

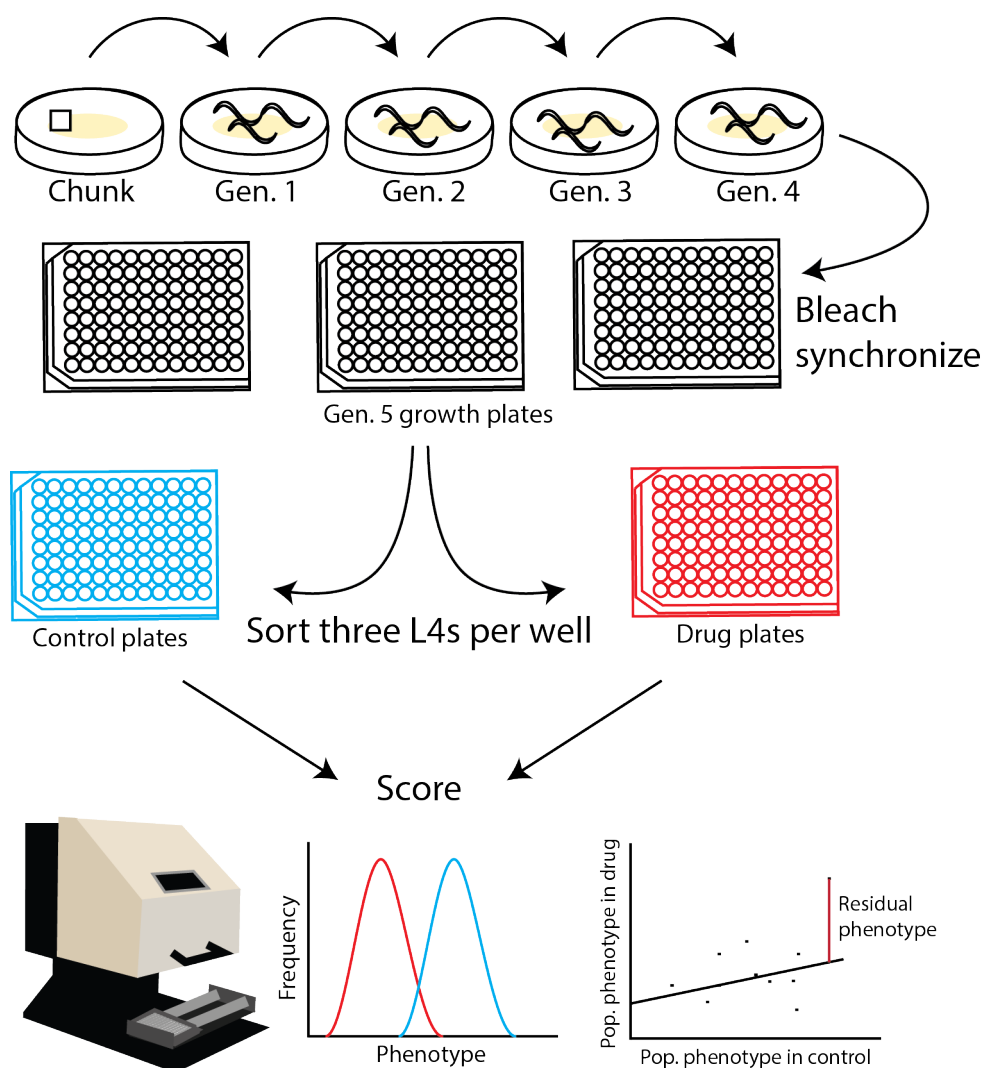


Figure S1

A diagram shows the high-throughput fitness assay protocol. From top to bottom, left to right: Each strain is chunked onto a fresh plate, and animals of the L4 stage are transferred for four generations. Gravid adults from the fourth generation are bleached to synchronize growth of all strains. Embryos are aliquoted to 96-well growth plates containing K medium and 5 mg/mL of bacterial lysate. After 48 hours, three L4 larvae are sorted from each well of the growth plate to the corresponding well of either a control plate (which contains K medium, 10 mg/mL bacterial lysate, and 1% distilled water) or a drug plate (which contains K medium, 10 mg/mL bacterial lysate, 1% distilled water, and bleomycin). Animals are grown in the control and drug plates for four days at 20° with shaking. Then, animals are scored using the COPAS BIOSORT. For each well, all animals are measured for length (TOF) and optical density (EXT). The normal distributions represent hypothetical distributions of all animals in a given well of a control (red) plate or a drug (blue) plate. Summary statistics of these distributions are measured to obtain a population growth estimate for all animals within a well. For each trait measured, the average control phenotype for a given strain is plotted against the drug phenotype for each replicate of that strain. A linear model is fit to those data, and the residual phenotype is calculated and used for further analysis.

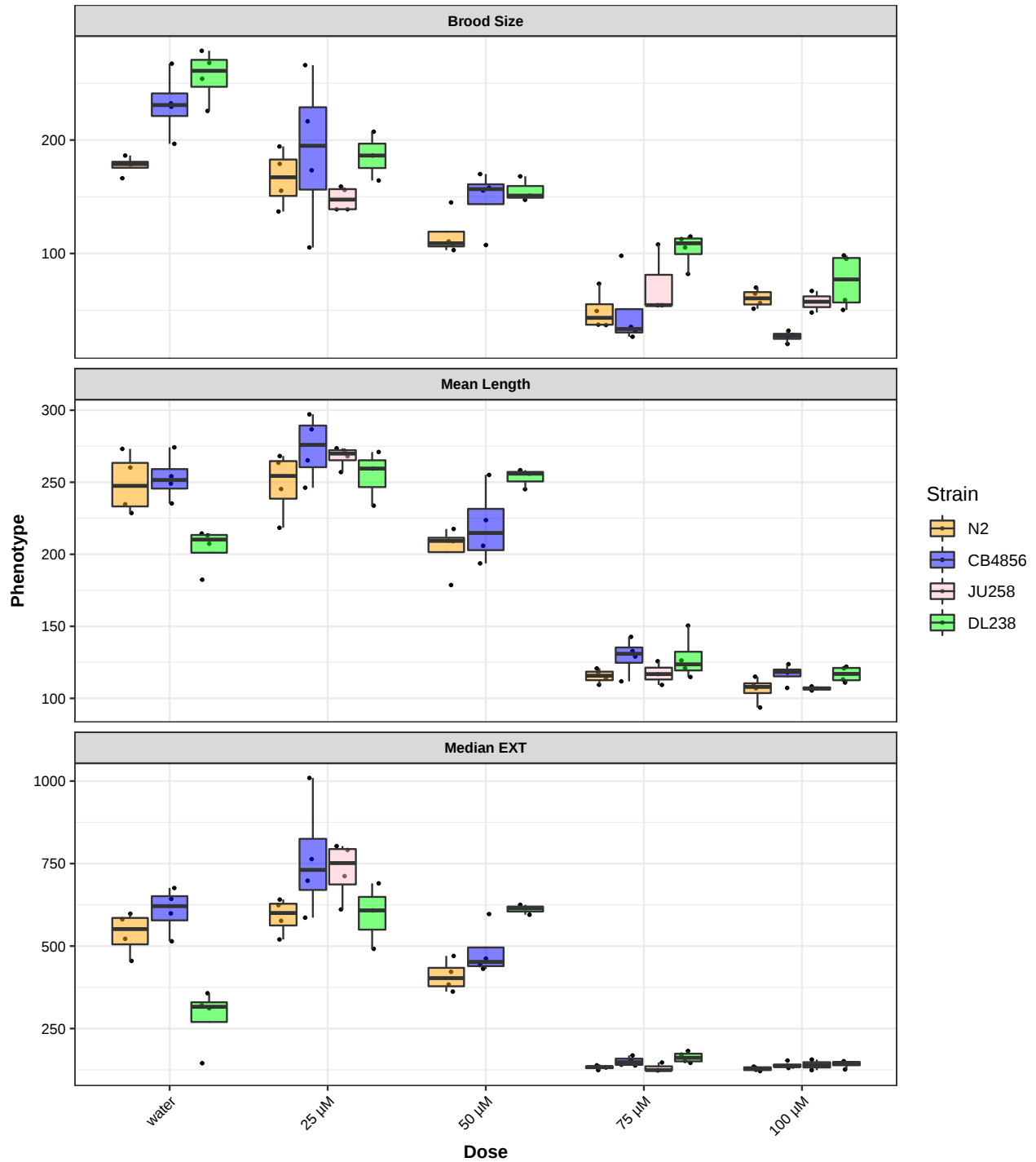


Figure S2

Dose-response phenotypes are shown for three high-throughput fitness traits: brood size (norm.n), mean length (mean.TOF), and median optical density (median.EXT). Phenotypes for each of the four wild isolates are shown as Tukey boxplots, colored by strain (N2 - orange, CB4856 - blue, JU258 - pink, DL238 - green). The x-axis shows the concentration of bleomycin (or water) to which the animals were exposed, and the y-axis shows the pruned phenotype. Each point is a biological replicate. The JU258 strain was pruned from the water and 50 µM bleomycin conditions because the wells were contaminated.

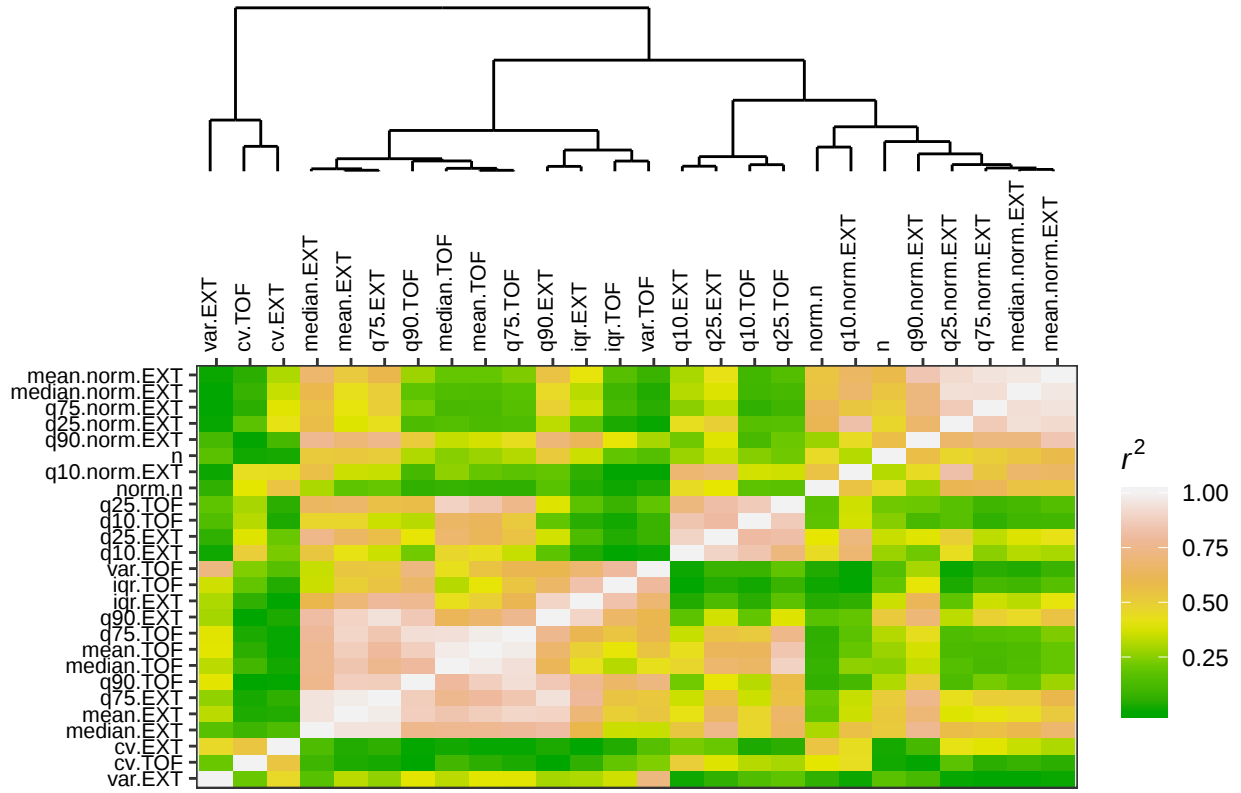


Figure S3

Pairwise correlations of high-throughput assay (HTA) traits are shown. **(Top)** A dendrogram of HTA traits, calculated from pairwise correlations of RIAIL phenotypes for each trait, is shown. **(Bottom)** The x- and y-axes list HTA traits for which the pairwise correlation coefficients (r^2) are shown as colored tiles, ranging from green to yellow to pink to white.

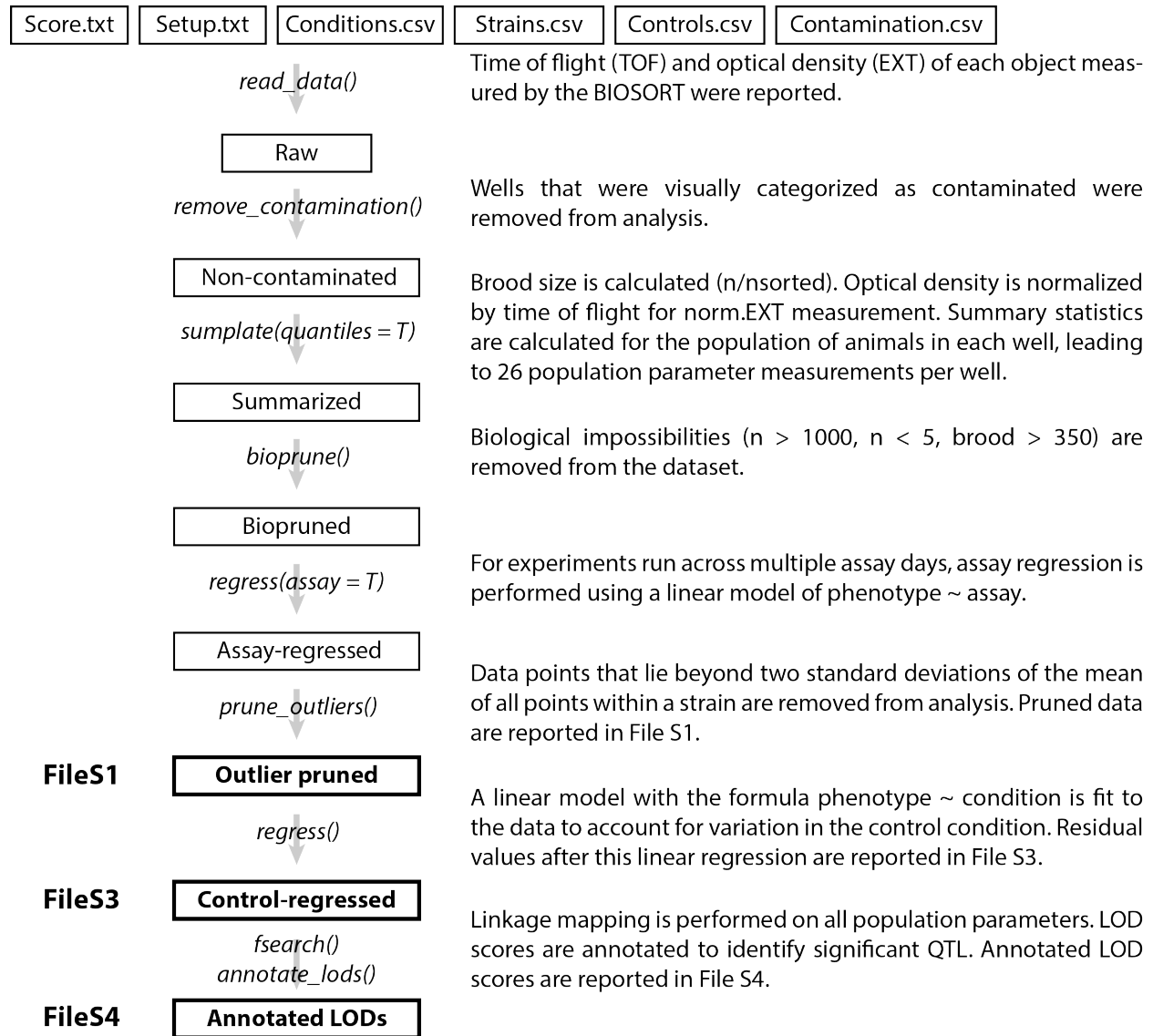
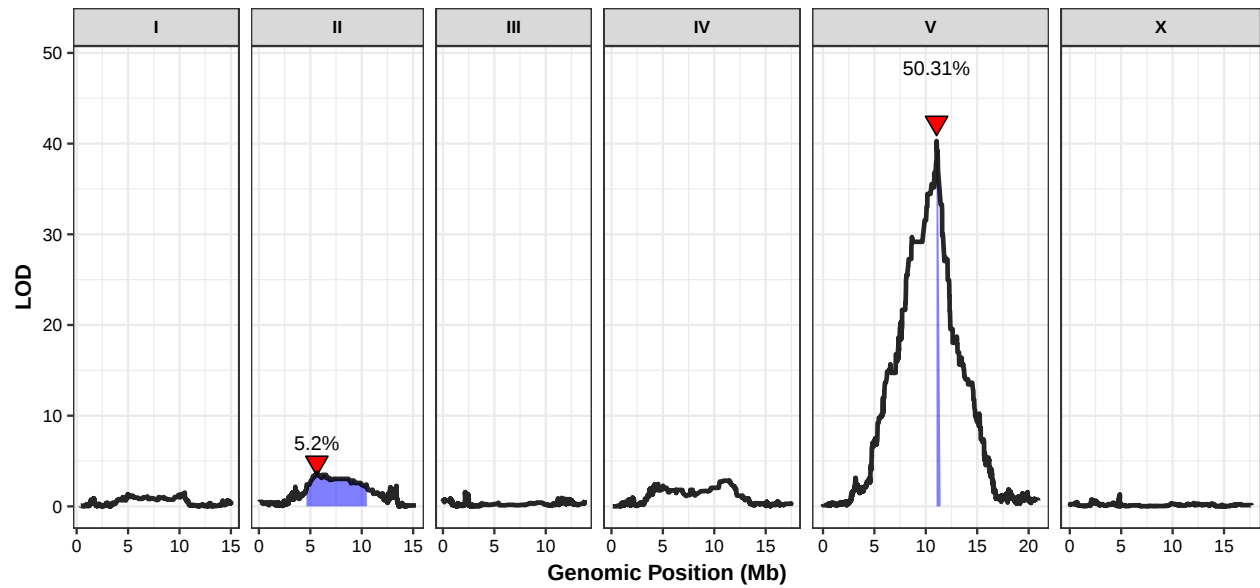


Figure S4

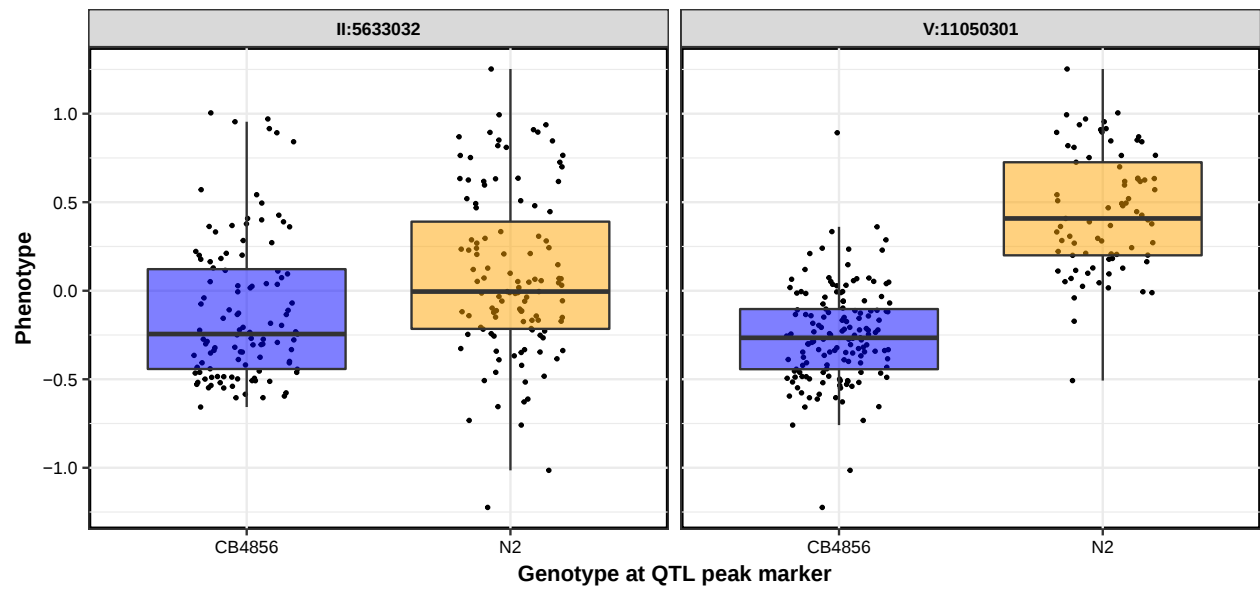
A flow chart from raw data to an annotated LOD score dataframe after linkage mapping is shown. **Center:** Intermediate datasets are listed as file names surrounded by a rectangle, and output files in figshare are in boldface with a thick black border. Arrows and italicized text indicate functions in R that were used during analysis. **Left:** File names of data output to figshare. **Right:** Descriptions of analysis steps performed. The *read_data* function merged the setup and score text files from the BIOSORT with csv files that indicated the conditions, strains, controls, and contamination of all wells measured with the high-throughput fitness assay for a particular experiment. Contamination was removed from the raw data using the *remove_contamination* function. Then, *sumplate* calculated brood size, normalized optical density (EXT) to animal length (TOF) for the norm.EXT measurement, and calculated population parameters (including mean, median, variance, covariance, and quantiles) for TOF, EXT, and norm.EXT measurements. Biological impossibilities were removed by the *bioprune* function. If an experiment was measured across multiple days, the *regress(assay = T)* function was used to account for between-day variation. Outliers were removed from each strain using the *prune_outliers* function. These pruned data were output to **File S1**. The *regress* function was used to calculate residual phenotype measurements for all traits within a strain. Regressed data were output to **File S3**. To perform linkage mapping, the *fsearch* and *annotate_lods* functions were used on all traits in **File S3**.

and mapping results were output to **File S4**.

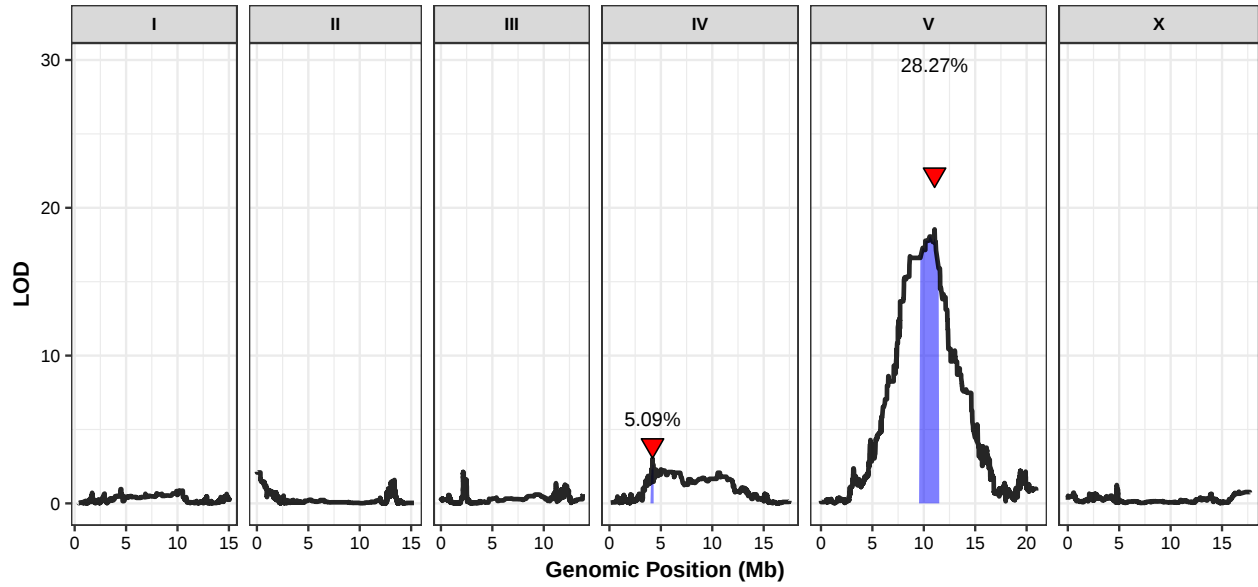
A Bleomycin cv.EXT



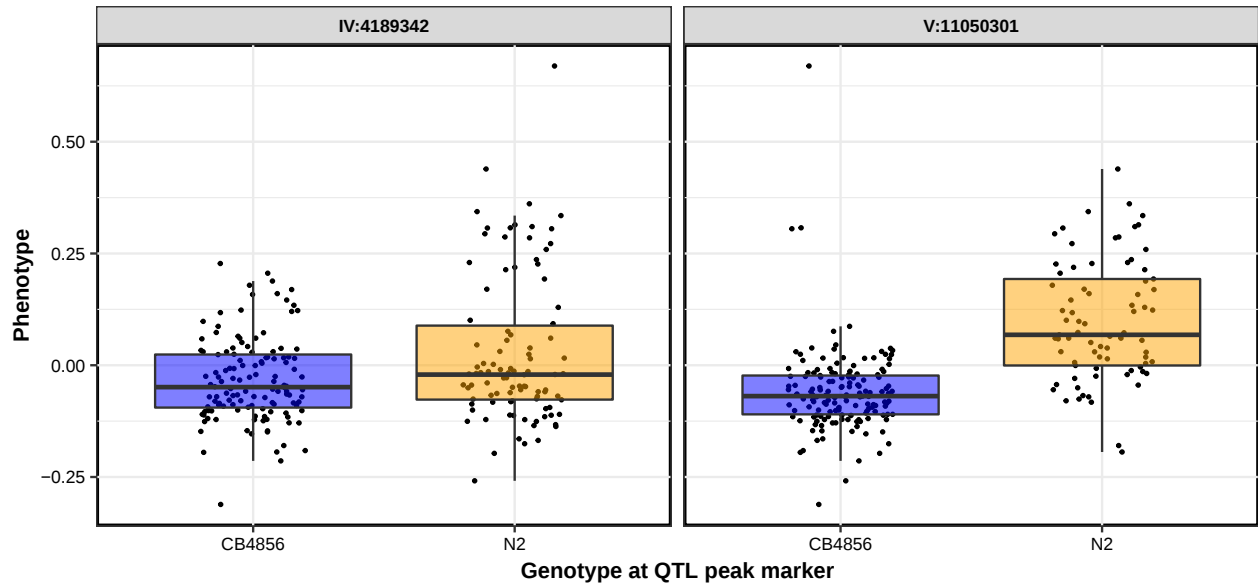
B



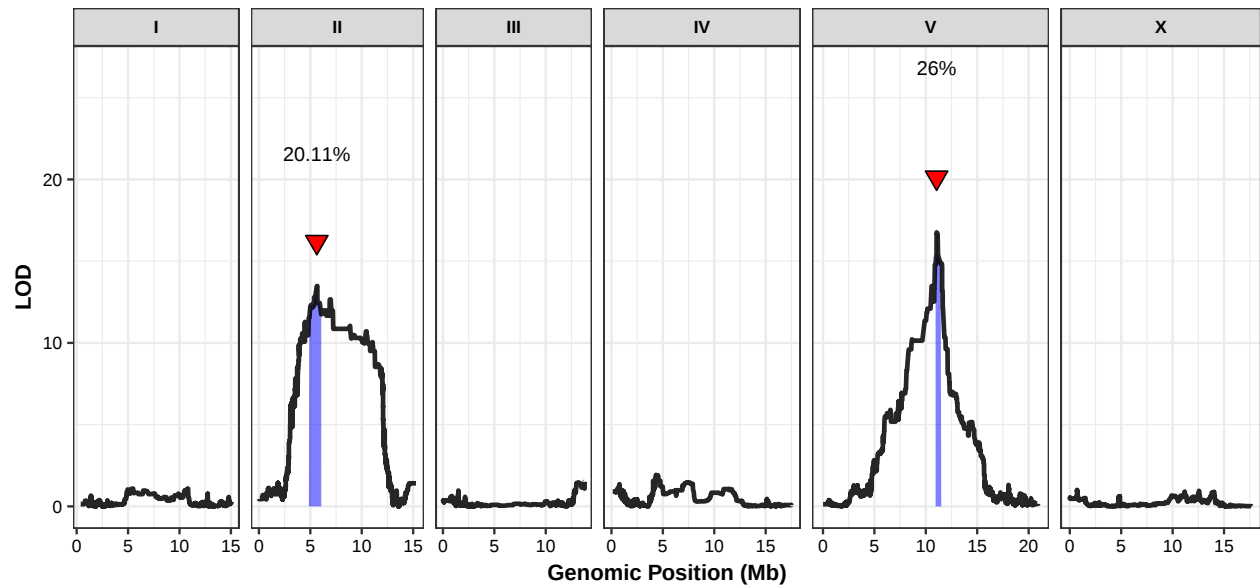
A Bleomycin cv.TOF



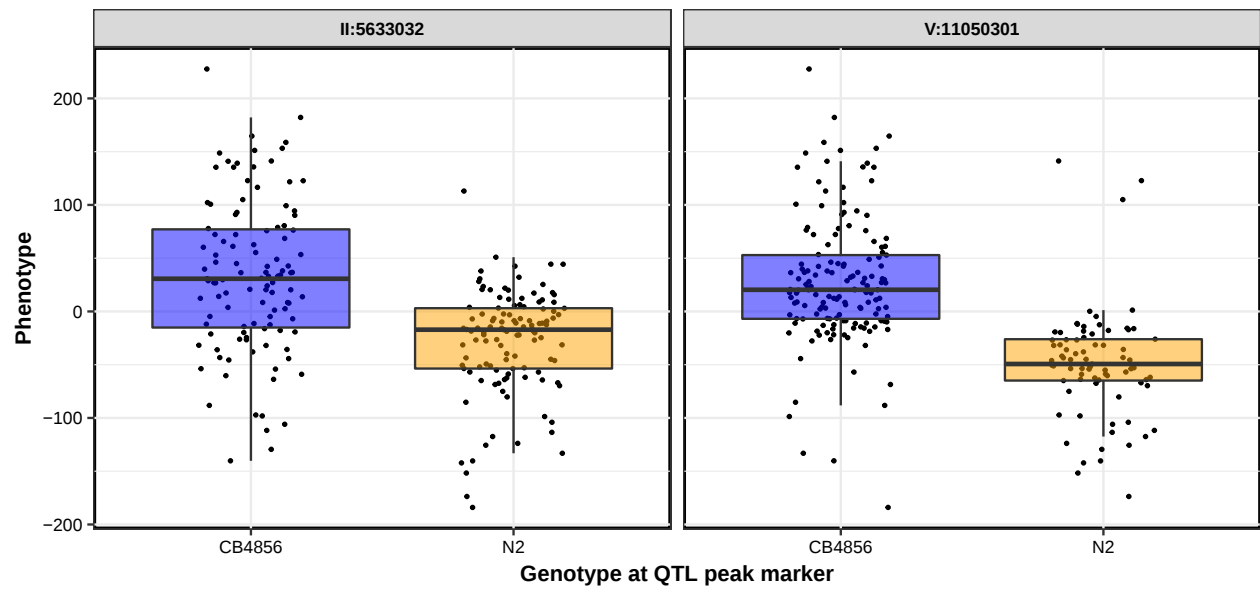
B



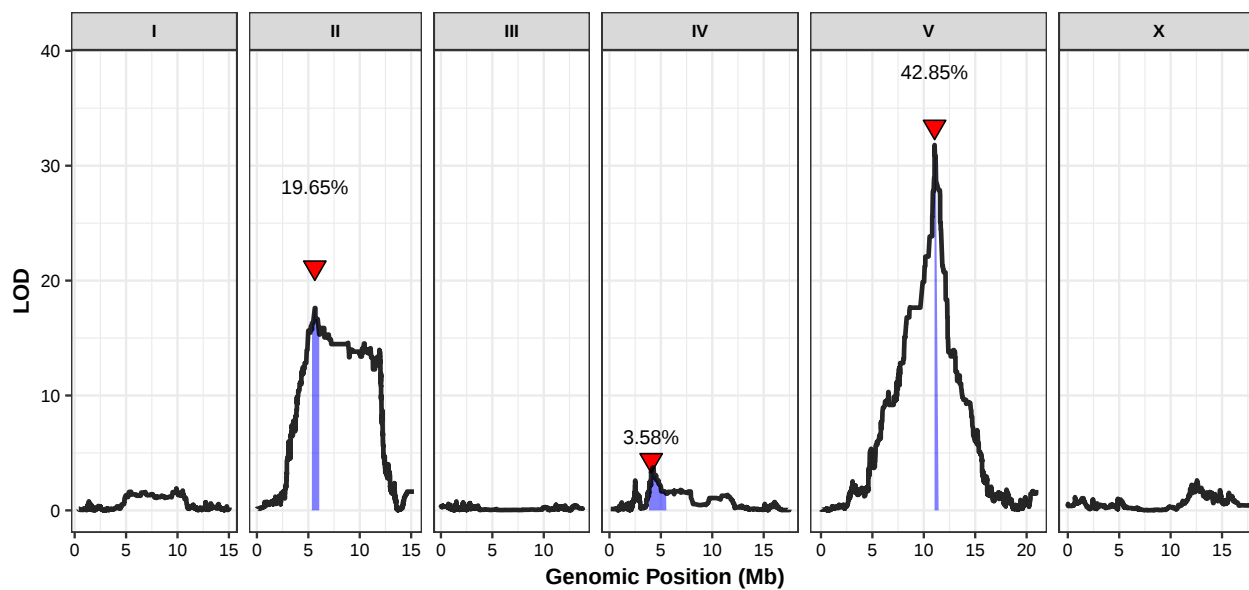
A Bleomycin iqr.EXT



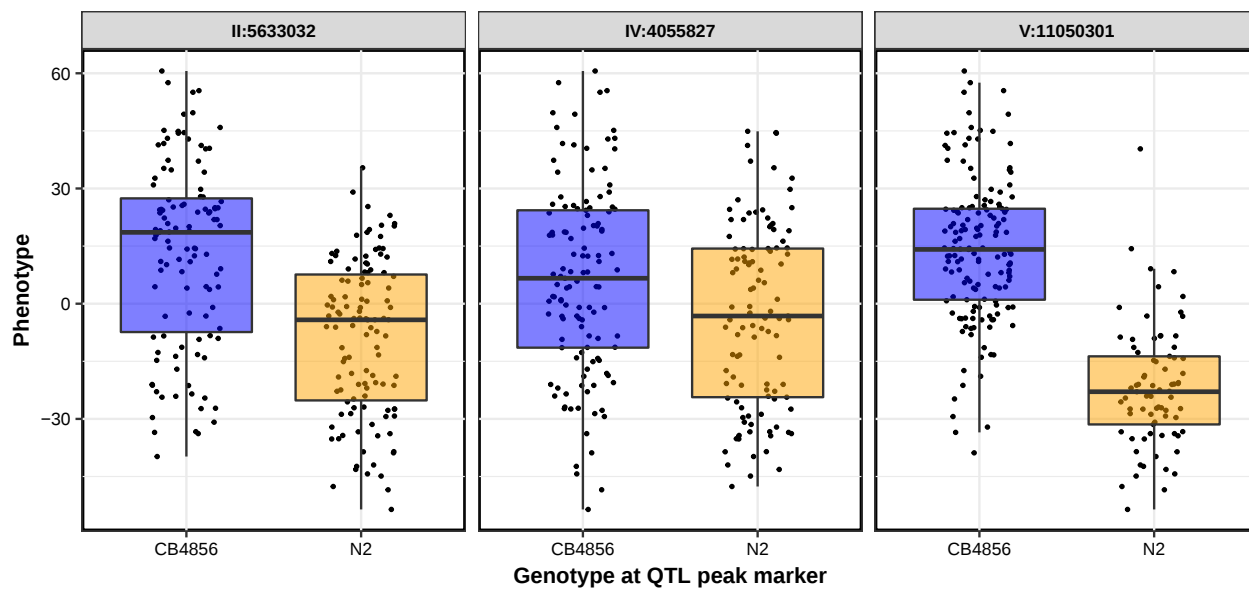
B



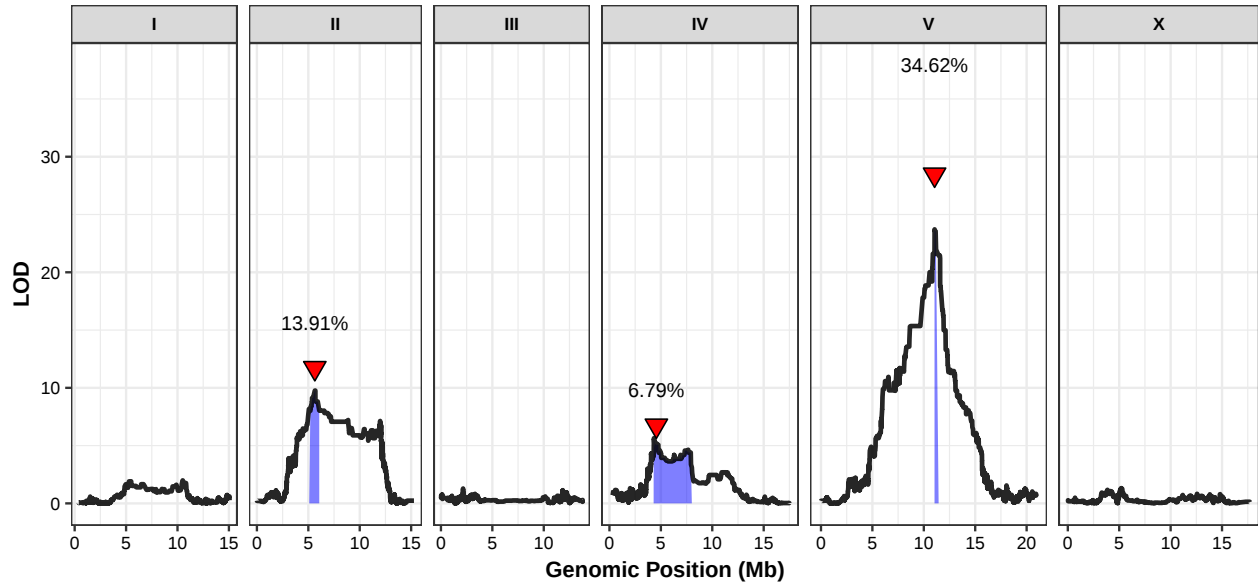
A Bleomycin iqr.TOF



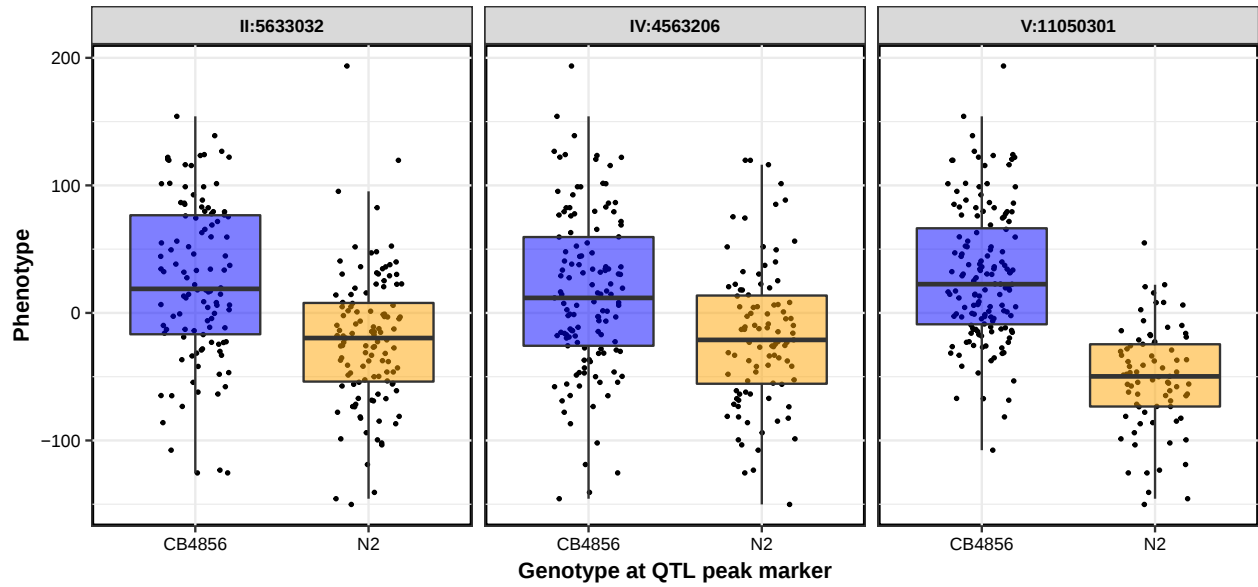
B



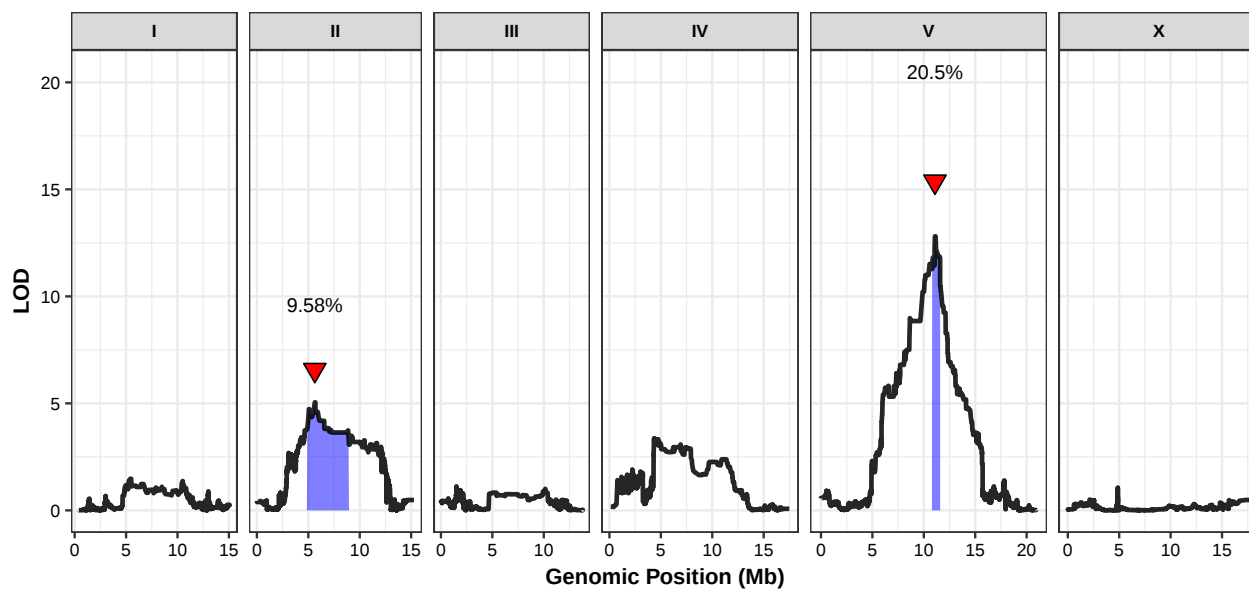
A Bleomycin mean.EXT



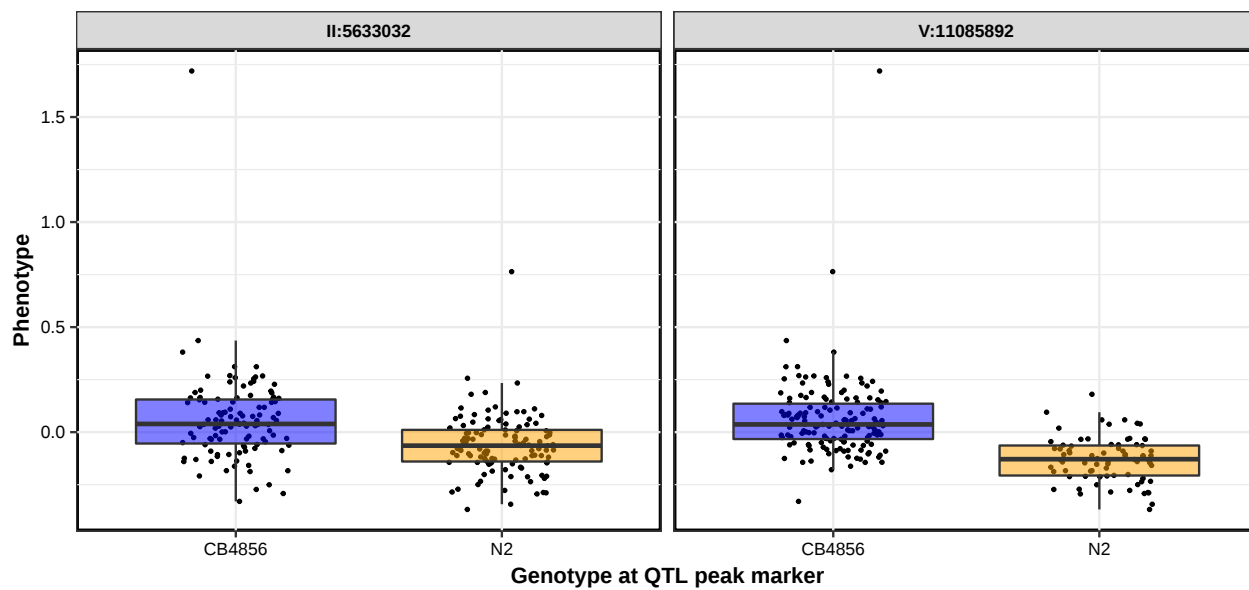
B



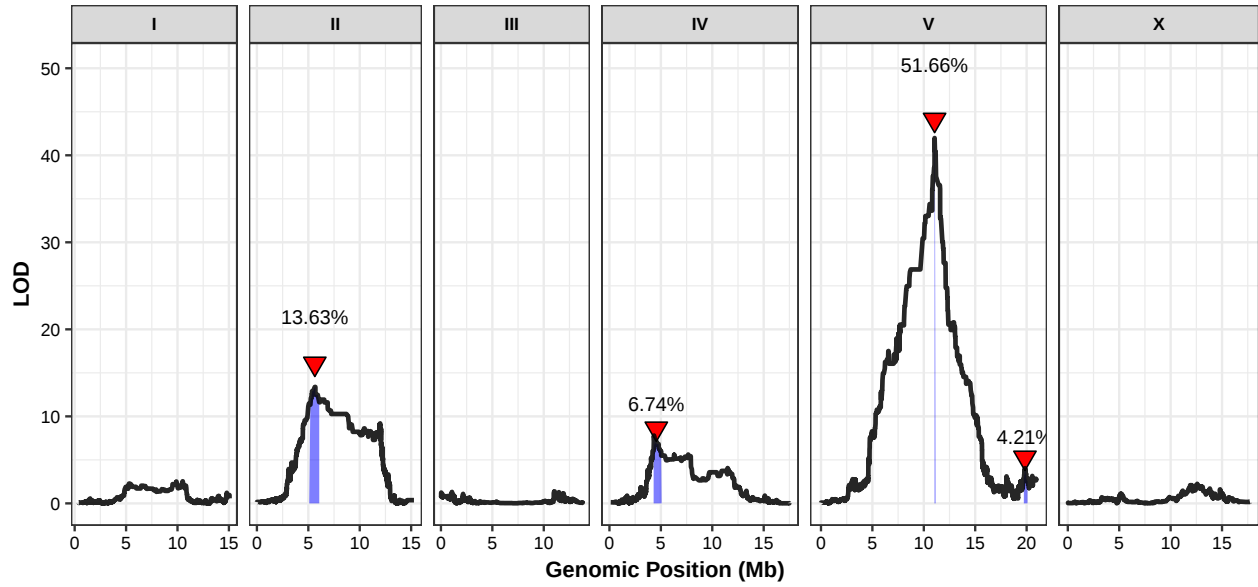
A Bleomycin mean.norm.EXT



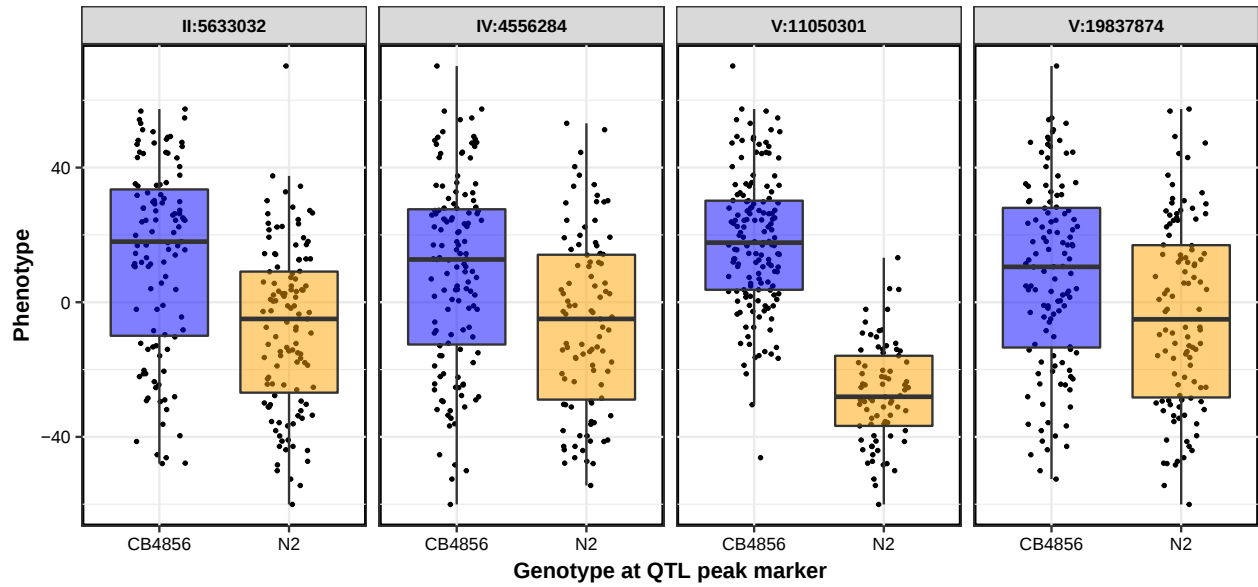
B



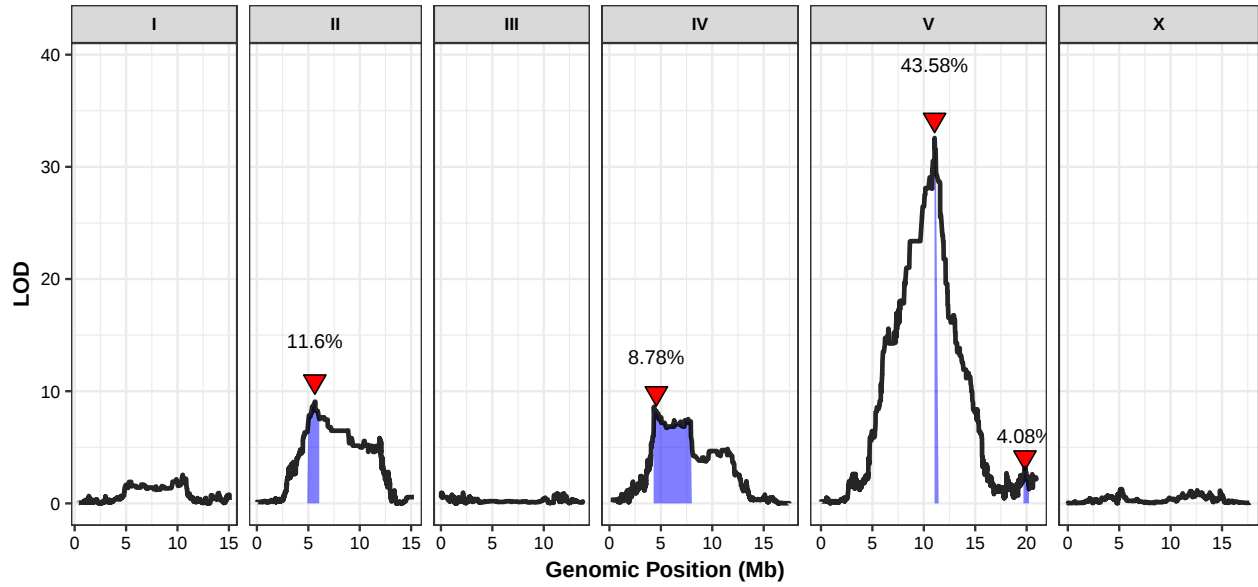
A Bleomycin mean.TOF



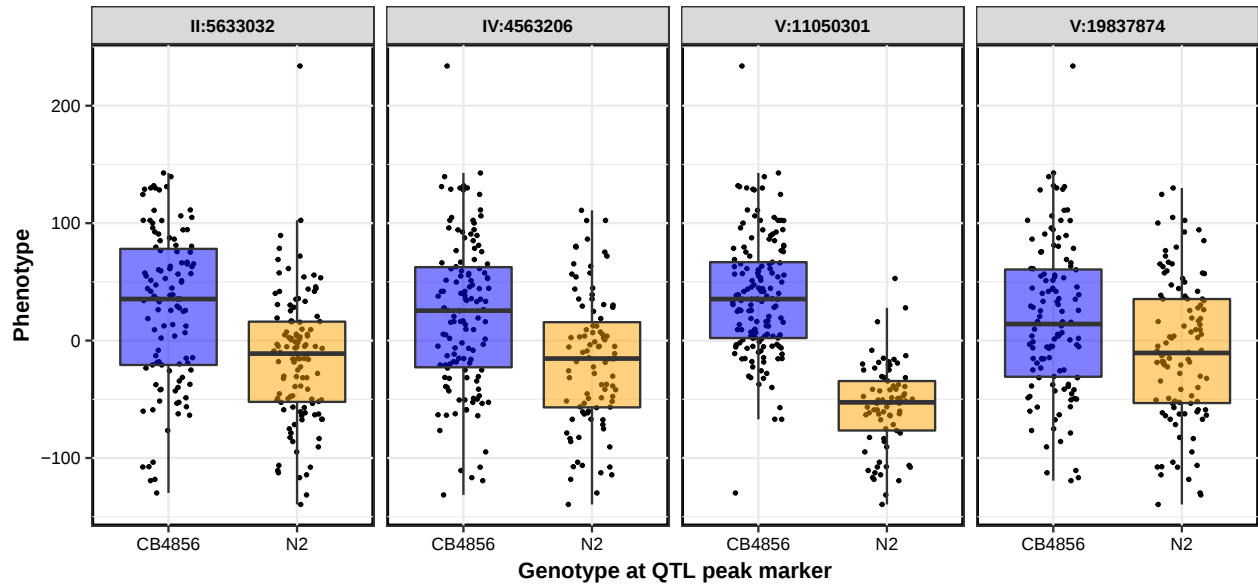
B



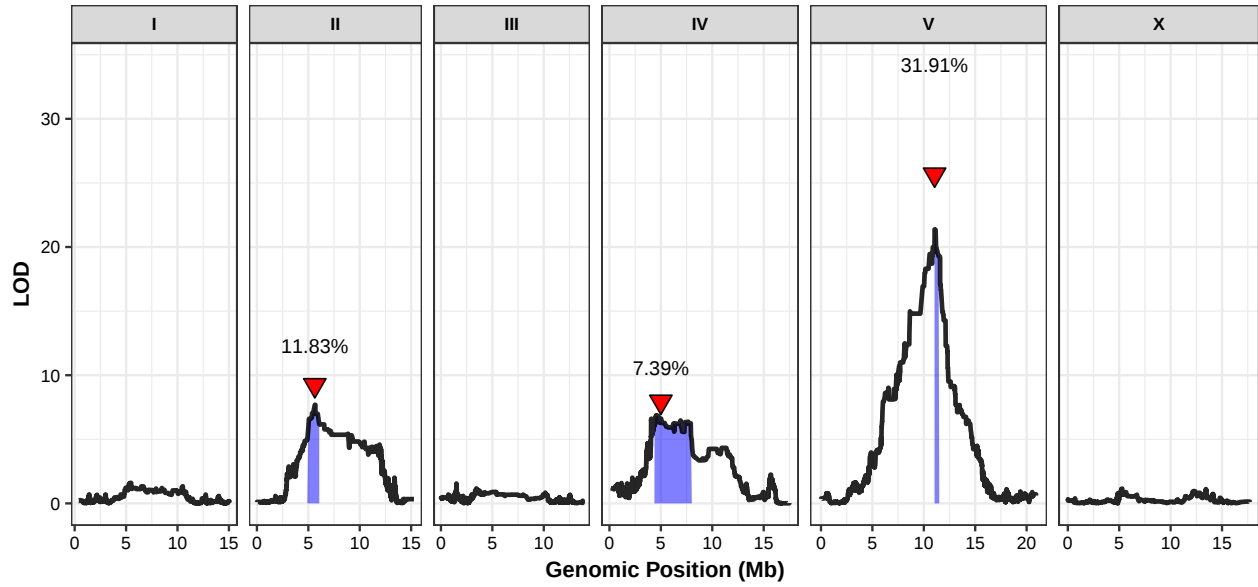
A Bleomycin median.EXT



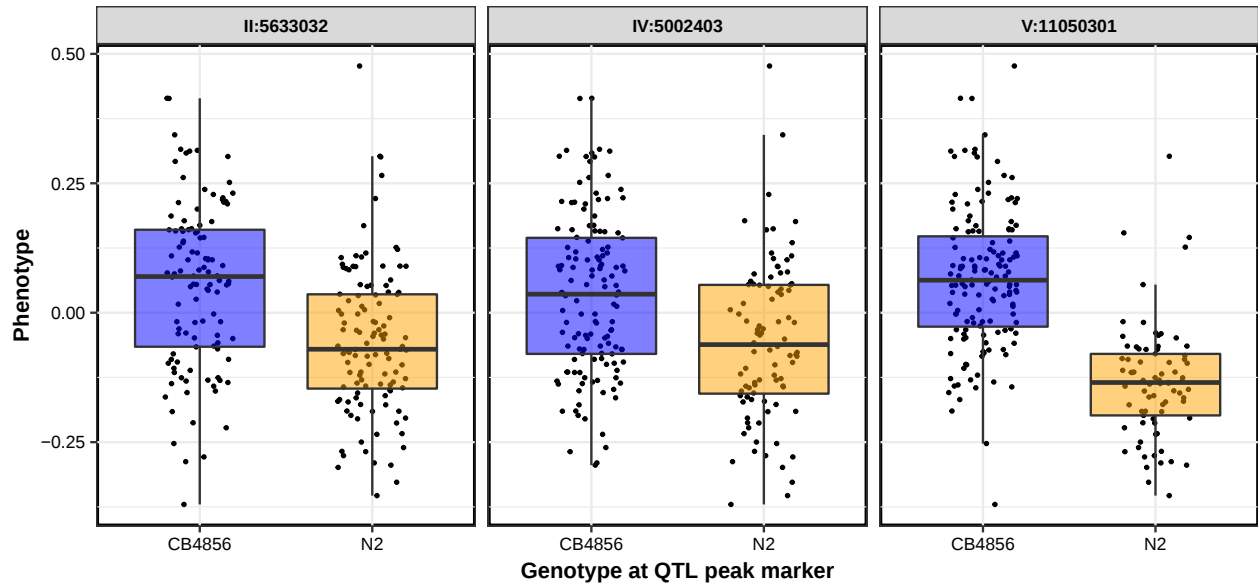
B



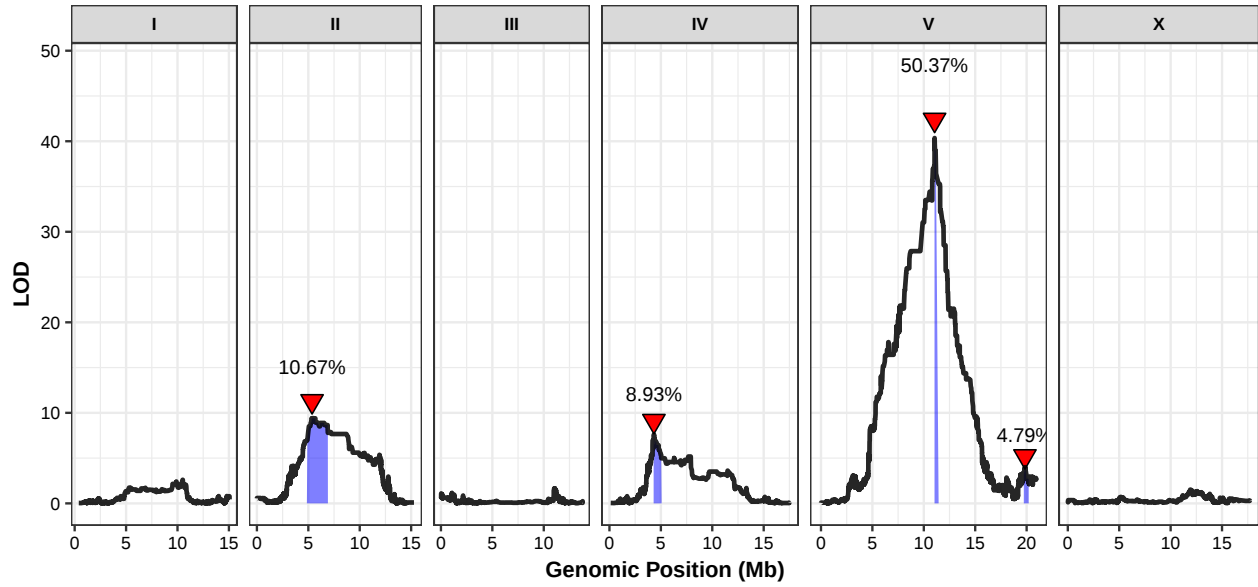
A Bleomycin median.norm.EXT



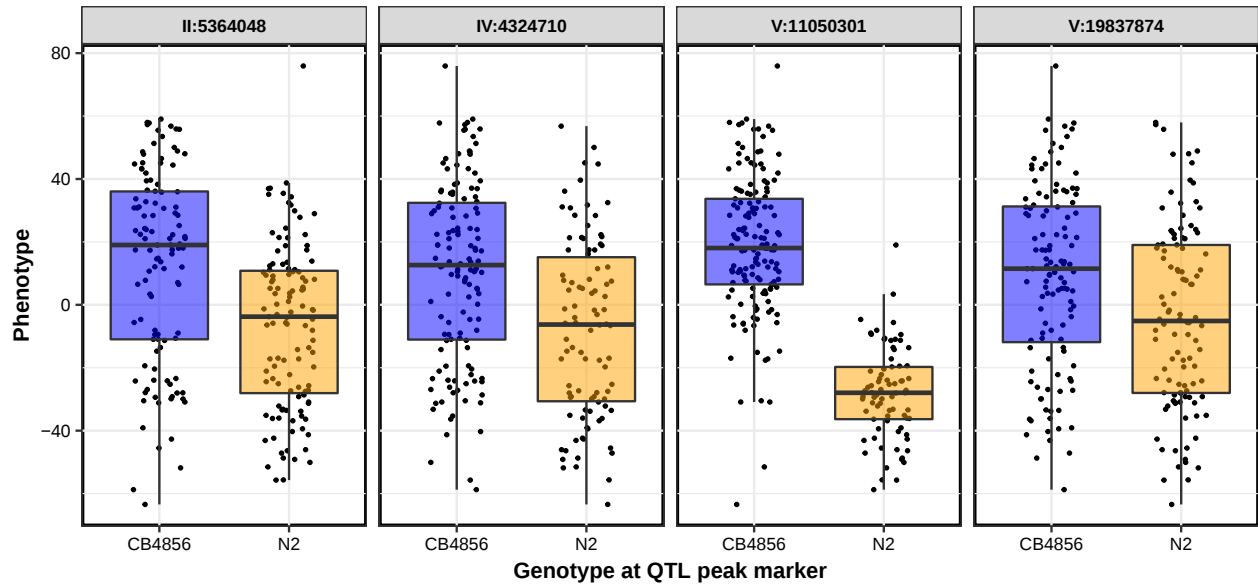
B

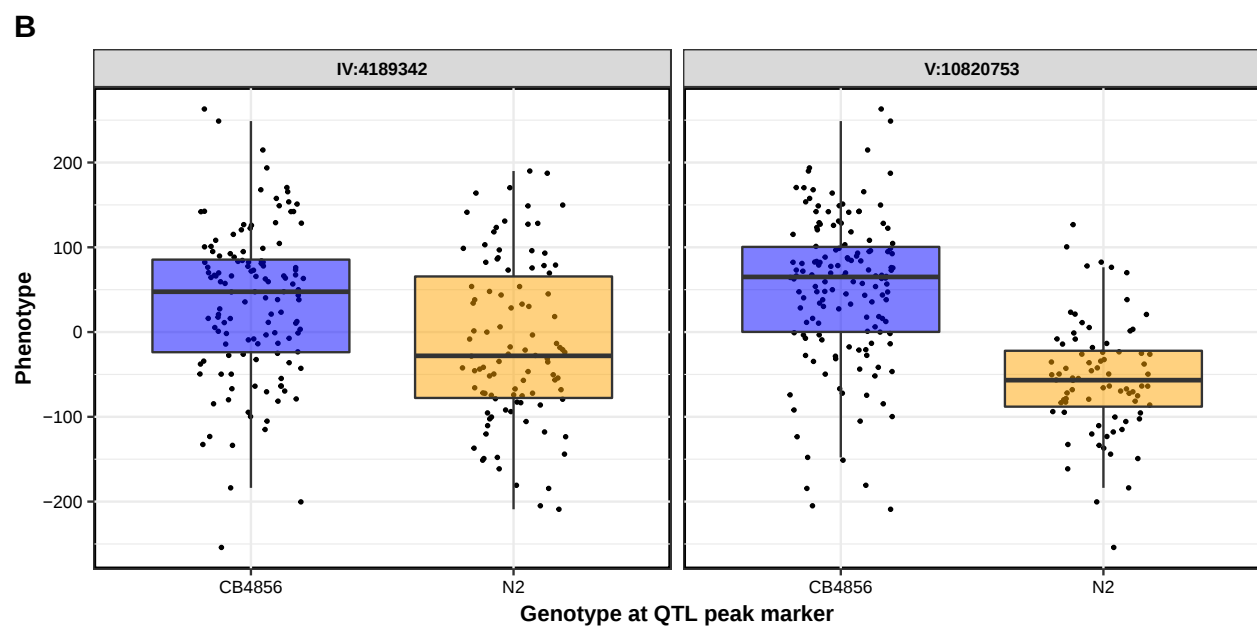
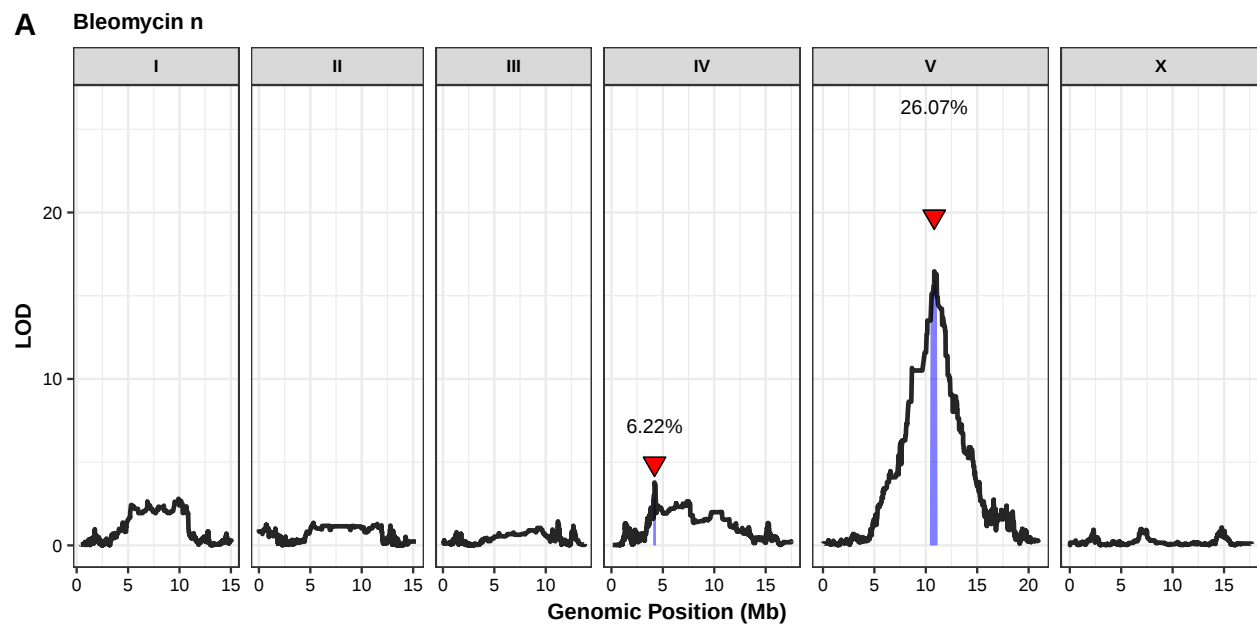


A Bleomycin median.TOF

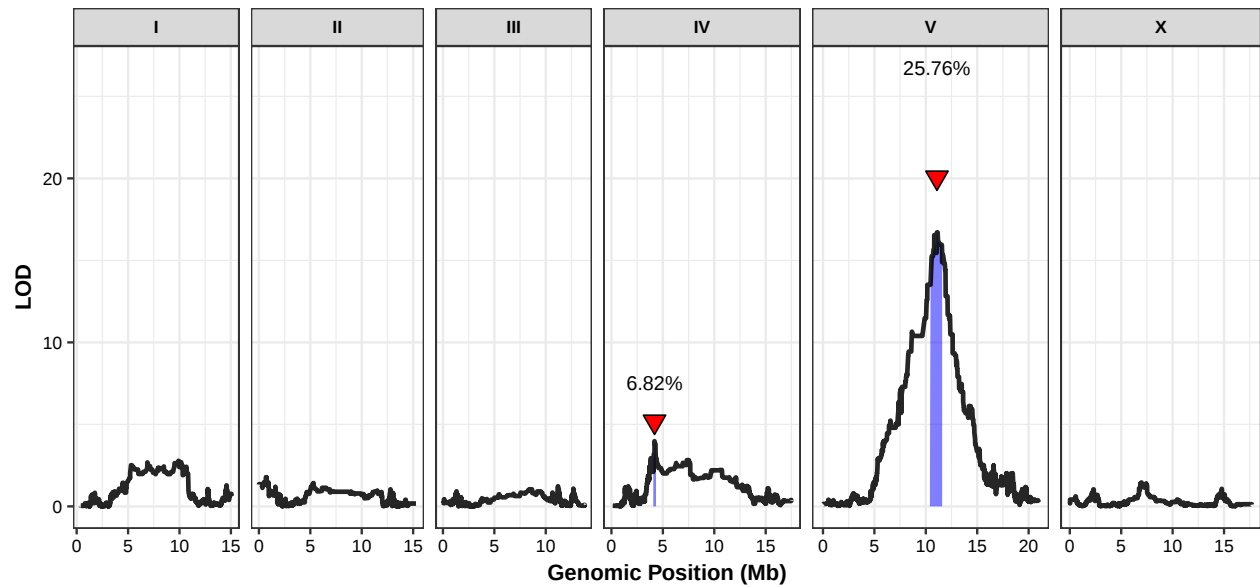


B

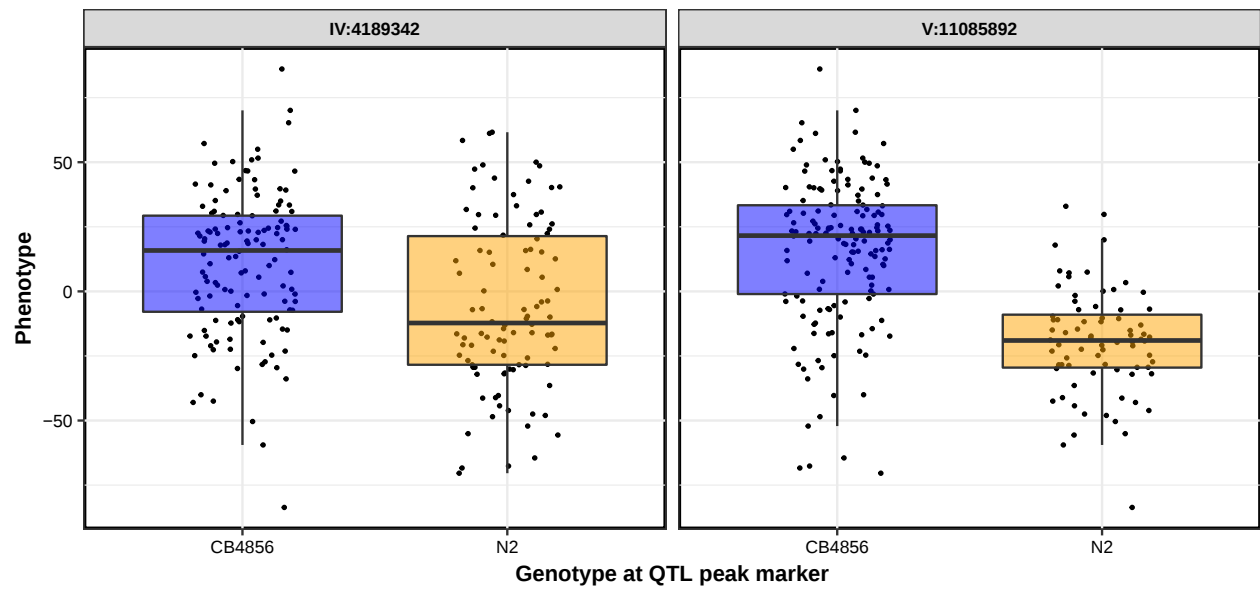




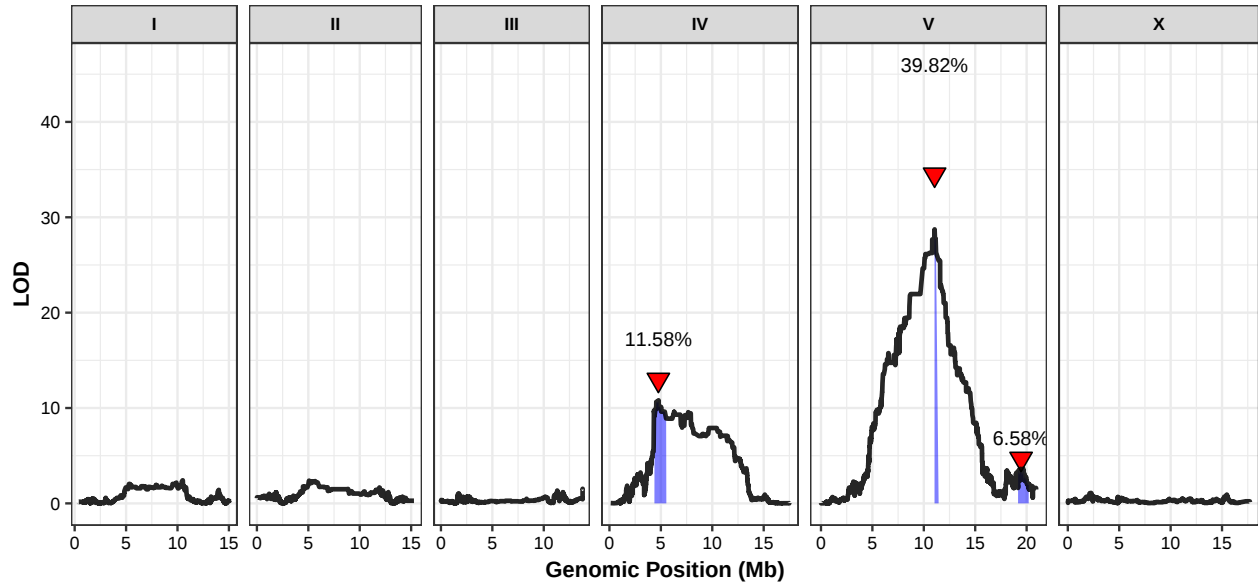
A Bleomycin norm.n



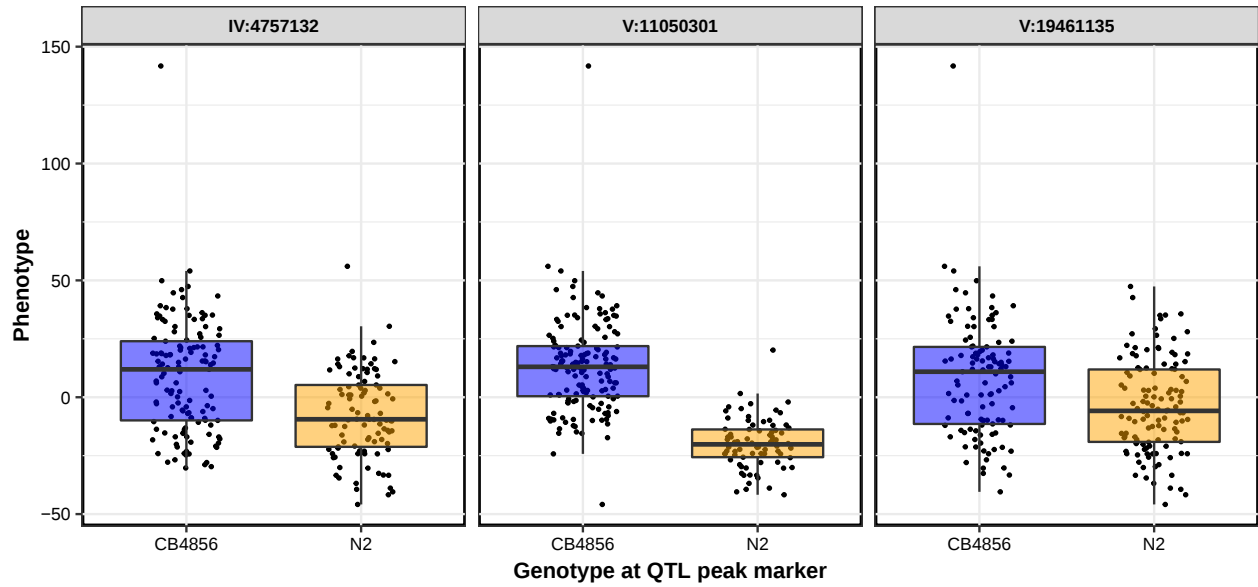
B



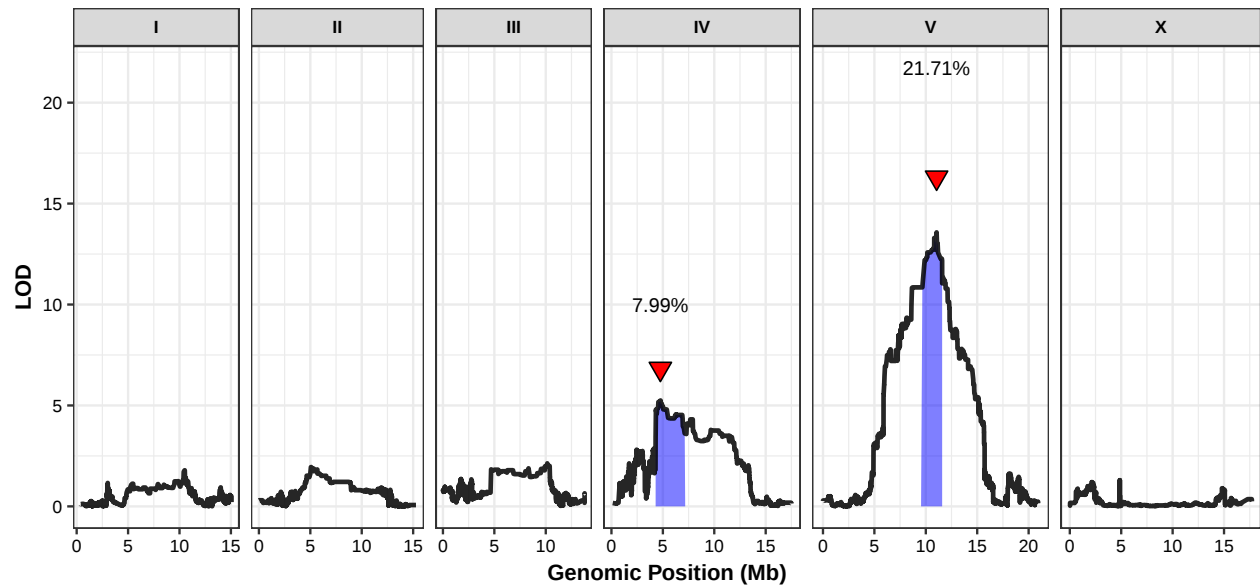
A Bleomycin q10.EXT



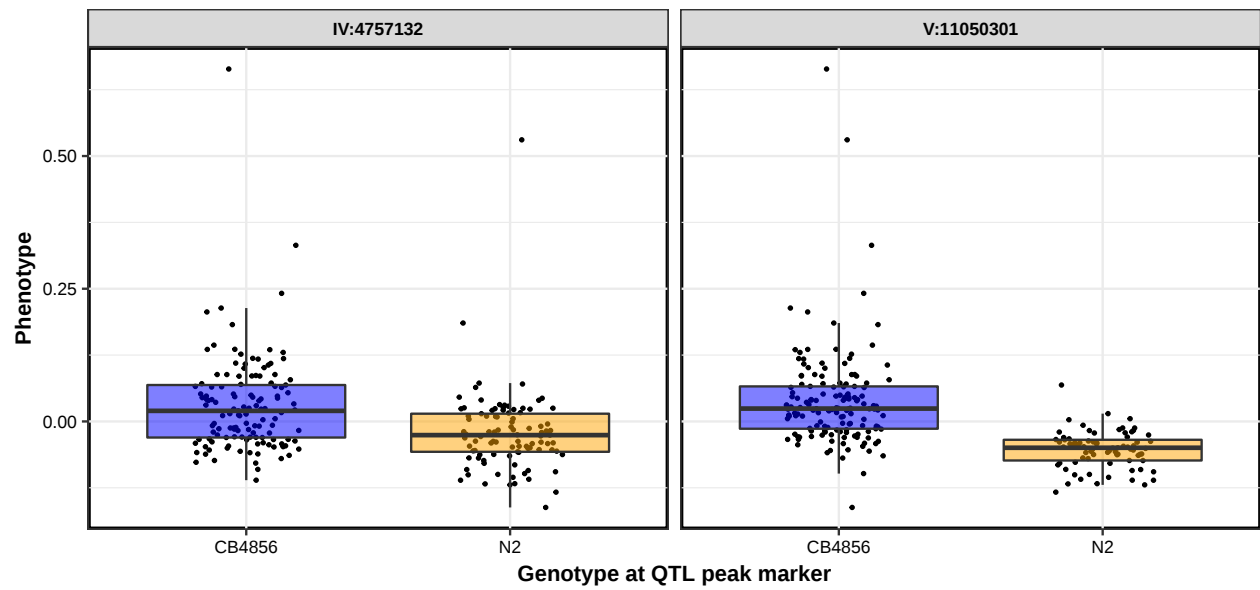
B



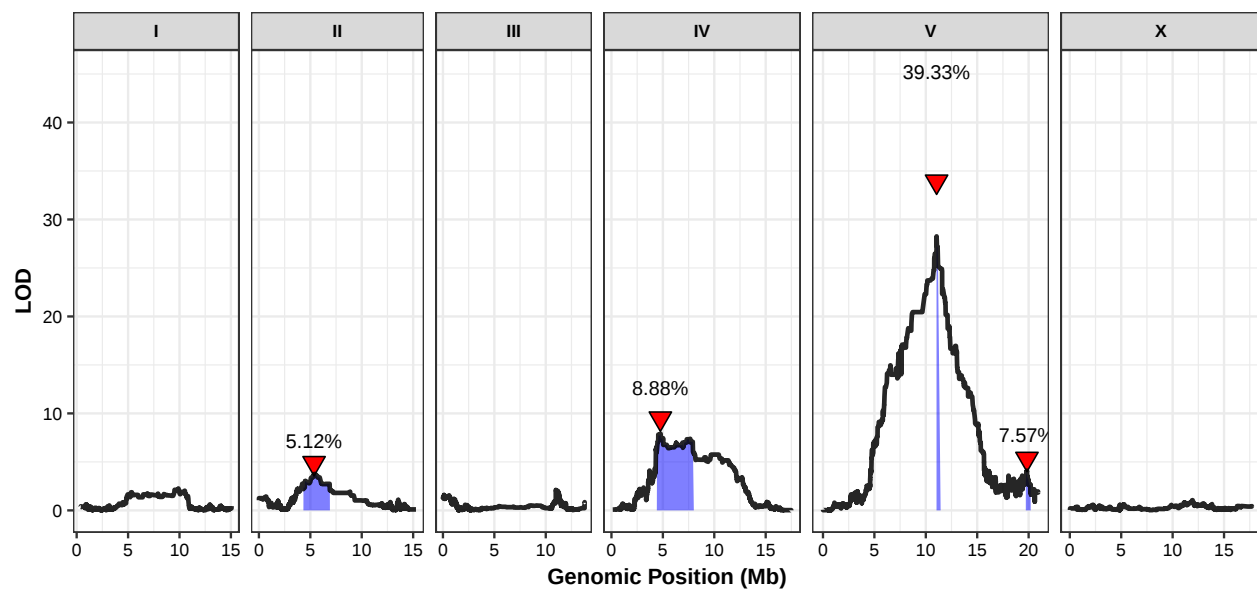
A Bleomycin q10.norm.EXT



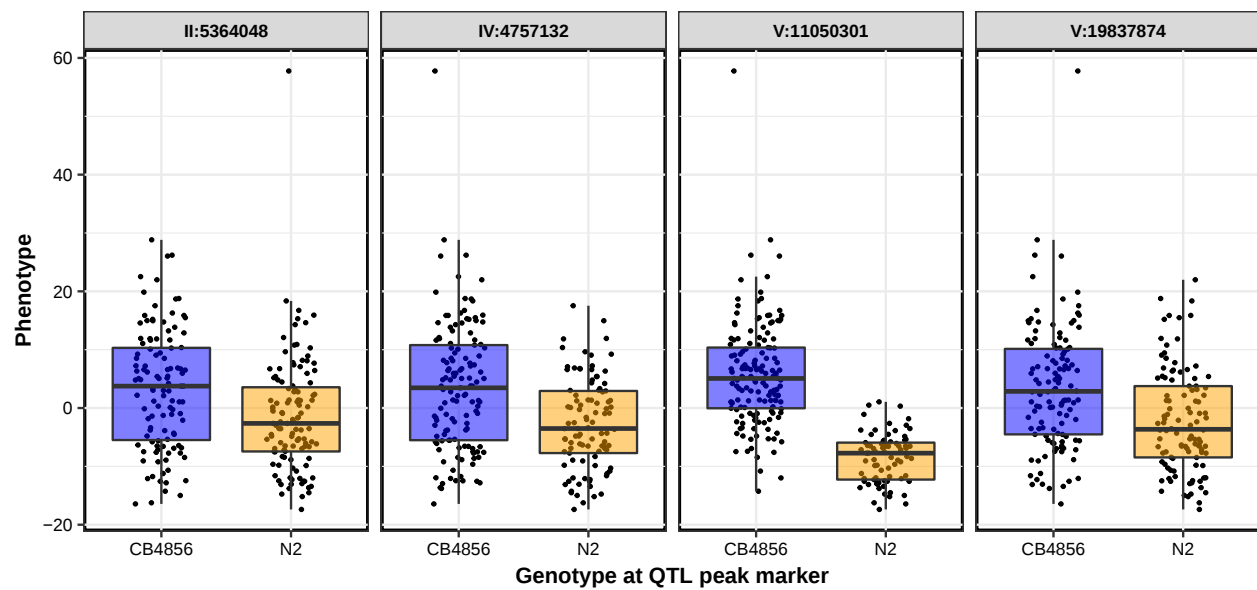
B



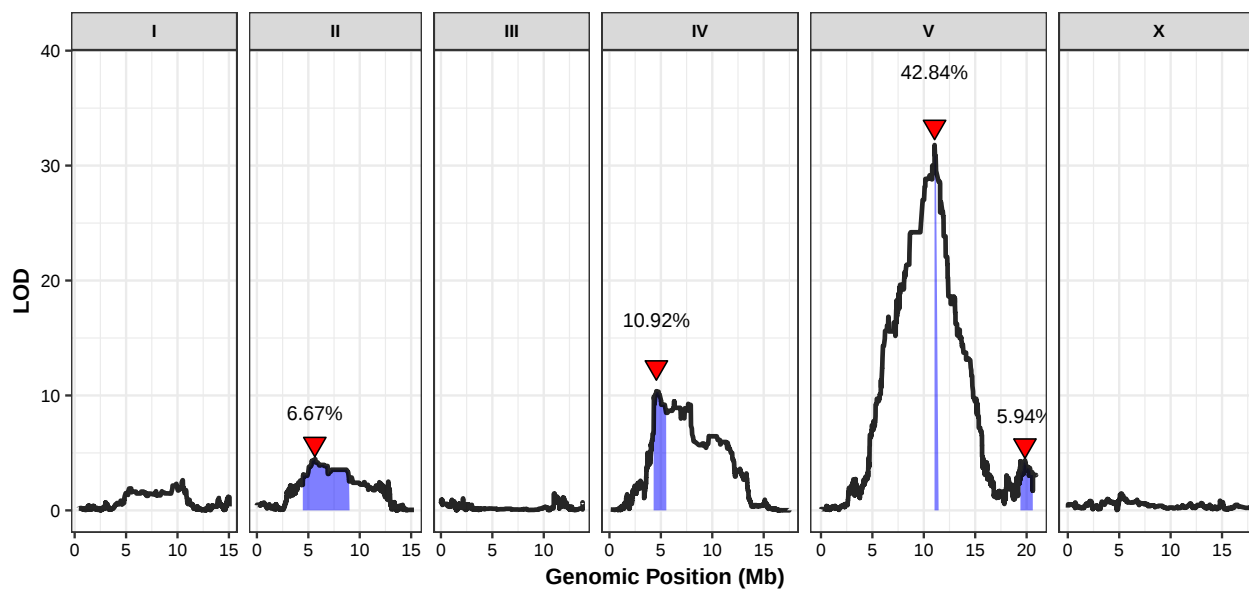
A Bleomycin q10.TOF



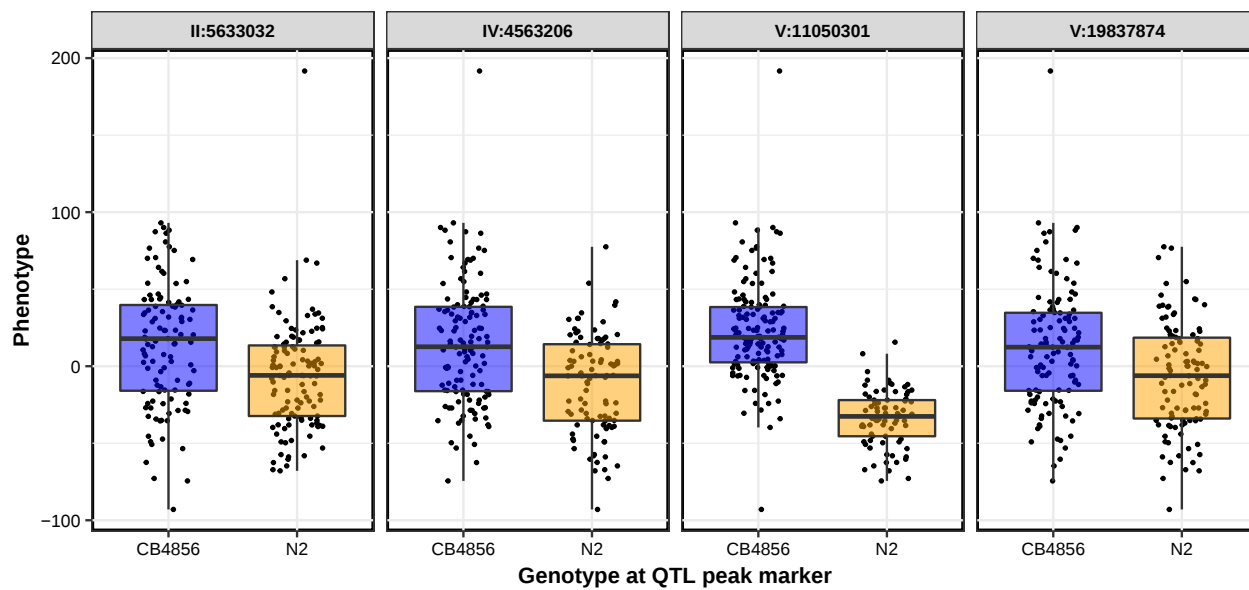
B



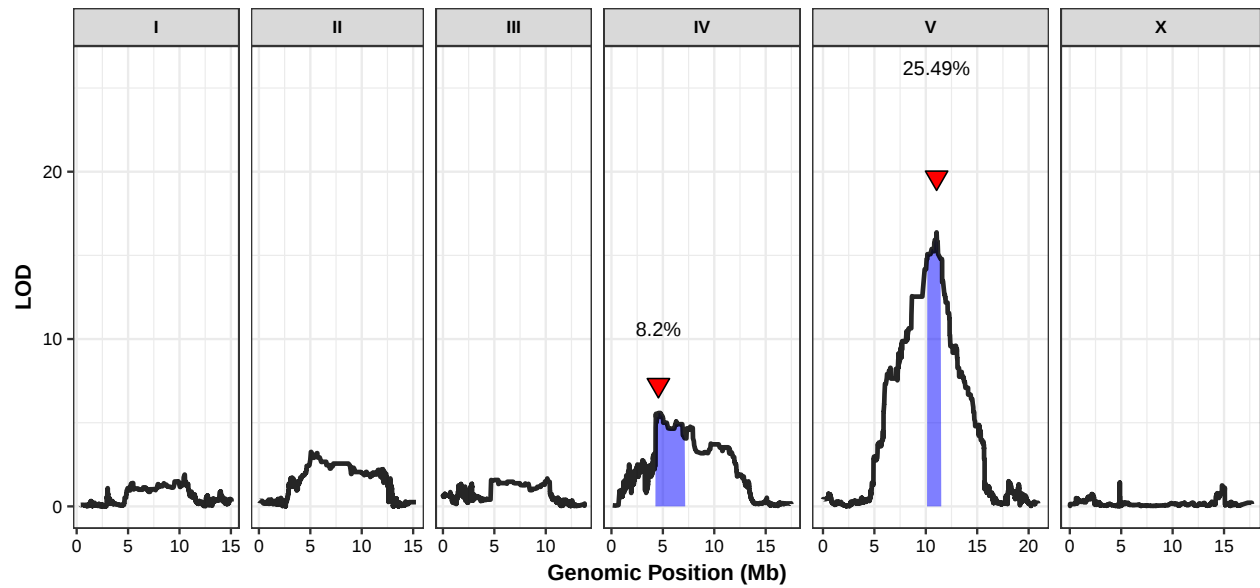
A Bleomycin q25.EXT



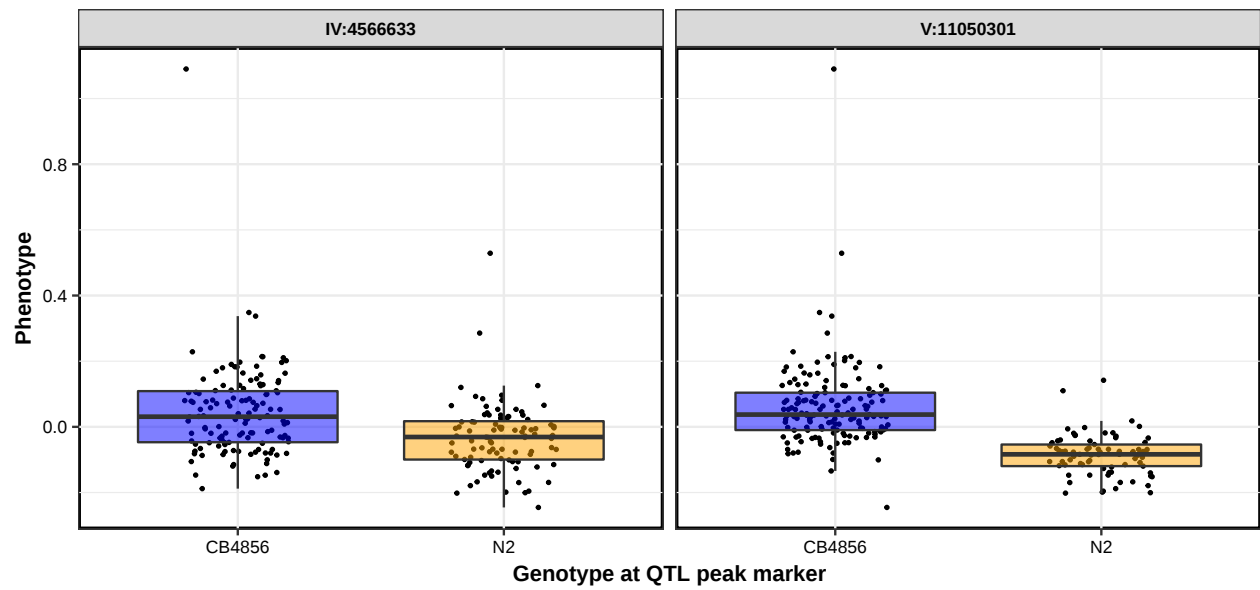
B



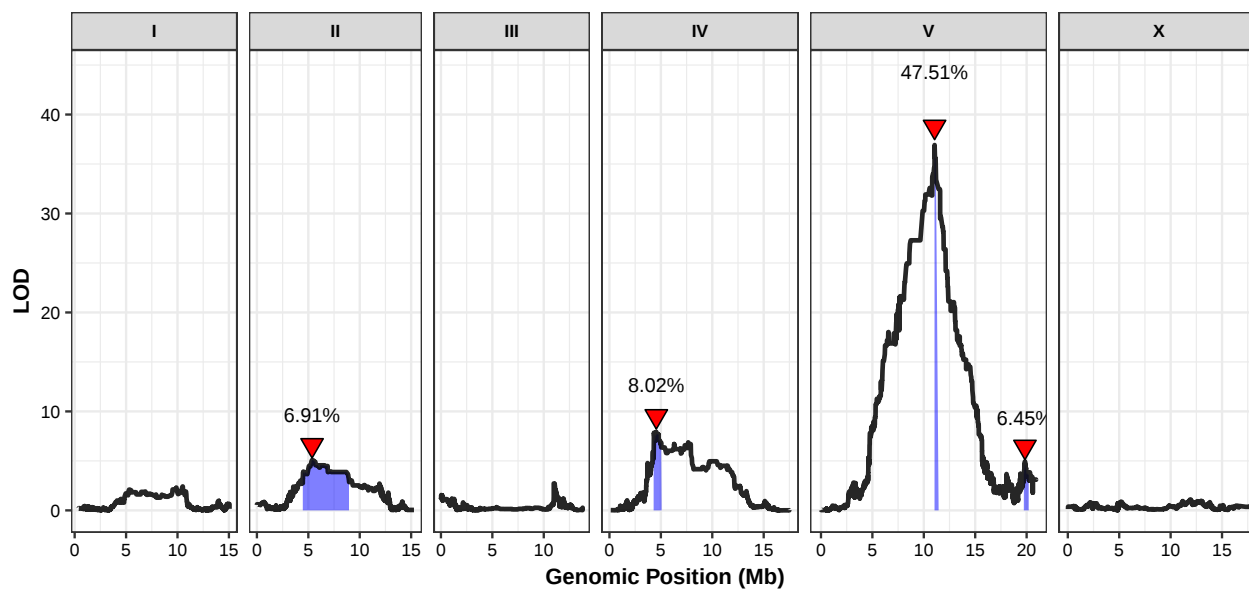
A Bleomycin q25.norm.EXT



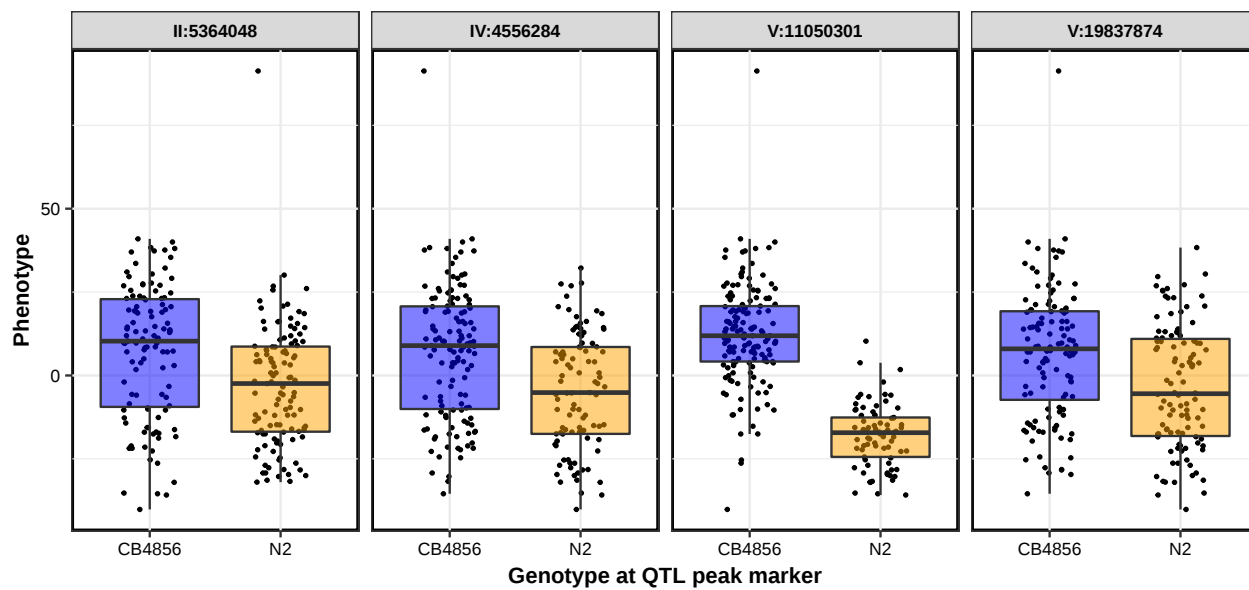
B



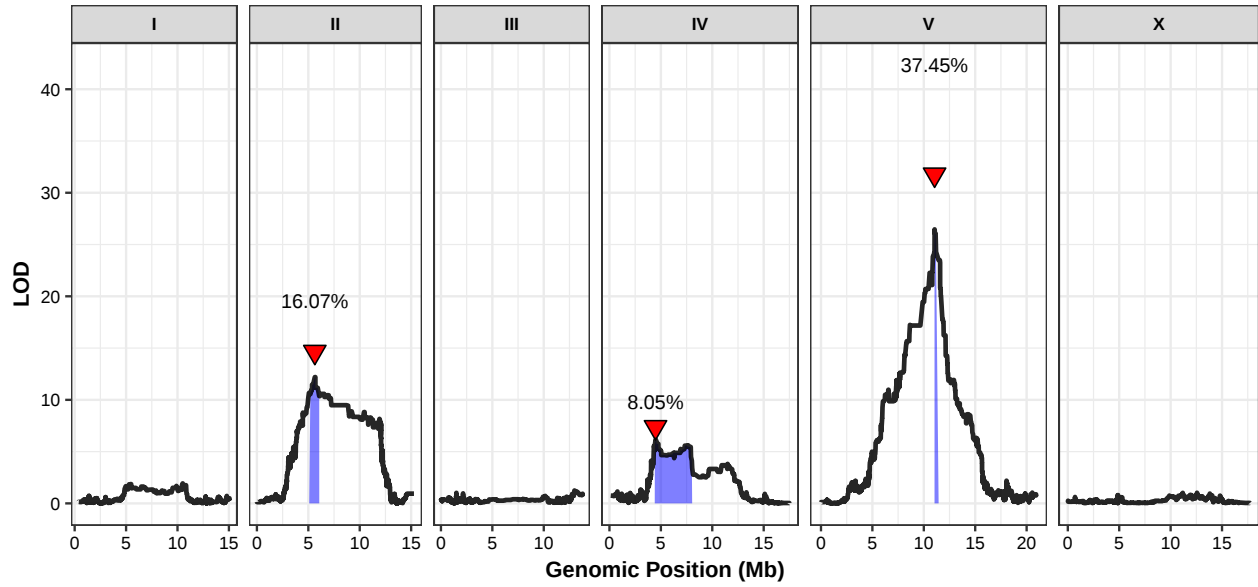
A Bleomycin q25.TOF



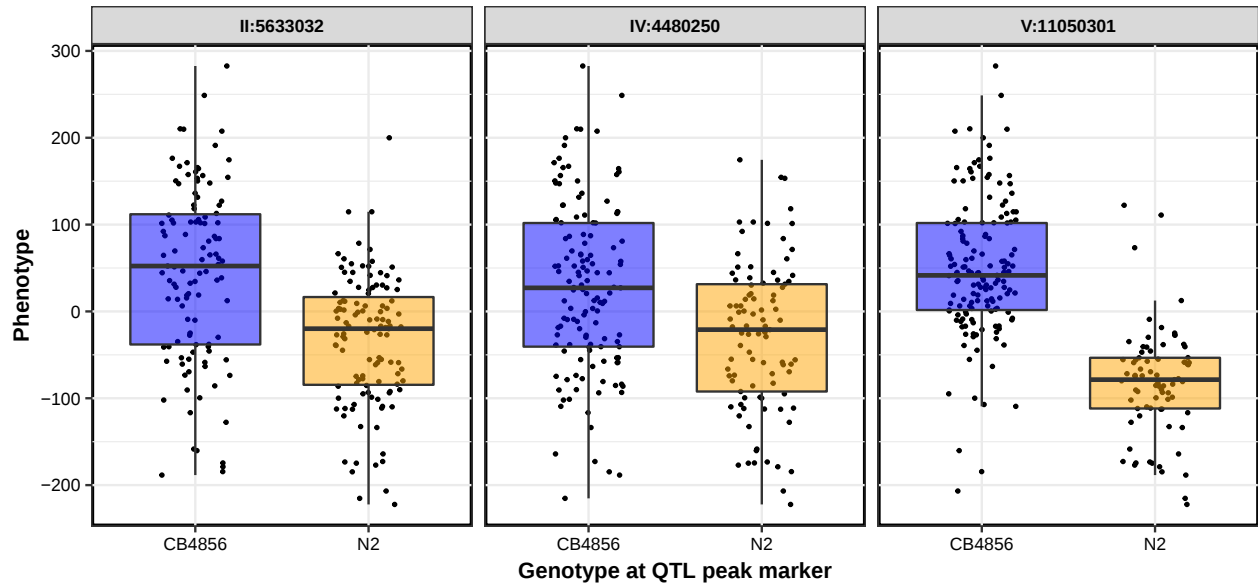
B



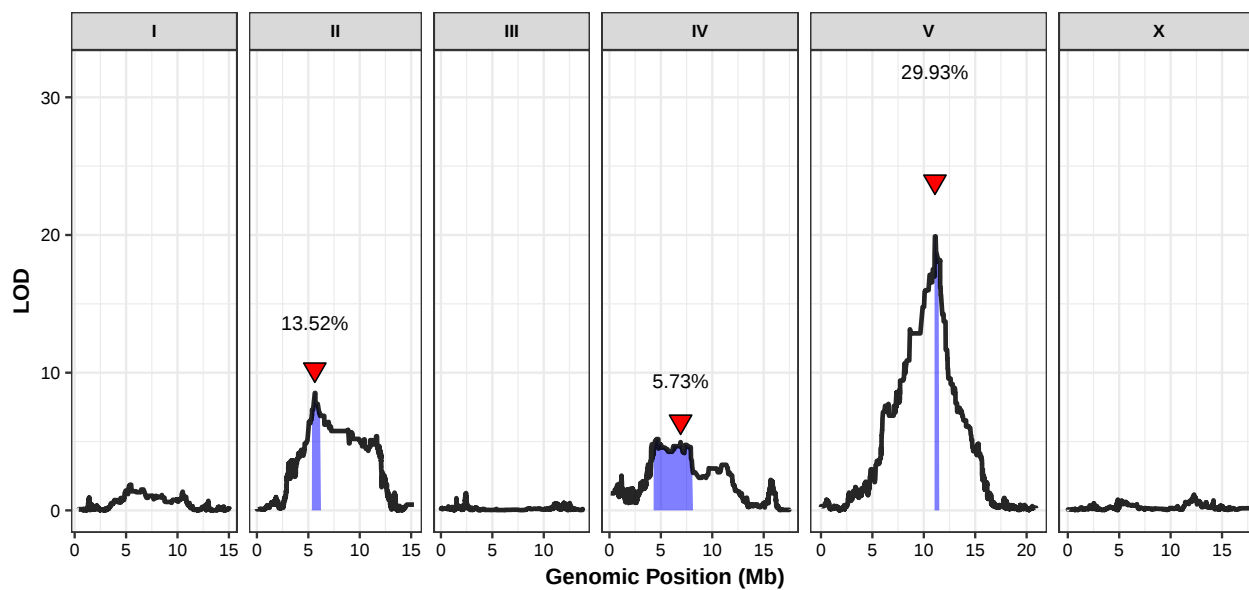
A Bleomycin q75.EXT



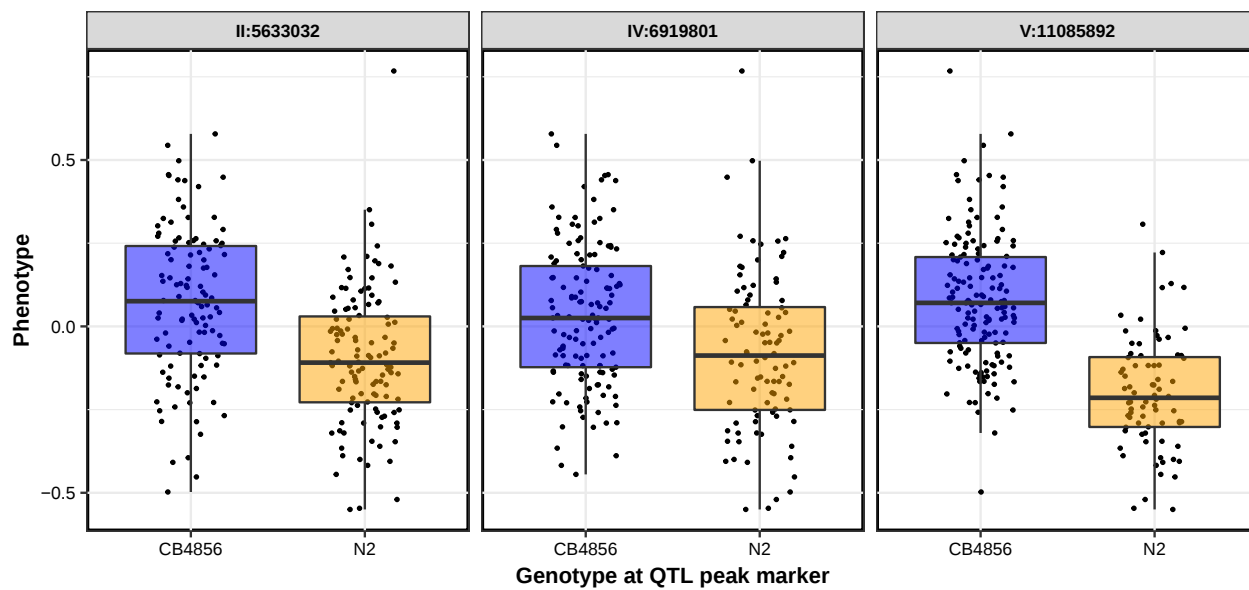
B



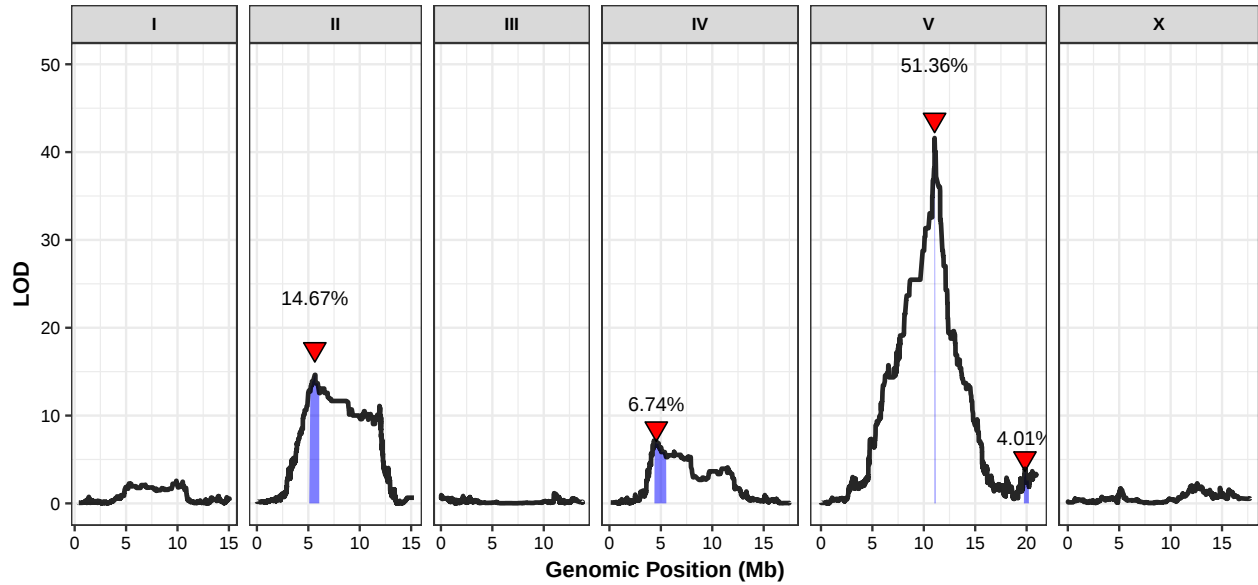
A Bleomycin q75.norm.EXT



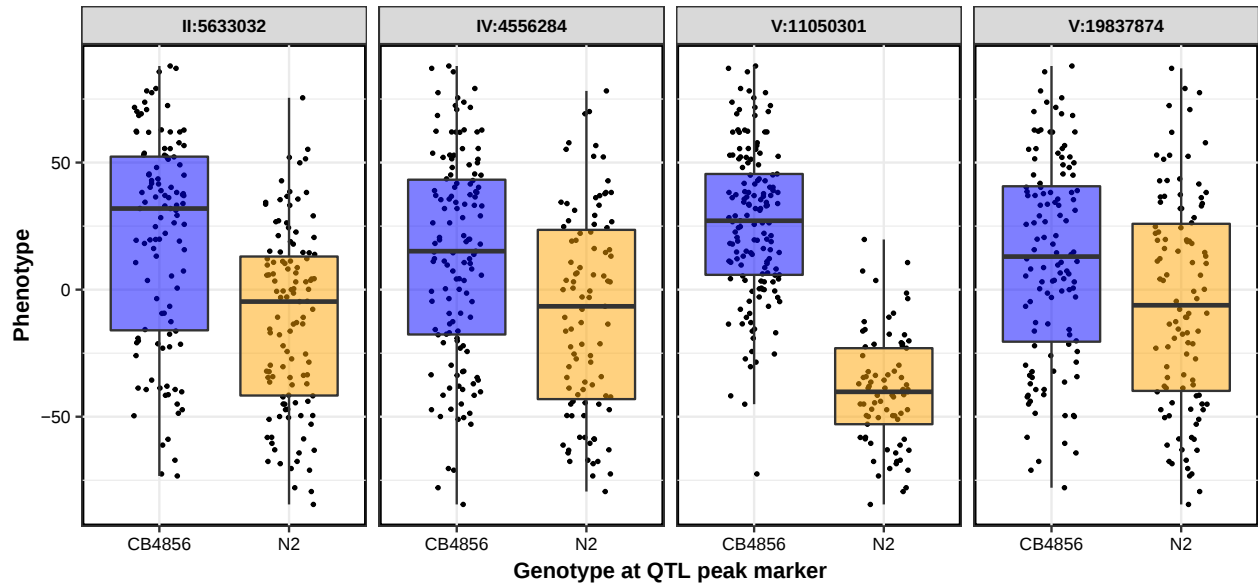
B



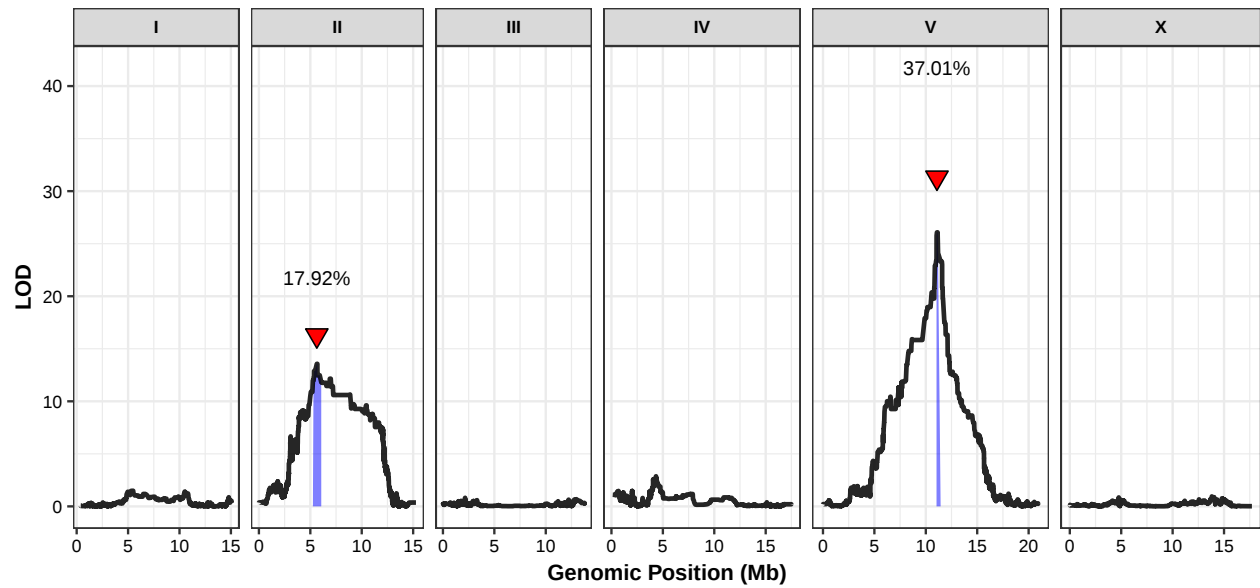
A Bleomycin q75.TOF



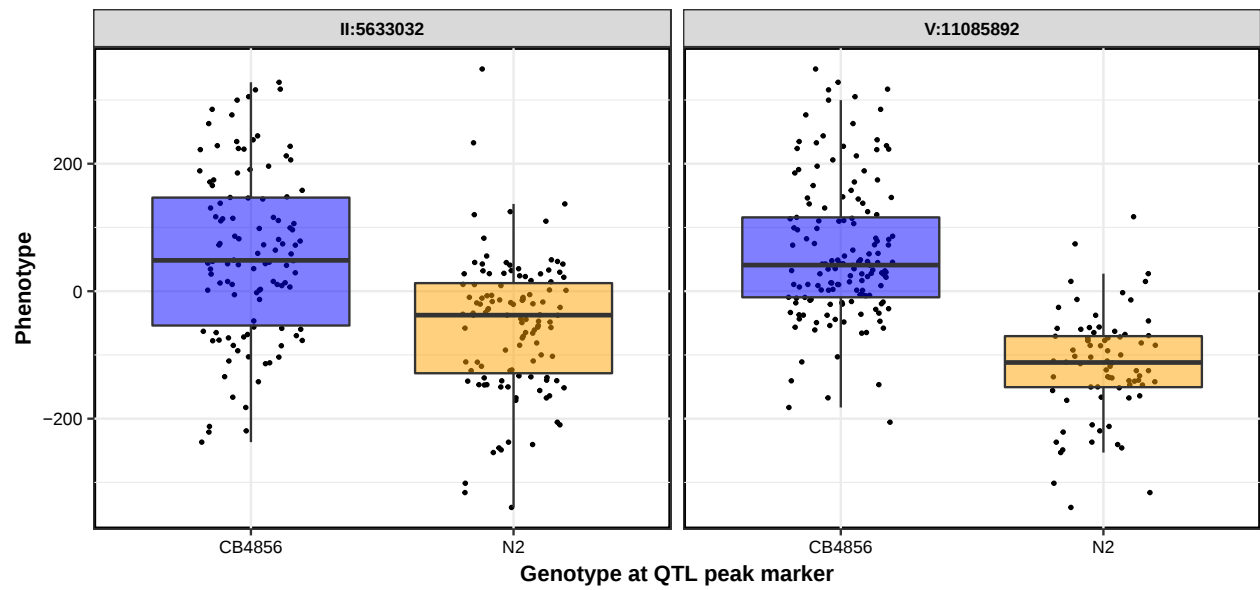
B



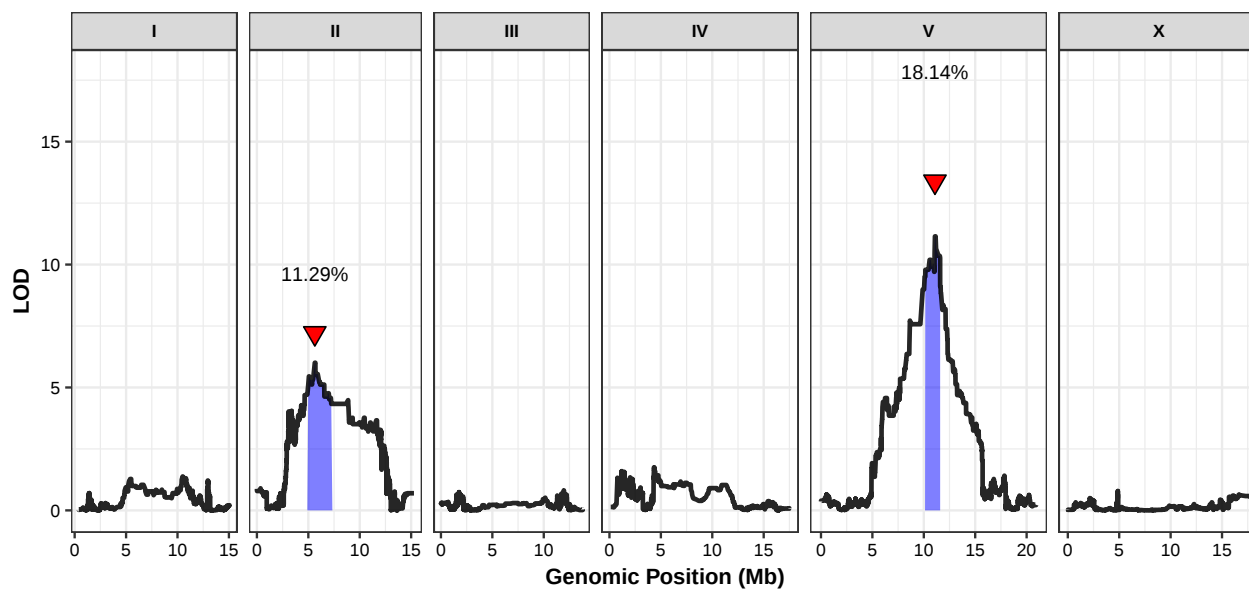
A Bleomycin q90.EXT



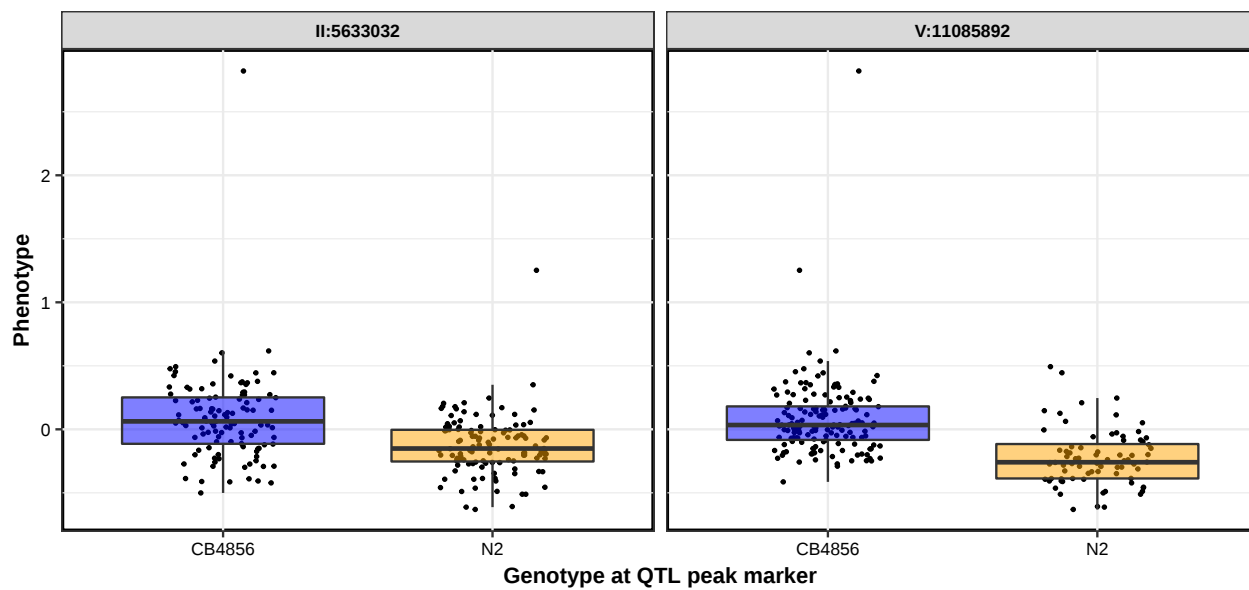
B



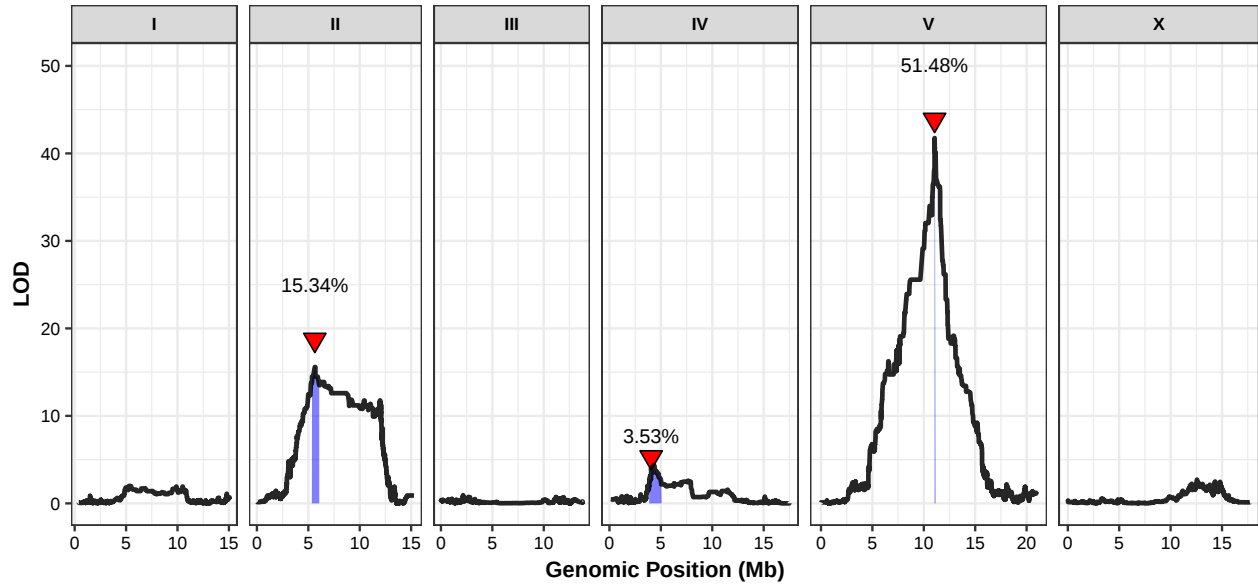
A Bleomycin q90.norm.EXT



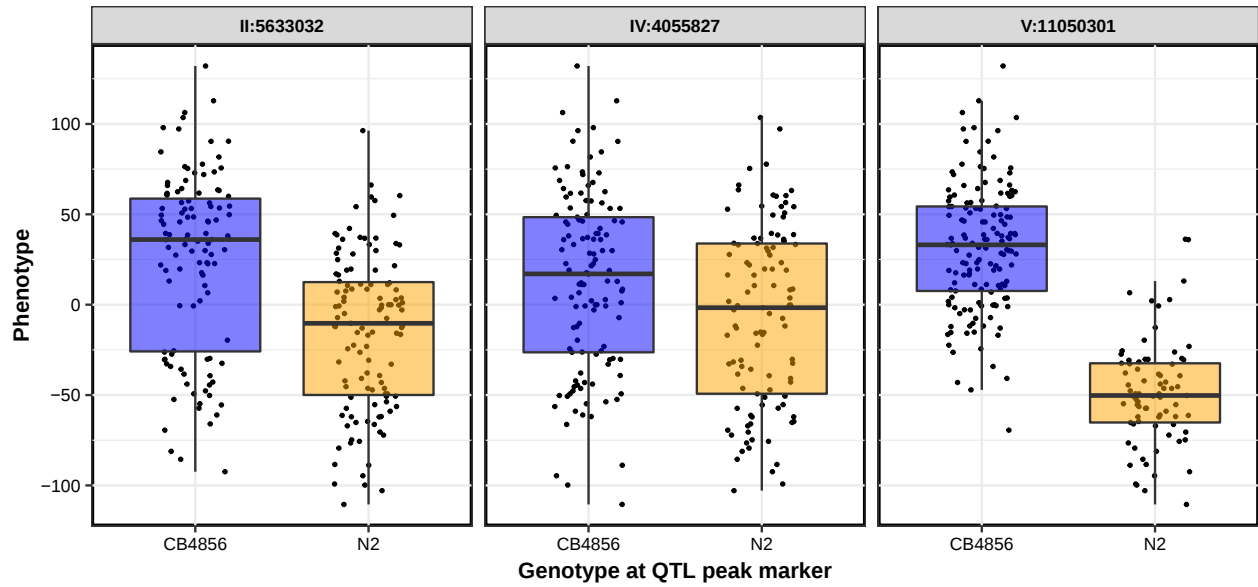
B

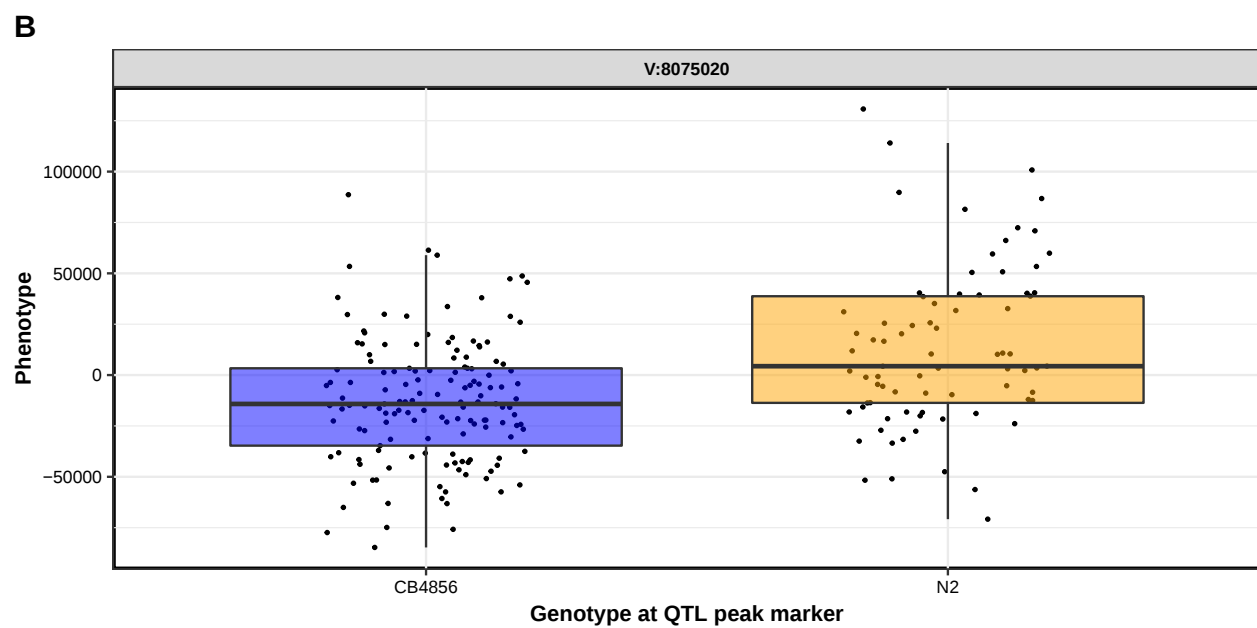
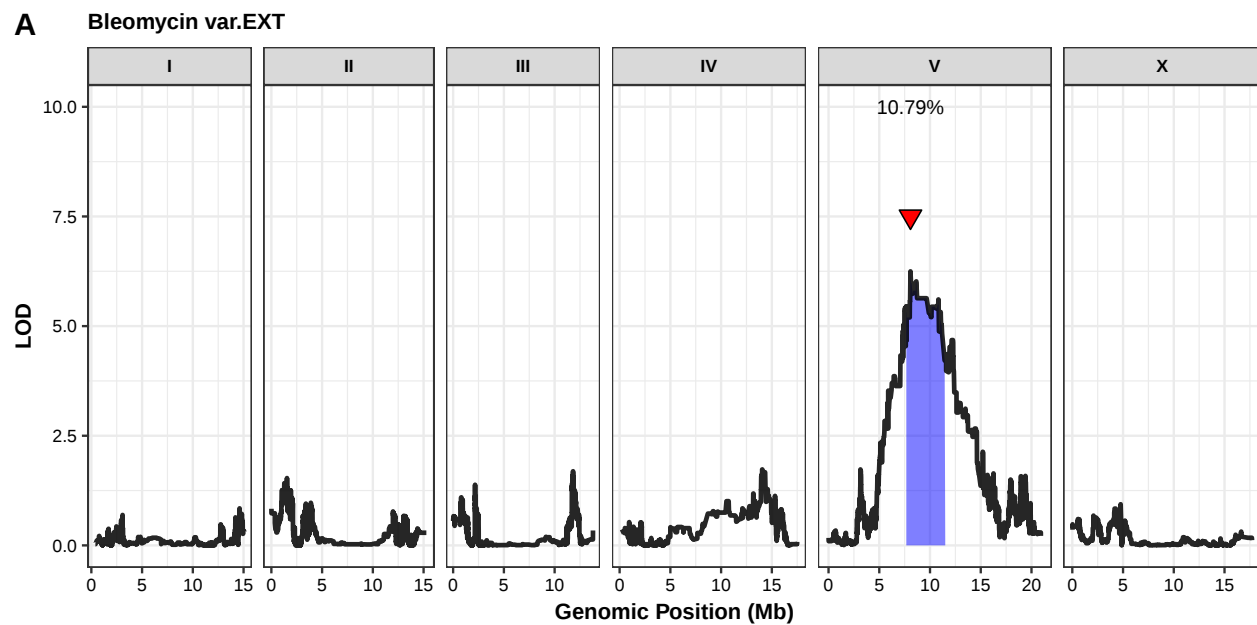


A Bleomycin q90.TOF



B





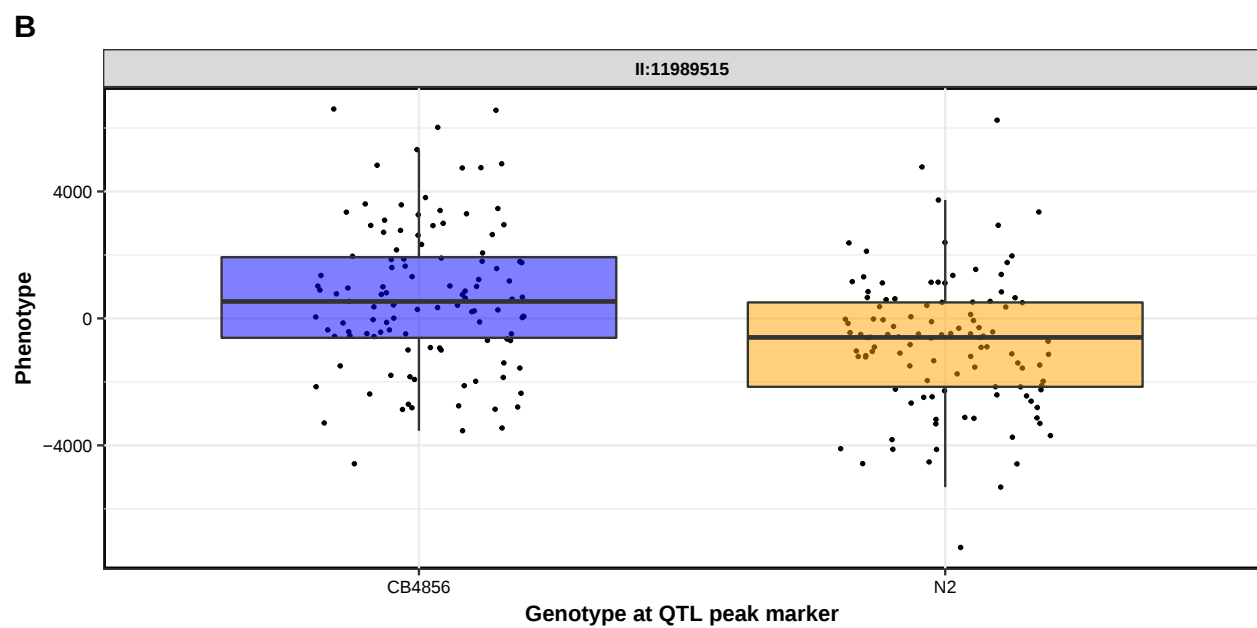
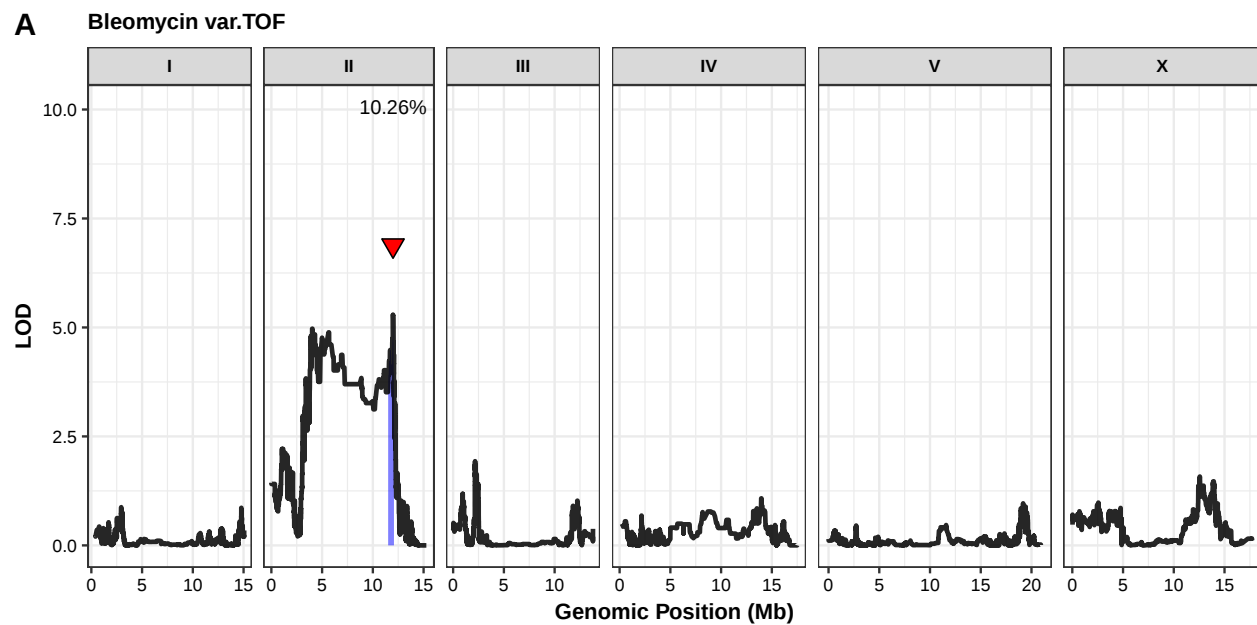


Figure S5

Linkage-mapping analysis of bleomycin-response variation for all 26 high-throughput traits are shown. **(A)** On the x-axis, each of 13,001 genomic markers, split by chromosome, were tested for correlation with phenotypic variation across the RIAIL panel. The log of the odds (LOD) score for each marker is reported on the y-axis. Each significant quantitative trait locus (QTL) is indicated by a red triangle at the peak marker, and a blue ribbon shows the 95% confidence interval around the peak marker. The total amount of phenotypic variance across the RIAIL panel explained by the genotype at each peak marker is shown as a percentage. **(B)** RIAIL phenotypes (y-axis), split by allele at each QTL peak marker (x-axis) are reported. For each significant QTL, phenotypes of RIAILs containing the N2 allele (orange) are compared to phenotypes of RIAILs containing the CB4856 allele (blue). Phenotypes are shown as Tukey box plots, and each point is the phenotype of an individual strain.

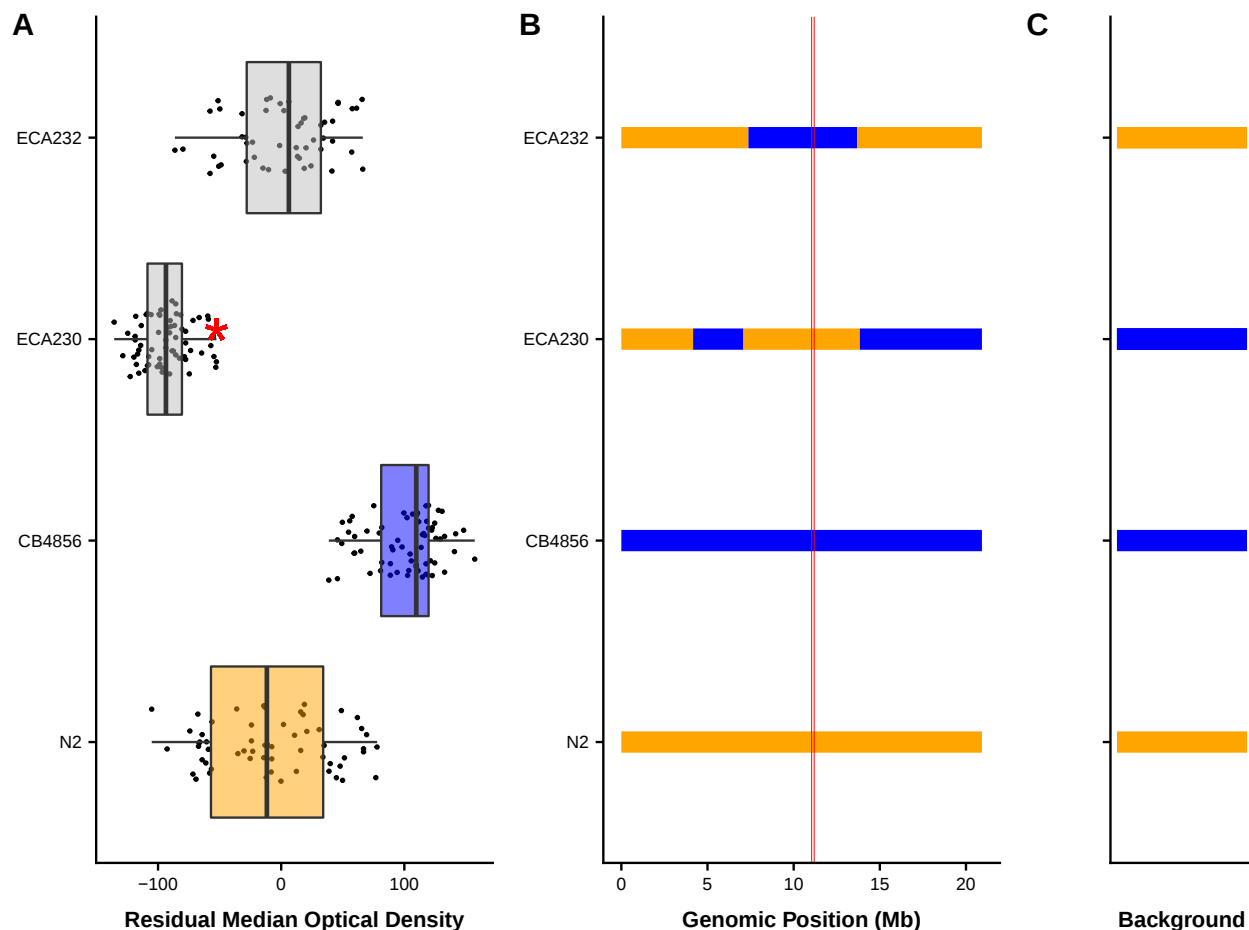


Figure S6

Phenotypes and genotypes of NIL strains are shown. **(A)** Phenotypes for each strain are shown as Tukey box plots, with strain name on the y-axis and residual bleomycin median optical density on the x-axis. Each point is a biological replicate. Parental strain box plots are colored by their genetic background, with orange indicating an N2 background and blue indicating a CB4856 genetic background. NILs are shown as grey box plots. A red asterisk indicates a significant difference between the phenotype of a given strain and the phenotype of the corresponding parental strain ($p < 0.05$, Tukey HSD). **(B)** Chromosomal representations of chromosome V are shown for each of the strains in A. Strain names are reported on the y-axis, and genomic position (Mb) is shown on the x-axis. Blocks of color indicate genotypes of genomic regions with orange indicating the N2 genotype and blue indicating the CB4856 genotype. Vertical red lines mark the confidence interval of the QTL from linkage mapping. **(C)** Background genotypes are represented as rectangles with colors indicating N2 (orange) or CB4856 (blue) genetic backgrounds.

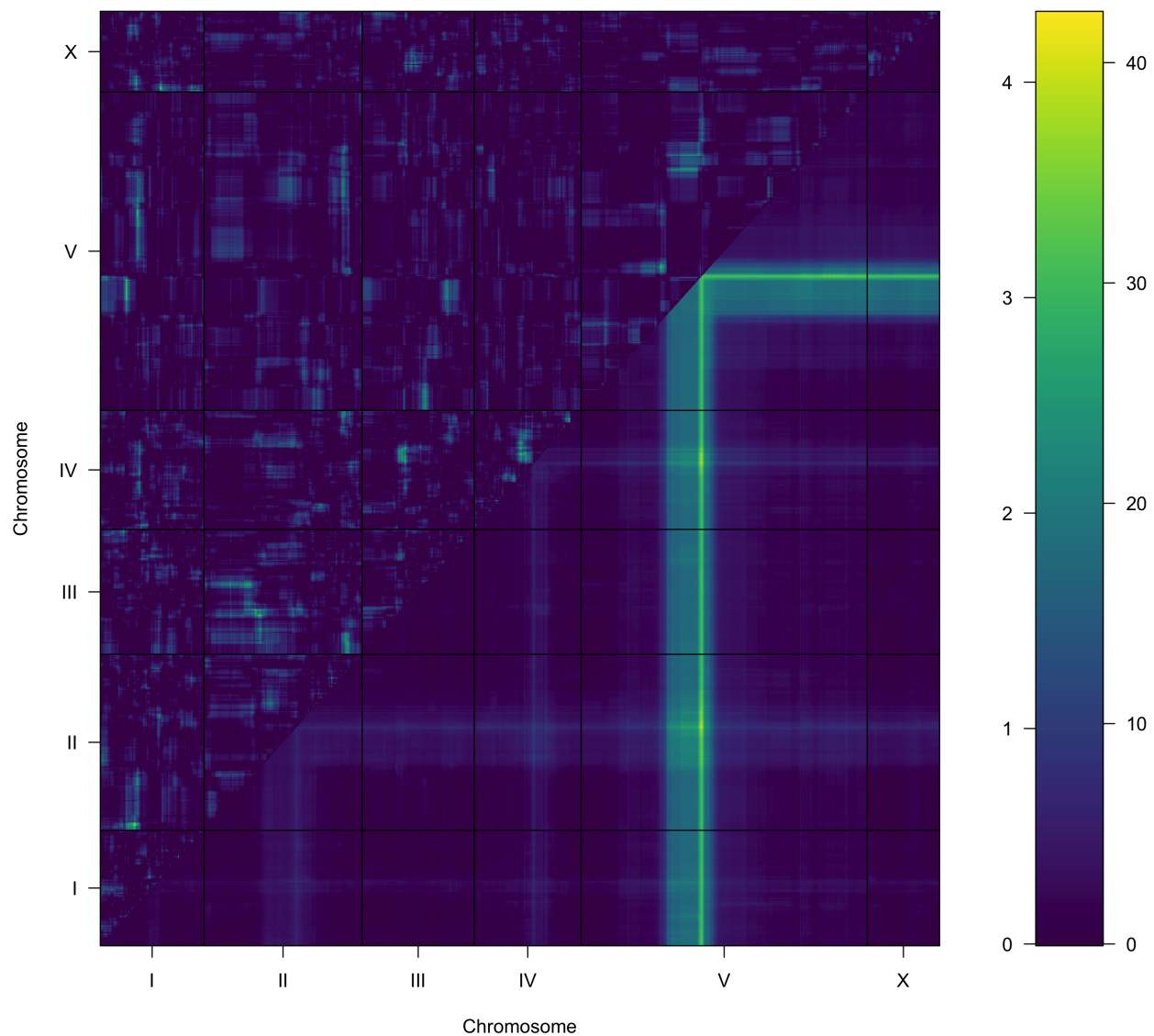


Figure S7

A two-factor genome scan for residual optical density in bleomycin is shown. Log of the odds (LOD) scores are shown for each pairwise combination of loci, split by chromosome. The upper-left triangle contains the epistasis LOD scores, and the lower-right triangle contains LOD scores for the full model. LOD scores are shown as colors, with lower scores in blue and higher scores in yellow, as shown in the color scale. The epistatic LOD score axis is on the left of the color scale and the full LOD score axis is on the right of the color scale.

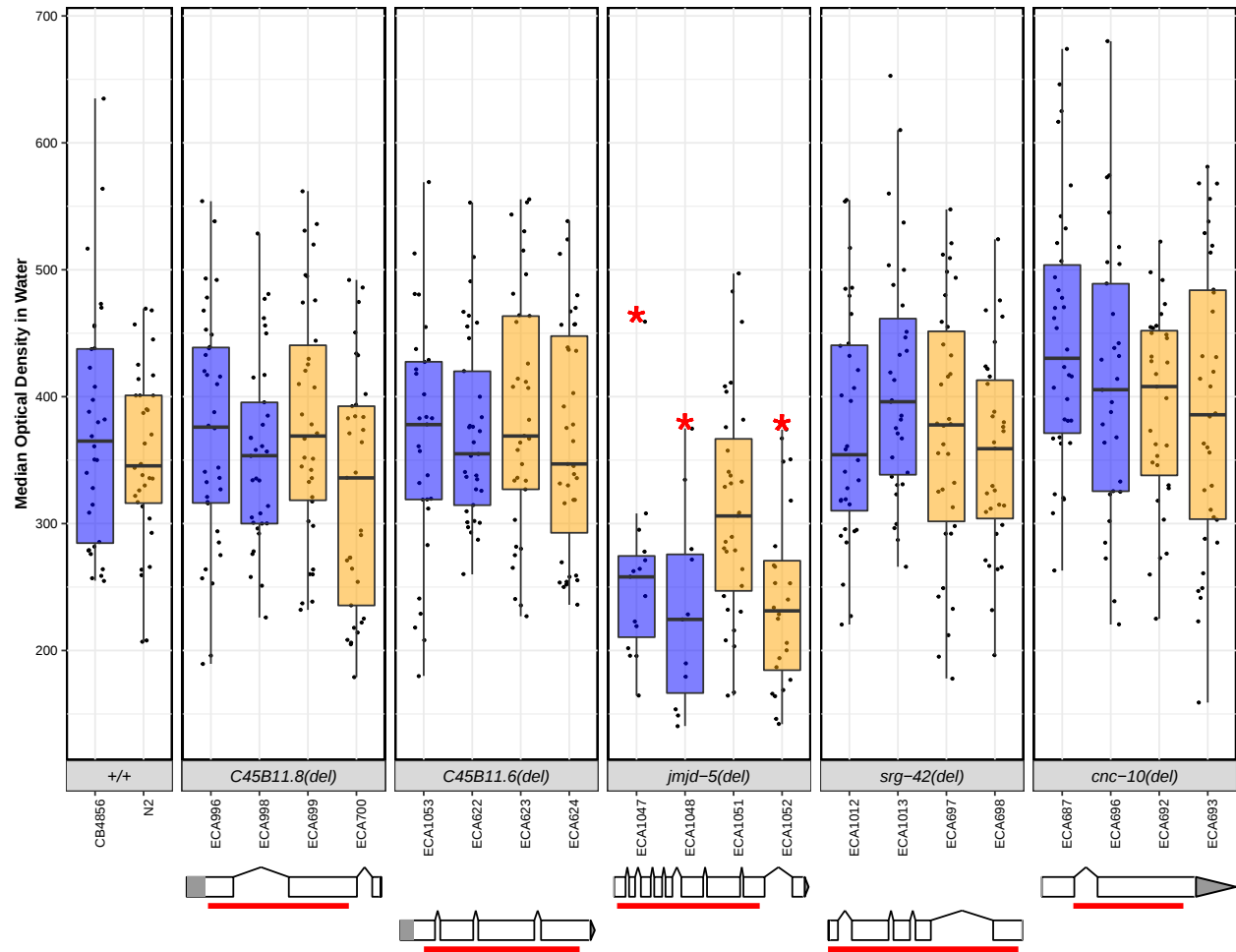


Figure S8

Control-condition phenotypes of deletion alleles for each candidate gene are shown. Deletion alleles for each candidate gene were tested in response to the control condition (lysate in K medium with 1% distilled water). Control-condition responses are shown as Tukey box plots, with the strain name on the x-axis, split by gene, and median optical density on the y-axis. Each point is a biological replicate. Strains are colored by their background genotype (orange indicates an N2 genetic background, and blue indicates a CB4856 genetic background). For each gene, two independent deletion alleles in each background were created and tested. Red asterisks indicate a significant difference ($p < 0.05$, Tukey HSD) between a strain with a deletion and the parental strain that has the same genetic background. Depictions of each deletion allele are shown below the phenotype for each candidate gene. White rectangles indicate exons and diagonal lines indicate introns. The 5' and 3' UTRs are shown by grey rectangles and triangles, respectively. The region of the gene that was deleted by CRISPR-Cas9 directed genome editing is shown as a red bar beneath each gene model.

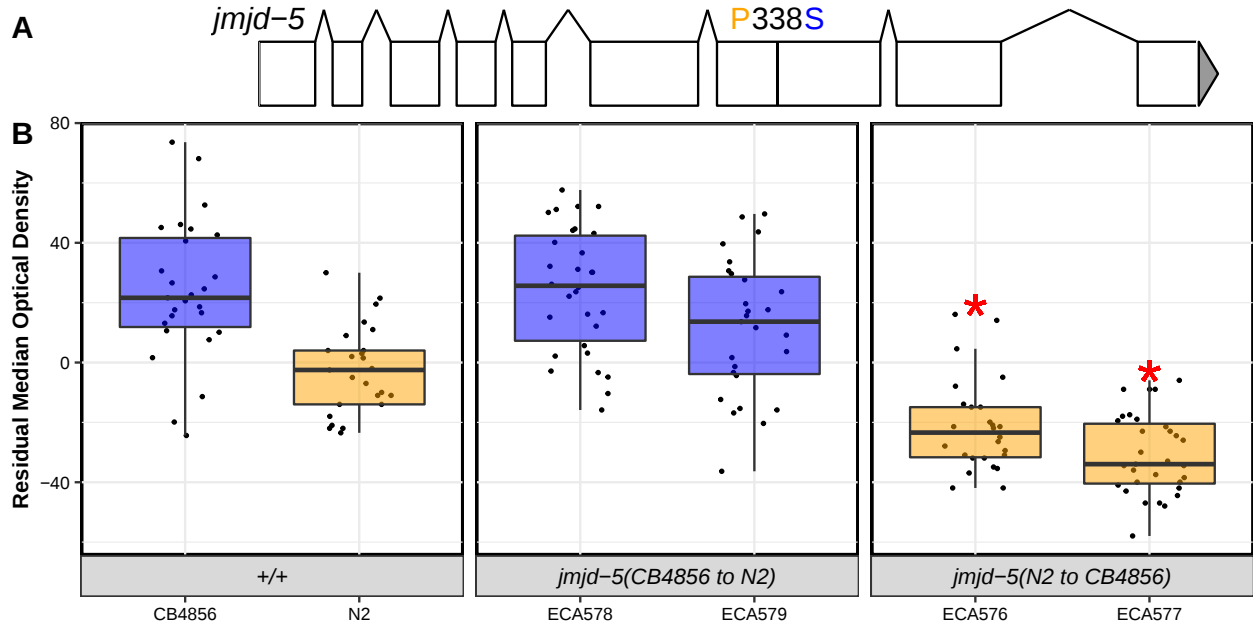


Figure S9

Reciprocal allele-replacement strains of *jmjd-5* were tested in response to bleomycin. **(A)** A model of *jmjd-5* is shown with white rectangles indicating exons and black diagonal lines indicating introns. The 5' and 3' UTRs are shown as a grey rectangle and triangle, respectively. The location of the amino-acid variant between the N2 and CB4856 strains is shown in black text above the gene depiction with the N2 residue in orange and the CB4856 residue in blue. **(B)** Bleomycin responses of allele-replacement strains are shown as Tukey box plots, split and colored by genetic background (N2 in orange and CB4856 in blue) with the strain name on the x-axis and residual median optical density on the y-axis. Each point is a biological replicate. Red asterisks indicate a significant difference between an allele-replacement strain and the parental strain sharing its genetic background ($p < 0.05$, Tukey HSD).

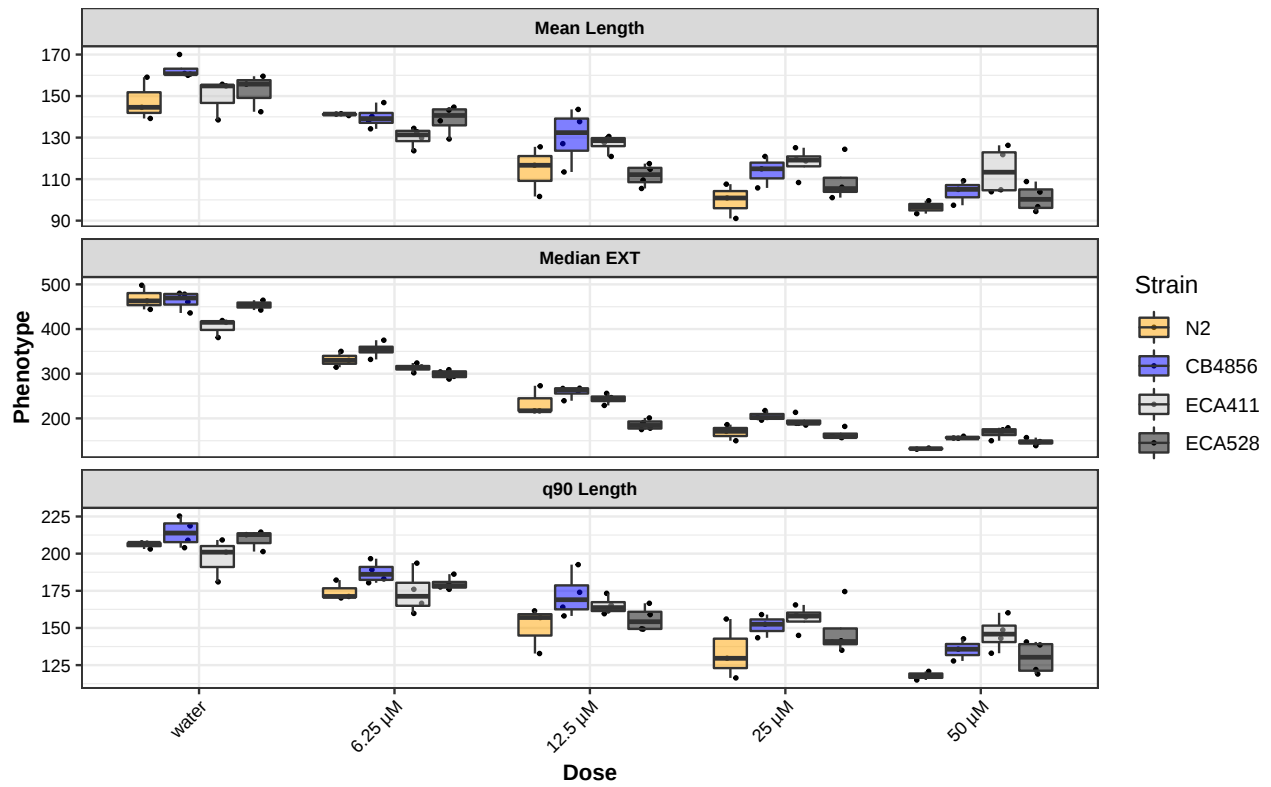


Figure S10

Dose-response phenotypes from the modified HTA are shown for three high-throughput fitness traits: mean length (mean.TOF), median optical density (median.EXT), and the 90th quantile of optical density (q90 EXT). Phenotypes for each of the four tested strains are shown as Tukey boxplots, colored by strain (N2 - orange, CB4856 - blue, ECA411 - light grey, ECA528 - dark grey). The x-axis shows the concentration of bleomycin (or water) to which the animals were exposed, and the y-axis shows the pruned phenotype. Each point is a biological replicate.

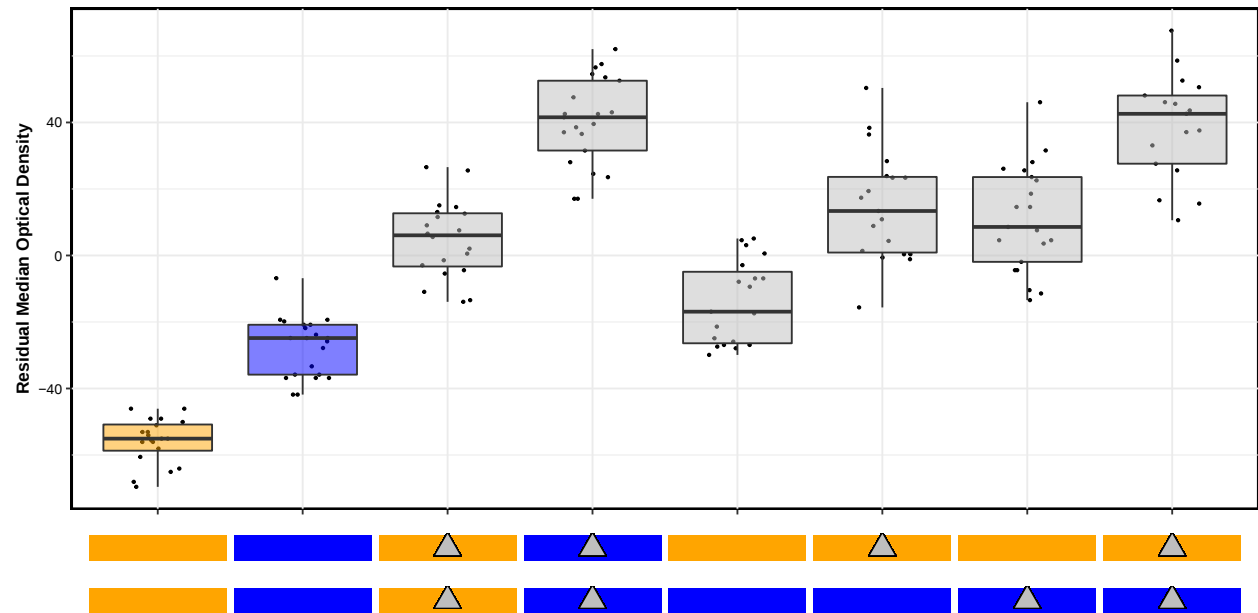


Figure S11

Results of the *jmjd-5* reciprocal hemizygosity assay are shown. The y-axis shows the residual median optical density for each strain reported along the x-axis. Bleomycin responses are reported as Tukey box plots where each point is a biological replicate. The genotypes of each strain are shown as colored rectangles beneath each box plot, where each rectangle represents a homolog (orange rectangles are an N2 genotype, and blue rectangles are a CB4856 genotype). The maternal homolog is shown on top and the paternal homolog is shown on bottom. Grey triangles indicate a deletion of *jmjd-5*, placed on the rectangle showing the background into which the deletion was introduced. The box plots for the parental strains (N2 and CB4856, on the left) are colored according to genotype

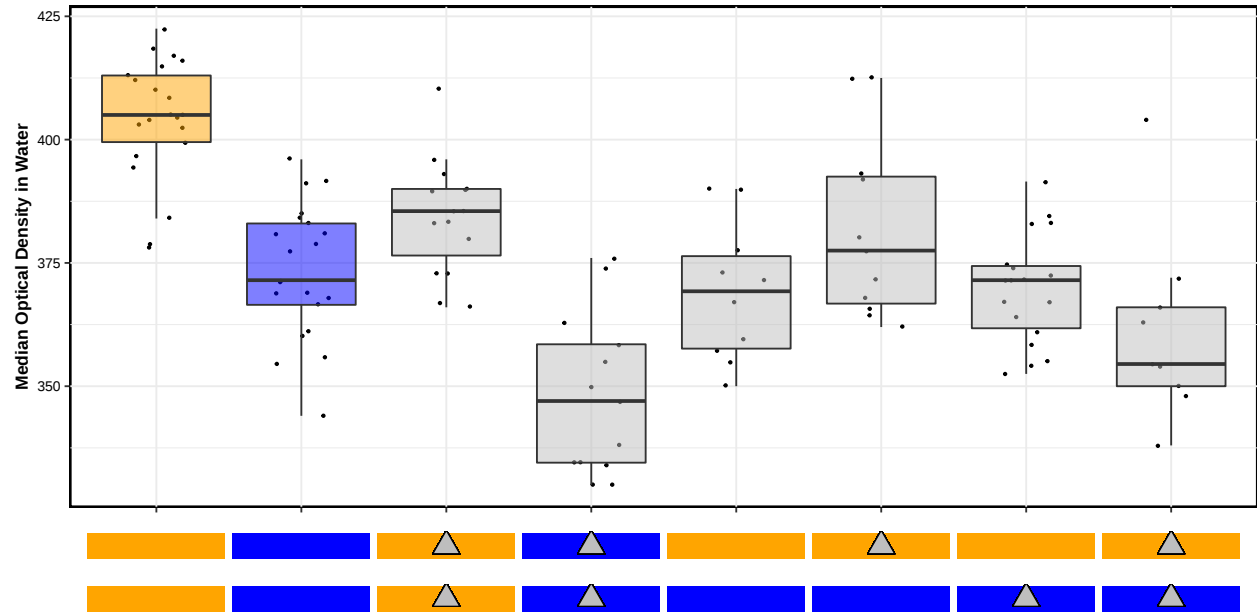


Figure S12

Control-condition phenotypes of the strains tested in the reciprocal hemizygosity experiment of *jmjD-5* are shown. The y-axis shows the pruned median optical density upon exposure to the control condition (lysate in K medium plus 1% distilled water) for each strain reported along the x-axis. Control-condition responses are reported as Tukey box plots where each point is a biological replicate. The genotypes of each strain are shown as colored rectangles beneath each box plot, where each rectangle represents a homolog (orange rectangles are an N2 genotype, and blue rectangles are a CB4856 genotype). The maternal homolog is shown on top and the paternal homolog is shown on bottom. Grey triangles indicate a deletion of *jmjD-5*, placed on the rectangle showing the background into which the deletion was introduced. The box plots for the parental strains (N2 and CB4856, on the left) are colored according to genotype

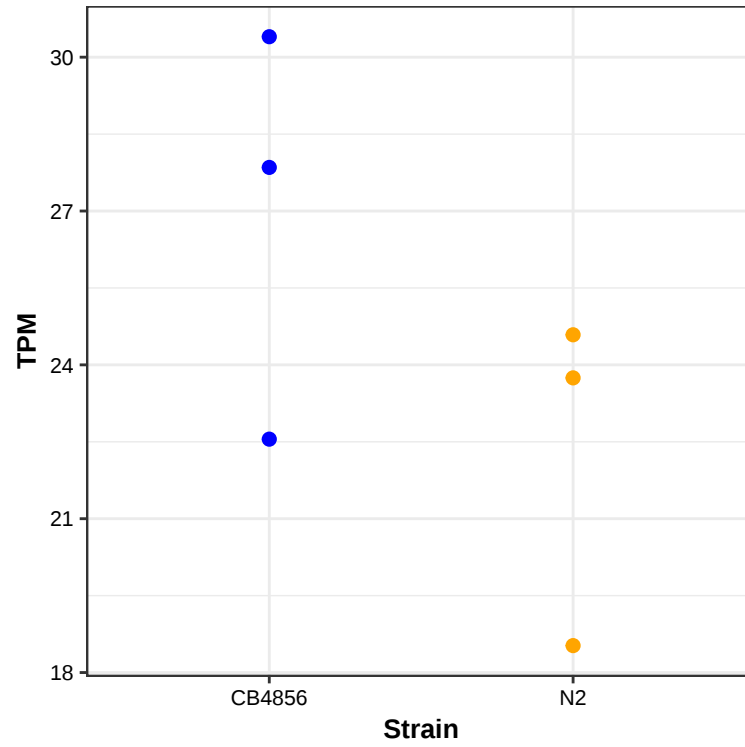


Figure S13

RNA-seq measurements of *scb-1* expression for young adult populations of N2 and CB4856 are reported. The x-axis indicates the sample (N2 or CB4856), and the y-axis shows the transcripts per million (TPM) estimate for each replicate. Replicates are plotted as dots, colored by strain (orange for N2 and blue for CB4856).

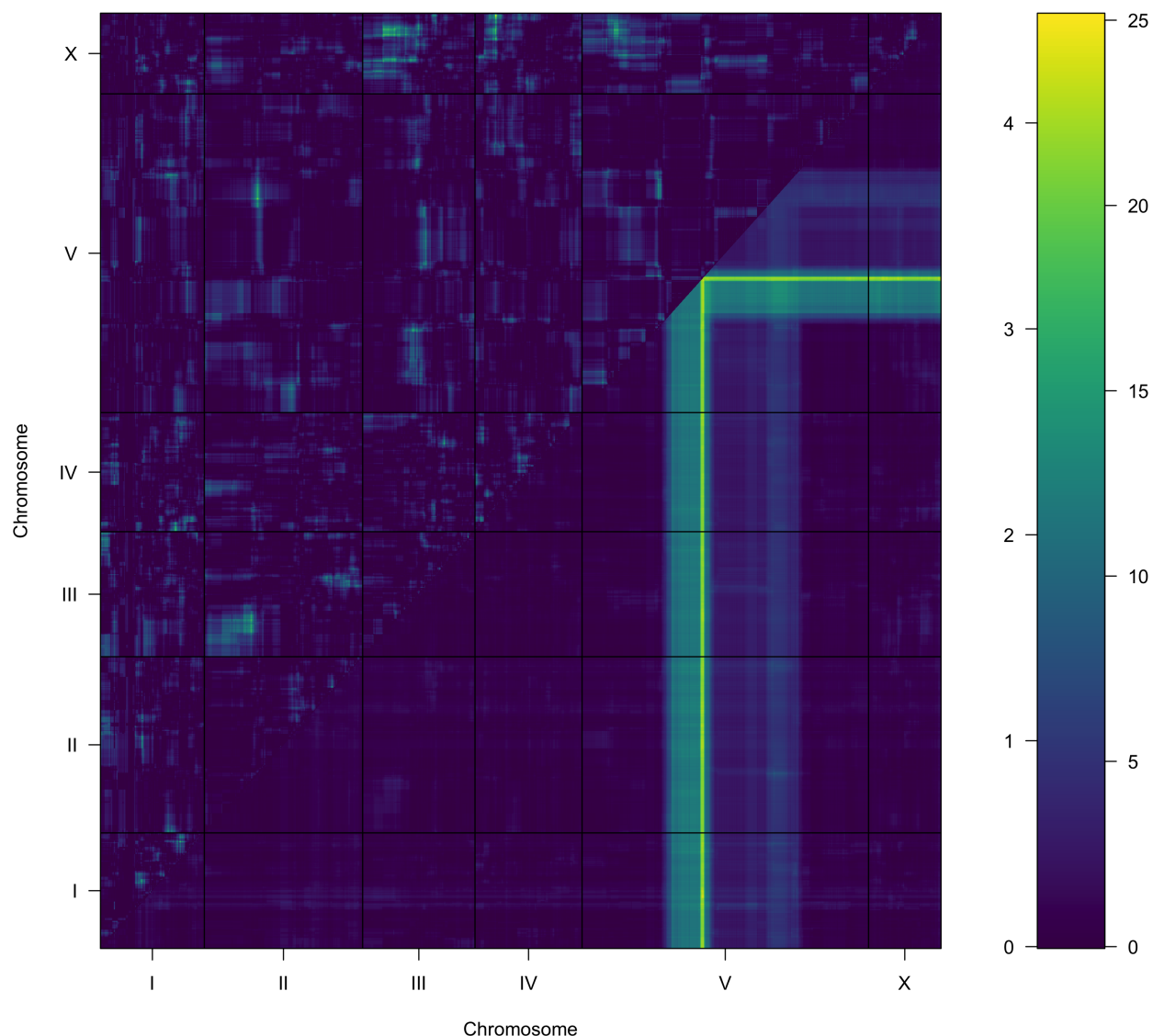


Figure S14

A two-factor genome scan for *scb-1* expression variation is shown. Log of the odds (LOD) scores are shown for each pairwise combination of loci, split by chromosome. The upper-left triangle contains the epistasis LOD scores, and the lower-right triangle contains LOD scores for the full model. LOD scores are shown as colors, with lower scores in blue and higher scores in yellow, as shown in the color scale. The epistatic LOD score axis is on the left of the color scale and the full LOD score axis is on the right of the color scale.

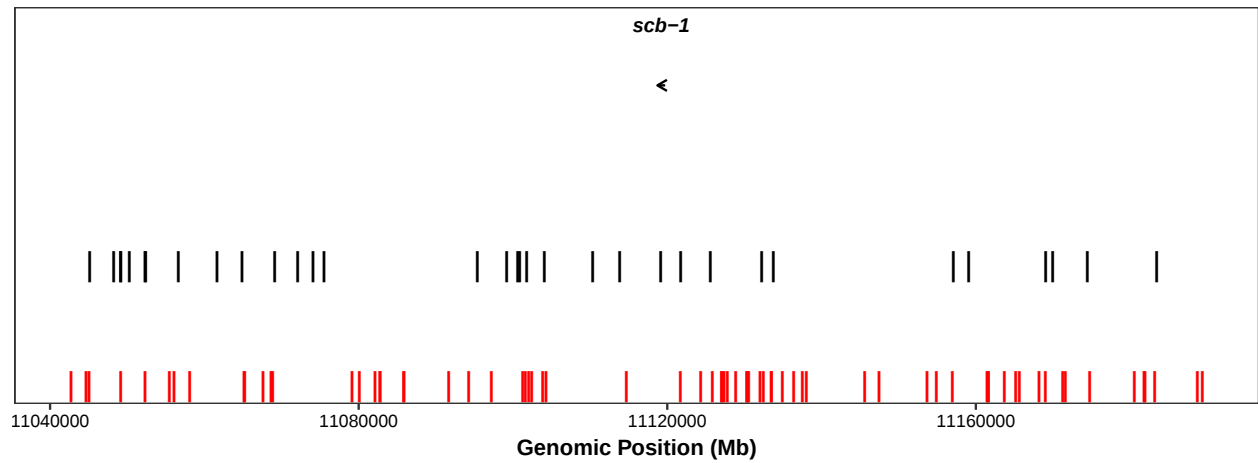


Figure S15

All variants in the QTL confidence interval for which CB4856 contains the alternate allele are plotted as vertical lines. On the x-axis, genomic position of the variant is shown. Variants are colored by minor allele frequency (less than 0.05 = red, greater than 0.05 = black). The location of *scb-1* is shown as an arrow, labeled with the gene name.

Bar chart showing median EXT values for various categories. The y-axis is labeled 'median EXT' and ranges from -100 to 100. The x-axis lists categories from N2 to G4946. Most categories have negative median EXT values, with a few positive values highlighted in blue: ED3017, CB4856, and CB4859.

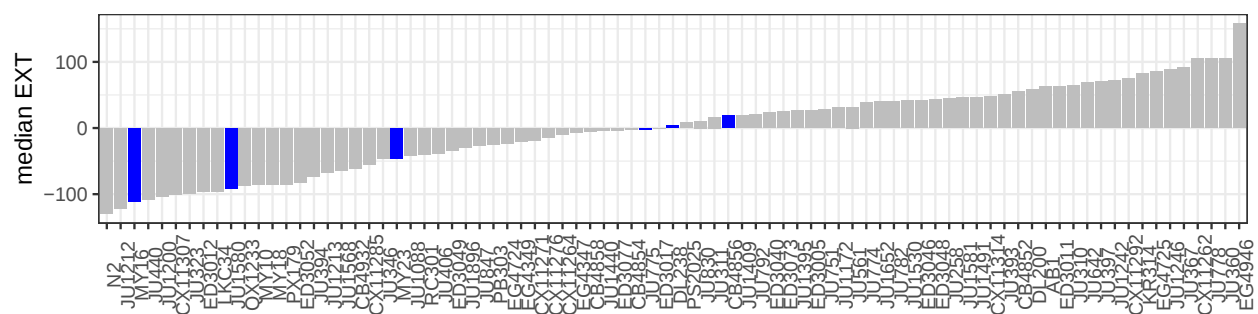
Bar chart showing median EXT values for various samples. The y-axis is labeled 'median EXT' and ranges from -100 to 100. The x-axis lists 48 samples. Most samples have negative median EXT values, with a few positive values. Two samples, ED3027 and CB4836, are highlighted with blue bars.

Sample	median EXT
N2	-110
JU12	-105
MY16	-100
JU440	-95
JU1200	-90
JU1300	-85
ED3027	-80
ED3028	-75
JU1530	-70
JU1233	-65
MY18	-60
ED3029	-55
ED3030	-50
JU1168	-45
CB4932	-40
CB4933	-35
JU1336	-30
JU1038	-25
PC301	-20
JU706	-15
ED3039	-10
JU1896	-5
PC302	0
ED3040	5
ED3041	10
ED3042	15
ED3043	20
ED3044	25
ED3045	30
ED3046	35
ED3047	40
ED3048	45
ED3049	50
ED3050	55
ED3051	60
ED3052	65
ED3053	70
ED3054	75
ED3055	80
ED3056	85
ED3057	90
ED3058	95
ED3059	100
ED3060	105
ED3061	110
ED3062	115
ED3063	120
ED3064	125
ED3065	130
ED3066	135
ED3067	140
ED3068	145
ED3069	150
ED3070	155
ED3071	160
ED3072	165
ED3073	170
ED3074	175
ED3075	180
ED3076	185
ED3077	190
ED3078	195
ED3079	200
ED3080	205
ED3081	210
ED3082	215
ED3083	220
ED3084	225
ED3085	230
ED3086	235
ED3087	240
ED3088	245
ED3089	250
ED3090	255
ED3091	260
ED3092	265
ED3093	270
ED3094	275
ED3095	280
ED3096	285
ED3097	290
ED3098	295
ED3099	300
ED3100	305
ED3101	310
ED3102	315
ED3103	320
ED3104	325
ED3105	330
ED3106	335
ED3107	340
ED3108	345
ED3109	350
ED3110	355
ED3111	360
ED3112	365
ED3113	370
ED3114	375
ED3115	380
ED3116	385
ED3117	390
ED3118	395
ED3119	400
ED3120	405
ED3121	410
ED3122	415
ED3123	420
ED3124	425
ED3125	430
ED3126	435
ED3127	440
ED3128	445
ED3129	450
ED3130	455
ED3131	460
ED3132	465
ED3133	470
ED3134	475
ED3135	480
ED3136	485
ED3137	490
ED3138	495
ED3139	500
ED3140	505
ED3141	510
ED3142	515
ED3143	520
ED3144	525
ED3145	530
ED3146	535
ED3147	540
ED3148	545
ED3149	550
ED3150	555
ED3151	560
ED3152	565
ED3153	570
ED3154	575
ED3155	580
ED3156	585
ED3157	590
ED3158	595
ED3159	600
ED3160	605
ED3161	610
ED3162	615
ED3163	620
ED3164	625
ED3165	630
ED3166	635
ED3167	640
ED3168	645
ED3169	650
ED3170	655
ED3171	660
ED3172	665
ED3173	670
ED3174	675
ED3175	680
ED3176	685
ED3177	690
ED3178	695
ED3179	700
ED3180	705
ED3181	710
ED3182	715
ED3183	720
ED3184	725
ED3185	730
ED3186	735
ED3187	740
ED3188	745

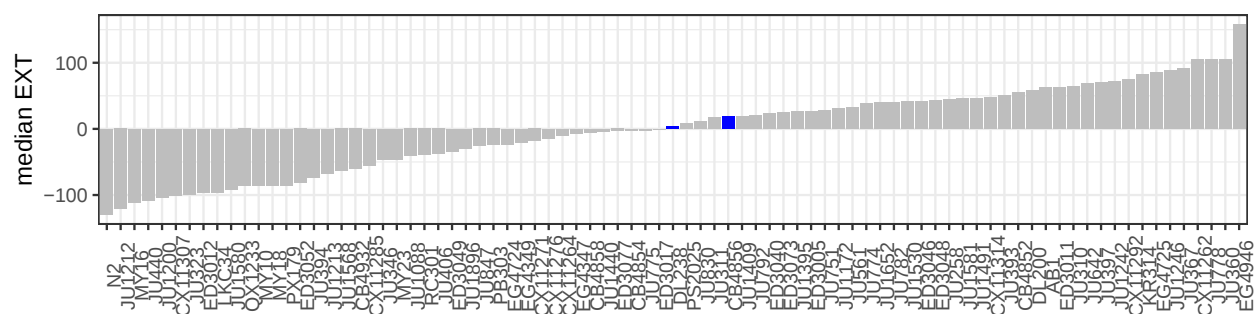
Figure 1 is a bar chart showing the median EXT values for 496 samples. The y-axis is labeled 'median EXT' and ranges from -100 to 100. The x-axis lists 496 sample IDs. Most bars are grey, but two bars (ED3027 and ED3028) are highlighted in blue. The values generally increase from left to right, starting around -120 and ending around 120.

Bar chart showing median EXT values for various taxa. The y-axis is labeled 'median EXT' and ranges from -100 to 100. The x-axis lists taxa names. Most taxa have negative median EXT values, with N2 and MY12 being the most negative. A few taxa have positive median EXT values, including ED3017, ED3018, and ED3019.

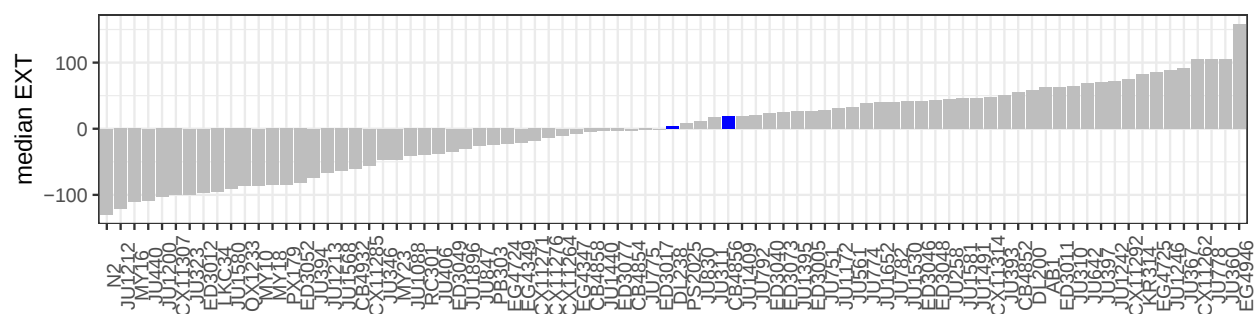
V:11104309



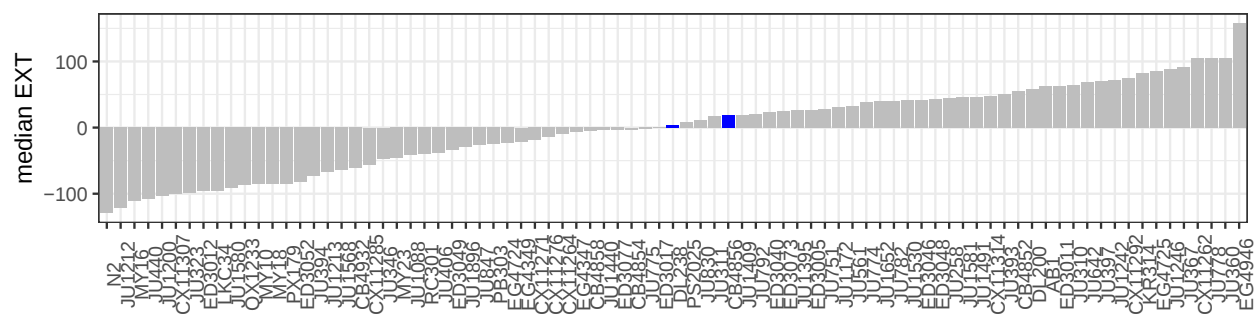
V:11125840



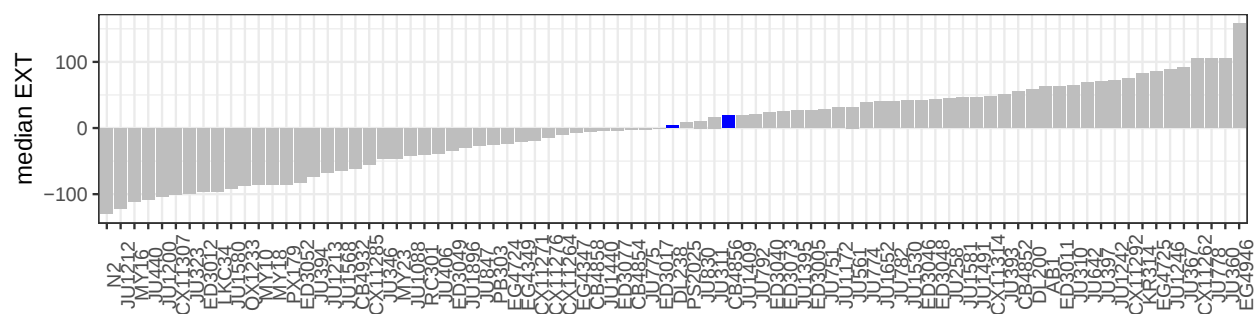
V:11127057



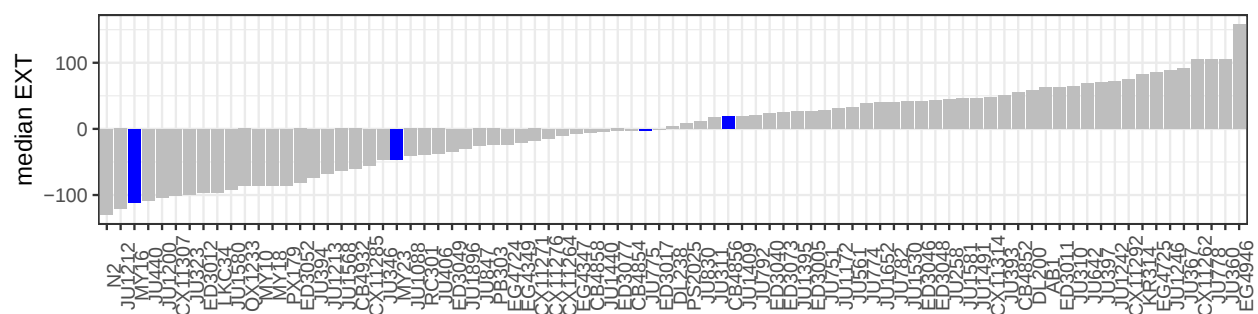
V:11127063



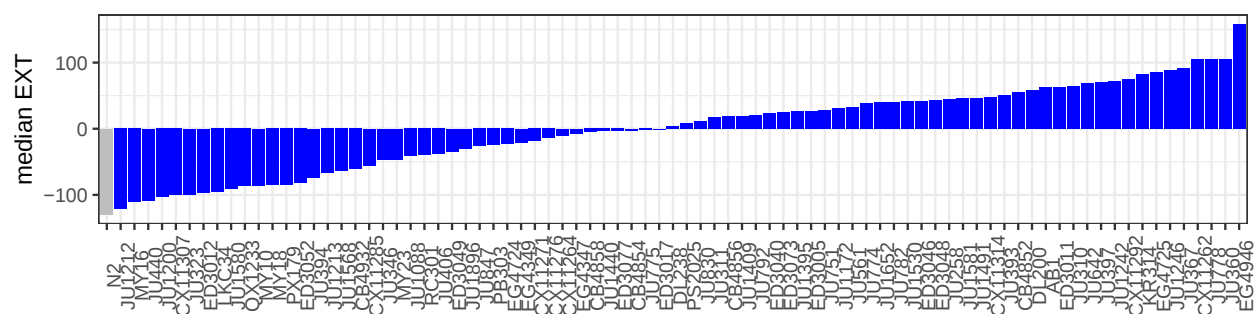
V:11127357



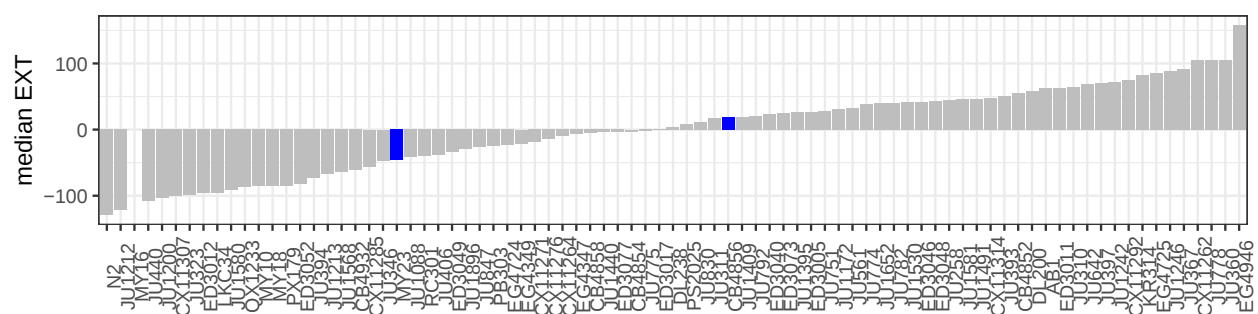
V:11128895



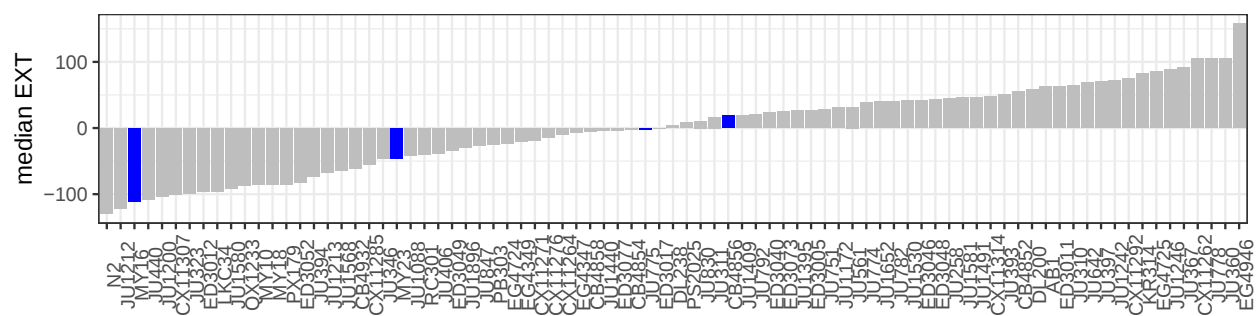
V:11130591



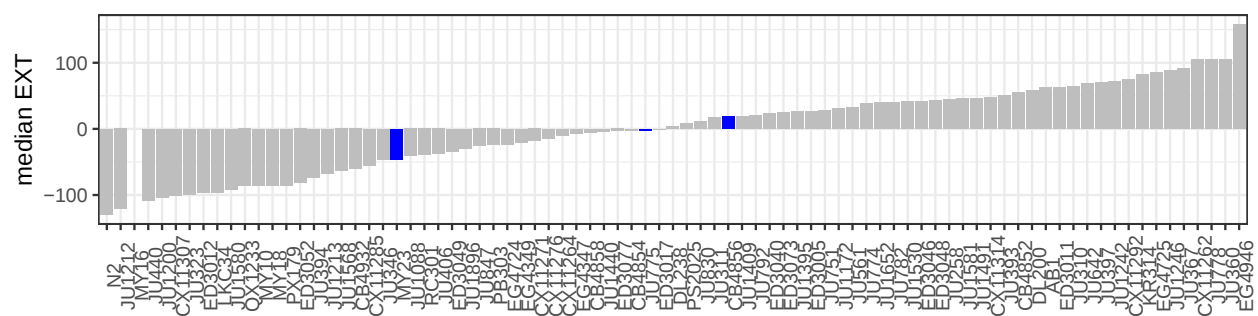
V:11133500



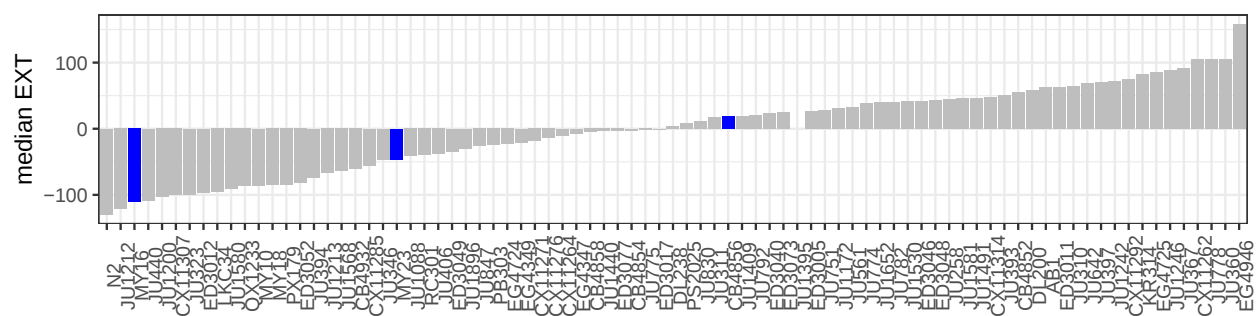
V:11133501



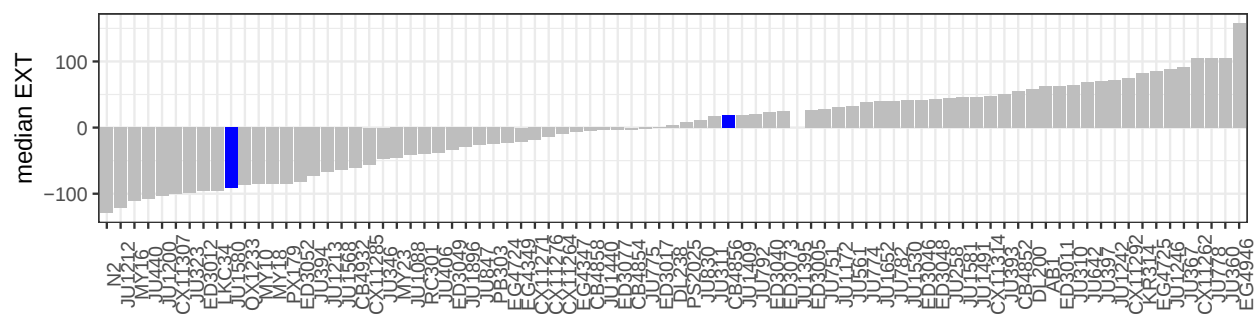
V:11154887



V:11163719



V:11168198



Bar chart showing median EXT values for various taxa. The y-axis is labeled 'median EXT' and ranges from -100 to 100. The x-axis lists taxa names. Most taxa have negative median EXT values, with some highlighted in blue: N2, MY16, ED3027, JX1330, JX1336, and JX1349. The taxa are ordered by their median EXT values, from most negative to most positive.

Bar chart showing median EXT values for various taxa. The y-axis is labeled 'median EXT' and ranges from -100 to 100. The x-axis lists taxa names. Most taxa have negative median EXT values, with some highlighted in blue. A few taxa have positive median EXT values, also highlighted in blue.

Taxon	median EXT (approx.)
N2	-120
MY12	-110
MY16	-100
MY17	-90
MY18	-85
MY19	-80
MY20	-75
MY21	-70
MY22	-65
MY23	-60
MY24	-55
MY25	-50
MY26	-45
MY27	-40
MY28	-35
MY29	-30
MY30	-25
MY31	-20
MY32	-15
MY33	-10
MY34	-5
MY35	0
MY36	5
MY37	10
MY38	15
MY39	20
MY40	25
MY41	30
MY42	35
MY43	40
MY44	45
MY45	50
MY46	55
MY47	60
MY48	65
MY49	70
MY50	75
MY51	80
MY52	85
MY53	90
MY54	95
MY55	100
MY56	105
MY57	110
MY58	115
MY59	120
MY60	125
MY61	130
MY62	135
MY63	140
MY64	145
MY65	150
MY66	155
MY67	160
MY68	165
MY69	170
MY70	175
MY71	180
MY72	185
MY73	190
MY74	195
MY75	200
MY76	205
MY77	210
MY78	215
MY79	220
MY80	225
MY81	230
MY82	235
MY83	240
MY84	245
MY85	250
MY86	255
MY87	260
MY88	265
MY89	270
MY90	275
MY91	280
MY92	285
MY93	290
MY94	295
MY95	300
MY96	305
MY97	310
MY98	315
MY99	320
MY100	325
MY101	330
MY102	335
MY103	340
MY104	345
MY105	350
MY106	355
MY107	360
MY108	365
MY109	370
MY110	375
MY111	380
MY112	385
MY113	390
MY114	395
MY115	400
MY116	405
MY117	410
MY118	415
MY119	420
MY120	425
MY121	430
MY122	435
MY123	440
MY124	445
MY125	450
MY126	455
MY127	460
MY128	465
MY129	470
MY130	475
MY131	480
MY132	485
MY133	490
MY134	495
MY135	500
MY136	505
MY137	510
MY138	515
MY139	520
MY140	525
MY141	530
MY142	535
MY143	540
MY144	545
MY145	550
MY146	555
MY147	560
MY148	565
MY149	570
MY150	575
MY151	580
MY152	585
MY153	590
MY154	595
MY155	600
MY156	605
MY157	610
MY158	615
MY159	620
MY160	625
MY161	630
MY162	635
MY163	640
MY164	645
MY165	650
MY166	655
MY167	660
MY168	665
MY169	670
MY170	675
MY171	680
MY172	685
MY173	690
MY174	695
MY175	700
MY176	705
MY177	710
MY178	715
MY179	720
MY180	725
MY181	730
MY182	735
MY183	740
MY184	745
MY185	750
MY186	755
MY187	760
MY188	765
MY189	770
MY190	775
MY191	780
MY192	785
MY193	790
MY194	795
MY195	800
MY196	805
MY197	8

Bar chart showing median EXT values for various taxa. The y-axis is labeled 'median EXT' and ranges from -100 to 100. The x-axis lists taxa names. Most taxa have negative median EXT values, with a few positive values. Two taxa, LU1333 and CB4856, are highlighted in blue.

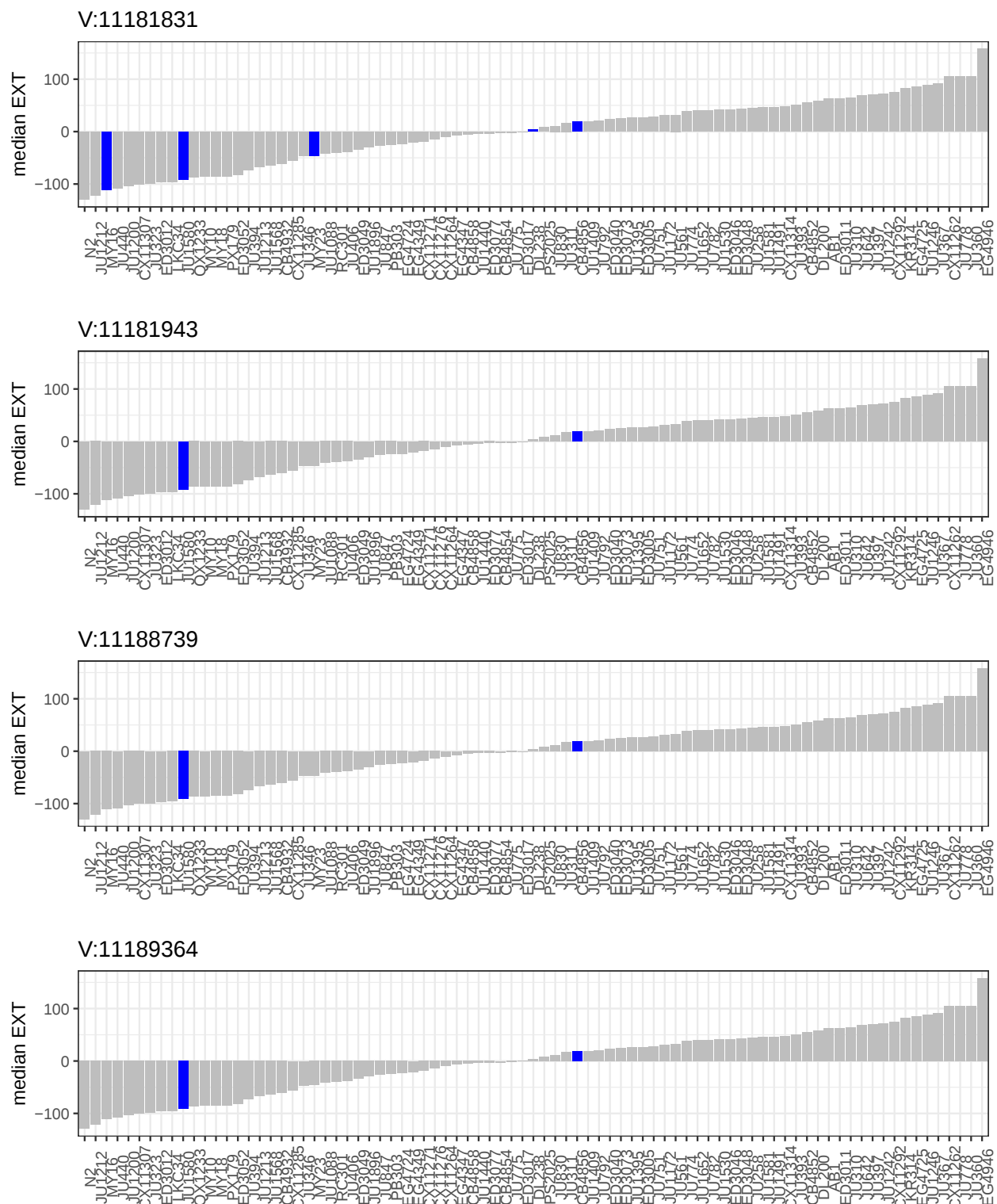


Figure S16

Phenotypes of all wild isolates assayed in bleomycin are shown for each of the 28 variants in the QTL region that are rare (MAF < 0.05) but not unique to CB4856. Each plot shows a bar plot indicating the distribution of residual bleomycin median optical density phenotypes across all 83 wild isolates assayed, arranged by phenotype. The x-axis indicates the strain name, and the y-axis indicates the phenotypic value. For each of

the 28 variants, the position is written above the phenotypic distribution. Strains with the alternate allele at that site are colored blue, and strains with the reference allele are colored in grey.

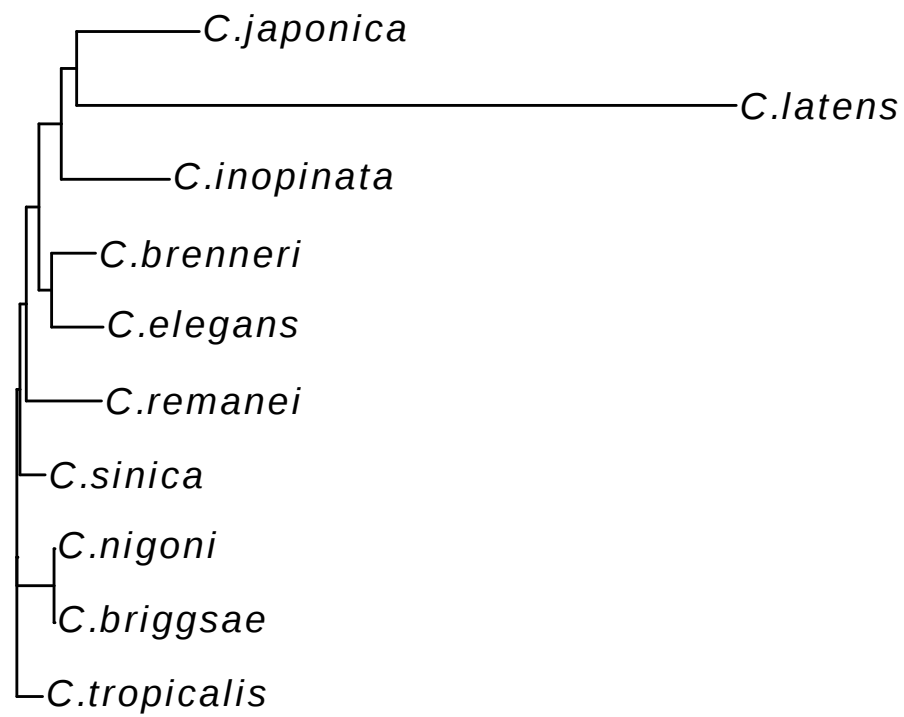


Figure S17

A neighbor-joining tree from from a multiple-sequence alignment of SCB-1 homologs is shown.