACTAGTGATCTCGATGGCCGAGTTAACTT	use with 1587
at ATG with homology to YFP (3')		01587	ACCATGATAATTCTCACCTTCTAGACATGTTGACATGGCTGTACTGG	
<i>THI UAS::PacI</i>		02549	CGTGAAGGGATGATGGGGAGGTTtttaatgATGGTATTTGGATCTGGATAAG	use with 1587
		02550	CTTATCCAGCATTTTAAACCGATTTtaatgAACTCCGCCATTCTACCTTACCG	use with 1588
<i>THI UAS::PMU3 UAS</i>		02670	TTGATGGCCAGGTGacgtacacgAATGCTTTGGAACTGCTGAA	use with 1587
		02671	ATTTCAAAGAACCTTGTtgcgtacacgAATCTGGCCATTCTACCT	use with 1588
<i>CgTH10 p + ScTH12 site (forward)</i>		02707	TGATGTGGCggaaaaccccttaatgAGGTTttccctctaagtGAATGCTTTGGTAA	use with 1587
<i>CgTH10 p + ScTH12 site (reverse)</i>		02708	AAGCATTcaacttagagggaaACCTCcttaagggtttccGGCCATTCTTCACGAG	use with 1588
<i>CgTH10 p + ScTH12 site (forward) + THI UASΔ</i>		02709	TGATGTGGCggaaaaccccttaatgAGGTTttccctctaagtGAATGCTTTGGTAA	use with 1587
		02710	AAGCATTcaacttagagggaaACCTCggaaaaccccttagAGGCGCTTACACGAG	use with 1588
		02711	CTCTGAAAAGGATGATGGCGggaaaaccccttagAGGCGTAACTGCTTTGGATCTGG	use with 1587
		02712	CCAAAAGGATTACCTCtttaagggtttccGGCCATTCTTCACGAGTCACCCAC	use with 1588
<i>CgTH10 p-YFP-pRS313</i>	-1000 bp	01784	ggtagccggccgcgtctagaactatggatccGGATATCCATTCACT	use with 01786
	-500 bp	01785	ggtagccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01786
	-400 bp	01846a	ggtagccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01786
	-300 bp	01847a	ggtagccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01786
	-295 bp	02499	ccgcgggtggccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01786
	-290 bp	02500	ccgcgggtggccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01786
	-285 bp	02501	ccgcgggtggccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01786
	-280 bp	02441	ccgcgggtggccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01786
	-260 bp	02442	ccgcgggtggccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01786
	-240 bp	02443	ccgcgggtggccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01786
	-220 bp	02444	ccgcgggtggccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01786
	-200 bp	01848a	ggtagccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01786
	-180 bp	02445	ccgcgggtggccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01786
	-100 bp	01849a	ggtagccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01786
at ATG with homology to YFP (3')		01786	accatgtataatttttaccccttagacatTTTGAAGTTCTCCGGCT	
<i>PMU3 UASΔ</i>		02603a	CGATCAACTGAACGAGTCCTGGAAACCTCTGTCATTGAGCTTTTG	
		02604	ATGACGAGGTTTCGAGCAGCAGTCCTGTCAGTGTGTTG	use with 01786
				use with 01784
<i>CgTH10 p-YFP-pRS313</i>	-950 bp	01249	ggtagccggccgcgtctagaactatggatccGGATATCCATTCACT	use with 01250
	-750 bp	01408	ggtagccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01250
	-650 bp	01409	ggtagccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01250
	-550 bp	01410	ggtagccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01250
	-450 bp	01411	ggtagccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01250
	-430 bp	01443	ggtagccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01250
	-410 bp	01444	ggtagccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01250
	-390 bp	01445	ggtagccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01250
	-370 bp	01446	ggtagccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01250
	-350 bp	01412	ggtagccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01250
	-250 bp	01413	ggtagccggccgcgtctagaactatggatccGGATATCCATTTCCTTT	use with 01250
at ATG with homology to YFP (3')		01250	accatgtataatttttaccccttagacatTTTGAAGTTCTCCGGCT	

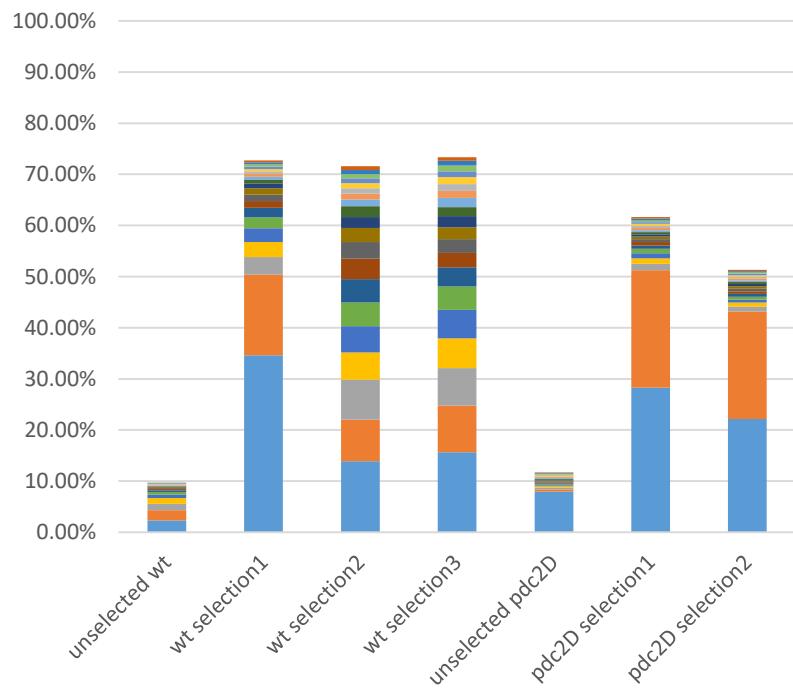


Figure S1



## Figure S2

Frequency of top 20 most abundant sequences in each sample



## Table S2

Raw data from Next Generation sequencing runs sorted by abundance of sequence							
sample 1 unselected wt	sample 2 selection1 wt	sample 3 selection2 wt	sample 4 selection3 wt	sample 5 unselected pdc2D	sample 6 selection1 pdc2D	sample 7 selection2 pdc2D	
CATTACCGG	2.3%	ATCGTATA	34.6%	CAGTTACCCGG	13.9%	CACGTACCGG	15.7%
ATCGTATTTA	2.0%	TTCGGTAA	15.8%	TTCGGTAA	8.1%	ATCGTATTTA	9.1%
CGTAGTCATG	1.3%	TCACGGCAT	3.4%	ATCGTATTTA	7.8%	TTAACCTTGAC	7.3%
CACACTTACG	1.1%	GTCGTTAGTGA	2.9%	CCTTCCTCGTG	5.3%	TTTCGTTAA	5.8%
CATTCGGAGC	0.7%	ATCGTATTATG	2.7%	TTAACCTTGAC	5.1%	CAACTTACG	5.6%
CGTACCATGCG	0.5%	ATCGTATTCAA	2.1%	CACACTTACG	4.6%	CGTCTCCGTG	4.6%
ATCGTATTATG	0.3%	CGTACATAAAG	1.9%	CGTAGTTCATG	4.6%	CGTAGTTATG	3.7%
CACTGTGACG	0.2%	ATCGTATGCAG	1.3%	CATTCGGACG	4.0%	CATTCGGACG	2.8%
CGCGTACTACA	0.2%	ATCGTTATCAA	1.3%	CTTCGTTAGAA	3.3%	CACTGTGACG	2.7%
ATCGTATTCAA	0.2%	TTCGTTATAAG	1.2%	CACTGTGACG	2.7%	ATCGTATCAA	2.4%
CAAAAAGTATT	0.1%	CCCGTACTACA	0.9%	ATCGTATTATG	2.2%	CGTACCATGCG	2.2%
CAACTAGTGCG	0.1%	AAGATCACGTT	0.7%	CGTACCATGCG	2.1%	CTTCGTTAGAA	1.8%
CGTTAAGTATG	0.1%	ATTCGTTATGA	0.6%	CGTGTAAAGG	1.3%	CGTCATGGTG	1.8%
GCAACTGACTT	0.1%	ATCGTATTAA	0.6%	CGTACATAAAG	1.1%	ATCGTATATG	1.4%
TACATGAAAT	0.1%	ATCGTATTAA	0.5%	CGTCATGGGTG	1.0%	CGTGTAAAGG	1.3%
ATCTTACCATC	0.1%	ATCTTATTTA	0.5%	TCACGGGCAT	1.0%	CGTACATAAAG	1.3%
ATCGTTCATAA	0.1%	ATCGTAA	0.5%	CTCTCTGTACG	0.9%	TTTCGTTCCAG	1.2%
CTCAGTTGTC	0.1%	CTCGTAACAA	0.5%	GTGTTAGTGA	0.9%	TTTCGTTCAA	1.1%
TACTCTGTACG	0.1%	TCAAGTGCCT	0.4%	GTGTTACCGG	1.1%	GTTCGACTGAT	0.2%
CATGTGAGTACG	0.1%	GATATCATTCT	0.3%	ATCGTATCAA	0.7%	CAACTTACG	0.3%
6128 out of 8986 unique	483 out of 2117 unique	356 out of 1750 unique	416 out of 2082 unique	4347 out of 8175 unique	764 out of 2398 unique	2677 out of 10382 unique	
Percent unique	68.2%	22.8%	20.3%	20.0%	53.2%	31.9%	25.8%

## Table S3

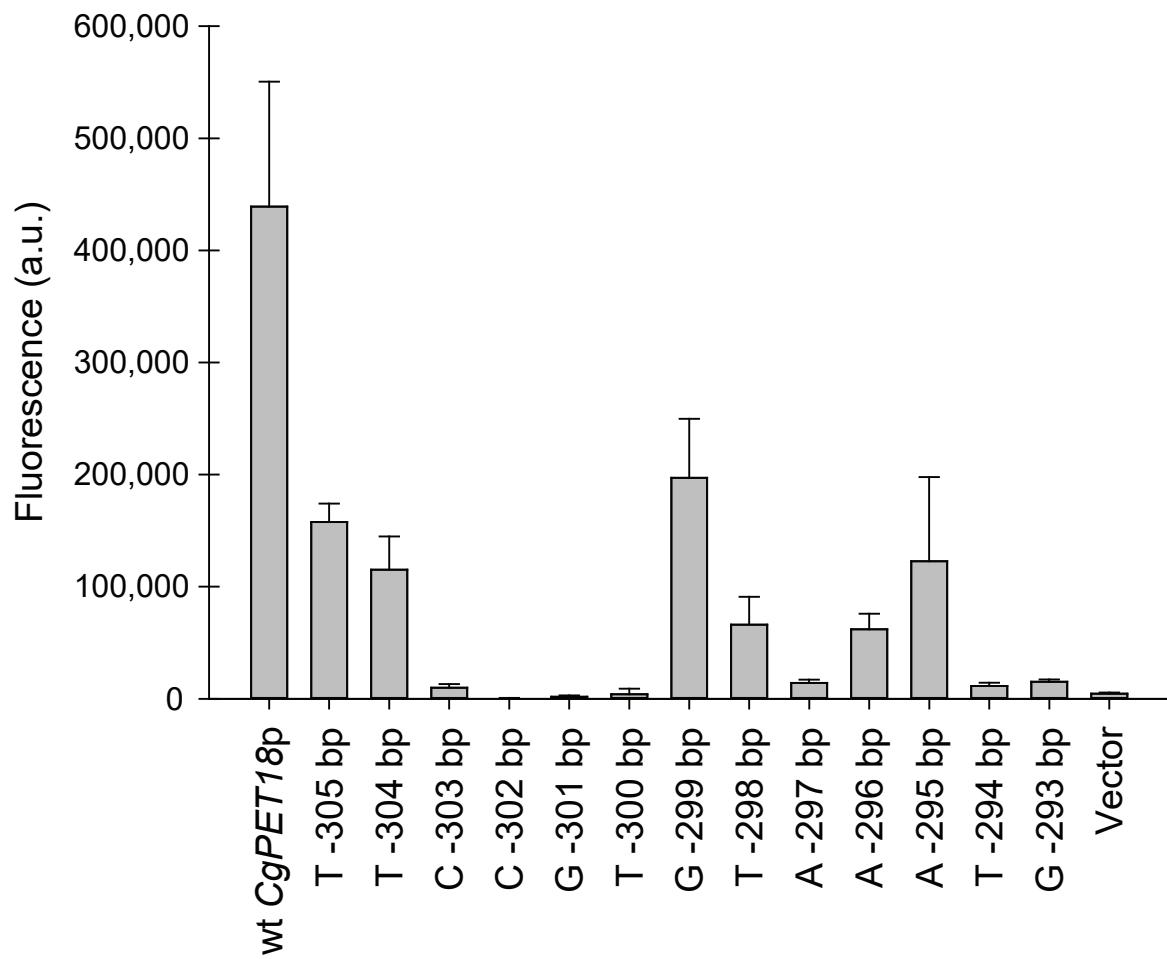
**Sequences that were highly enriched in wild-type by selection in no thiamine conditions**

	Fold enrichment	Final proportion	Fold enrichment in <i>pdc2Δ</i>	times observed in wt selection
A	TTTCGTGTAAA 888x	9.9%	<3x	3
B	TTTACCCCTGAC 561x	6.2%	6.8x	2
C	CGTTCTCCGTG 445x	5.0%	<3x	2
D	TCACGTGGCAT 161x	18.0%	<3x	3
E	CGTACATAAAG 133x	1.5%	<3x	3
F	CGTCATGGGTG 126x	1.4%	<3x	2
G	CGTGTAAAGGG 119x	1.3%	<3x	2
H	TTCGTTATAAG 91x	1.0%	<3x	2

## Figure S3

CgPET18	--TTCCGTGTAAATG-
A	--TTTCGTGTAAA---
G	-----CGTGTAAAGGG
B	TTTACCCCTGAC-----
CgPET18	--TTCCGTG-TAAATG-
C	-----CGTTCTCCGTG
F	-----CGTCATGGGTG
CgTHI20	--TTCACTCTAATT
CgTHI10	--TTCCCTCTAAAGTG
CgTHI4	--TGCCCTTGAAATTG
H	--TTCGTTATAAG-
CgPMU3	(AGA) -GACGTACAACG-
E	(AGA) --CGTACATAAAG-
D	(AGA) TCACGTGGCAT-

Figure S4



## Figure S5

### *ScTHI5*

TGCATTTATCGGTTTGGCTATGCAATGAGTACAGTCCAATTGAACGGCTCATGAACATGAAATATCGAATTATT  
 ACGTAAATAGCCAAAACCGATACTACGTACTCATGTCAGGTTAACCTGCCGAGTAGTTACTTATATGCTTAATAAG  
 -290      -280      -270      -260      -250      -240      -230      -220      -210  
 GCGAATCCCTGTG  
 CAACTAGCAGAAG GCTTG  
 AAATATTACTTATTCAAGATTATGAAAATGGTGCTGTTATAGCAAAATAAGCAACATCAATCTCTGCGACGACAGCAAACAAA  
 ATAGCAATTCTGCTGTTAACCTTACACGACAATATCGTTATTCGTTAGTTAGAGAACGCTGCTGCGTTGTTGTATATT  
 TGGAGCA  
 -190      -180      -170      -160      -150      -140      -130      -120      -110  
 TTATCATCTCATGAAACAATATTCTGTCCATTCTTTACCTTTCAATTAAACACACTTCCAAGCT  
 TAATAGTAGAAGTACTGTTATAAGAACAGGTAAGAAAACAAGAAAATGGAAAAGTTAAAATATT  
 -90      -80      -70      -60      -50      -40      -30      -20      -10  
 ATGTCTACAGACAAG  
 M S T D K  
 TACAGATGTCTGTTG

### *ScTHI20*

GATAGATTAATGATCCCCGATGAAGGTTAGGAAACCTTAAAGCTAGCCTATTAAACCATCAGACAAAGCATACTGA  
 CTATCTAATTACTAGGGCTACTTCCAATCCTTGGGAATCTCGATACCGCTGACCCTTTTTAGGATAATAATT  
 -190      -180      -170      -160      -150      -140      -130      -120      -110  
 GTCCCCCTTCGGG  
 CAGCTGGCAGAAA  
 AAATCCTATTATTAAACCATCAGACAAAGCATACTGA  
 GACTTCGATAAAGGTTGCATTCTTGCTTATTATTTCTCTCAATGATACTAGACAGATTAATAATAATATAAGGAAA  
 AGCTCGCATTTACT  
 CTGAAGCTATTCCAACGTAAGGAACGCAATAATAAAAAGAGAGTTACTATGATCTGCTAATTATT  
 -90      -80      -70      -60      -50      -40      -30      -20      -10  
 ATGACCTATTCTACA  
 M T Y S T  
 TACTGGATAAGATGT

- [Yellow box] = candidate THI UAS
- [Green box] = candidate Thi2 binding site
- [Grey box] = Region required for expression in Figure 7B and 7C
- [Blue box] = site of mutation of THI UAS in Figure 7A

Name	Strand	Start	p-value	Sites
2. CgTHI10pr	-	3	1.00e-7	T CACTTAGAGGGAA AC
5. ScTHIS	+	14	7.52e-6	ACGAATTATT CAACTAGCAGAAG GCTTGAAATA
6. ScTHI20	+	21	9.95e-6	AGAGCTATGC CAGCTGGCAGAAA AAATCCTATT
1. CgPET18pr	-	3	1.06e-5	T CATTTACACGGAA CA
3. CgTHI4pr	-	3	1.12e-5	G CAATTCAAGGGCA TT
4. CgTHI20pr	-	3	2.34e-5	C AAAATTAGAGTGAA AT

Figure S6

