

Supplementary Material

**Evolutionary loss of genomic proximity to conserved noncoding elements impacted the gene expression dynamics during mammalian brain development**

Meenakshi Bagadia<sup>§</sup>, Keerthivasan Raanin Chandradoss<sup>§</sup>, Yachna Jain, Harpreet Singh, Mohan Lal, Kuljeet Singh Sandhu\*

Department of Biological Sciences  
Indian Institute of Science Education and Research (IISER) - Mohali  
Knowledge City, Sector - 81, SAS Nagar 140306, India

*<sup>§</sup>Equal contribution*

*\*To whom correspondence should be addressed*

Kuljeet Singh Sandhu

Assistant Professor

Department of Biological Sciences

Indian Institute of Science Education and Research (IISER) - Mohali

E. mail: [sandhuks@iisermohali.ac.in](mailto:sandhuks@iisermohali.ac.in)

## Supplementary Figure Legends

**Figure S1.** A flowchart illustrating overall strategy to obtain syntenic and LOP CNE-gene pairs.

**Figure S2.** Distribution of CNE density per gene (within 1Mb distance) in LOP and syntenic sets in human. P-value was calculated using two-tailed Mann Whitney U test.

**Figure S3.** Enrichment of (a) Mammalian Phenotype Ontology (MPO) terms, and (b) Bgee anatomical terms in syntenic (left panel) and rat-LOP (right panel) gene-sets. P-values were corrected using the Benjamini-Hochberg method.

**Figure S4.** Extension of epigenomic analyses of syntenic and rat-LOP CNEs. (a-b) H3K4me1 ChIP enrichment (over input DNA) around syntenic (black) and rat-LOP (red) CNEs in (a) fetal, and (b) post-natal tissues. (c) The strategy used to assess the CNE-TSS interactions using HiC datasets of human fetal and adult brains. All-to-all HiC interactions were filtered for TSS-to-all interactions for the genes in syntenic and rat-LOP sets. The resultant data was analogous to 4C and was analyzed using method atypical for 4C analysis. Loess regression line was fit to 4C counts as a function of genomic distance from the reference point (TSS in this case). The distance of 3-standard deviation from this regression line was taken as significance cut-off for the interactions impinging onto TSS. (d) Example of CNE-TSS interactions identified in the rat-LOP set. Upper and lower panels represent fetal and adult brain data. Red-line: 4C signal; grey line: H3K4me1; Black line: Loess fit; Dotted line: 3-standard deviation cut-off for significance.

**Figure S5.** Motif enrichment analysis of rat-LOP CNEs. 'Peak-motifs' from RSAT was used to identify over-represented sequence motifs in the rat-LOP CNEs while taking syntenic CNEs as background control. (a) Sequence motifs, their e-values and the matching transcription factors (TFs) from JASPAR. (b) Tissue specificity analyses of TFs. Red bars represent the tissues, wherein the TFs exhibit significant specificity ( $P < 0.05$ ). (c) Time course gene expression of TFs during human brain development. The red curve represents the average expression of TFs and grey color denotes 90% confidence interval.

**Figure S6.** Analysis of distance dependent categories of rat-LOP genes. (a) Enrichment of gene ontology terms (as revealed by 'toppgene' web-server) among genes in three different non-overlapping gene-sets prepared by calculating tertiles of CNE-gene distance distribution in rat-LOP set. (b) Corresponding developmental gene expression trajectories of human and rat brains. Red curve signifies rat-LOP genes, and the black line with 95% confidence intervals (grey) denotes null distribution obtained from rest of the genome. P-values were calculated using one-tailed t-tests for the fetus-specific upregulation of rat-LOP genes as compared to postnatal.

**Figure S7.** Analysis of gain of proximity instances in human. The gain of proximity instances were identified using the strategy shown. No enrichment was found in the GO process terms. The most enriched GO term is shown.

**Figure S8.** Analysis of 'loss of proximity' events that were common to rodents. (a) Enrichment of gene ontology terms among genes in rodent-LOP-2 set (syntenic in human and chimp, but lost proximity in rat and mouse, n=832). Rodent-LOP-1 set (intersection of rat-LOP and mouse-LOP, n=62) did not exhibit significant enrichment of any GO term at 10% FDR. (b) Developmental gene expression patterns of human, rat and mouse brains. Red curve signifies rodent-LOP genes, and black line denotes the null distribution. P-values were calculated using one tailed t-tests for the fetus-specific upregulation of rodent-LOP genes as compared to post-natal.

**Figure S9.** Enrichment of H3K4me1 mark on syntenic and rodent-LOP CNEs. Shown are the fetal-to-adult ratio of normalized H3K4me1 enrichment in +/- 200 bp from the center of the CNEs in rodent-LOP and syntenic sets. Rodent-LOP-1 set is obtained by inducing mouse data onto original rat-LOP data. Rodent-LOP-2 set was obtained by independently comparing CNE-gene pairs in primates (human and chimp) and rodents (rat and mouse). P-values were calculated using two-tailed t-tests.

**Figure S10. Analysis of mutually exclusive 'loss of proximity' events in mouse and rat.** Shown are the developmental gene expression trajectories for the genes that were syntenic in mouse and human but lost proximity in rat (n=125, left panel), and for the genes that were syntenic in rat and human but lost proximity in mouse (n=23, right panel). The correlation between loss of proximity and the fetal brain specific upregulation of genes can be seen. The gene-sets in this analysis were obtained by mapping mouse data onto the main datasets of CNE-gene pairs. P-values were calculated using one tailed t-tests for the fetal brain specific upregulation of LOP genes as compared to postnatal.

**Figure S11.** Comparison of rat-LOP CNE-gene pairs in rn5 and rn6 genome assemblies. (a) Shown is the scatter plot of CNE-gene distances (log10 scale) of the rat-LOP set in rn5 and rn6 assemblies. (b) Gene Ontology enrichment analysis of genes that had lost synteny consistently in both assemblies (83.5% of total). P-values are adjusted using the Benjamini-Hochberg method.

### **Supplementary Data legend**

Spreadsheet tables having raw datasets used in the analyses.

Figure S1

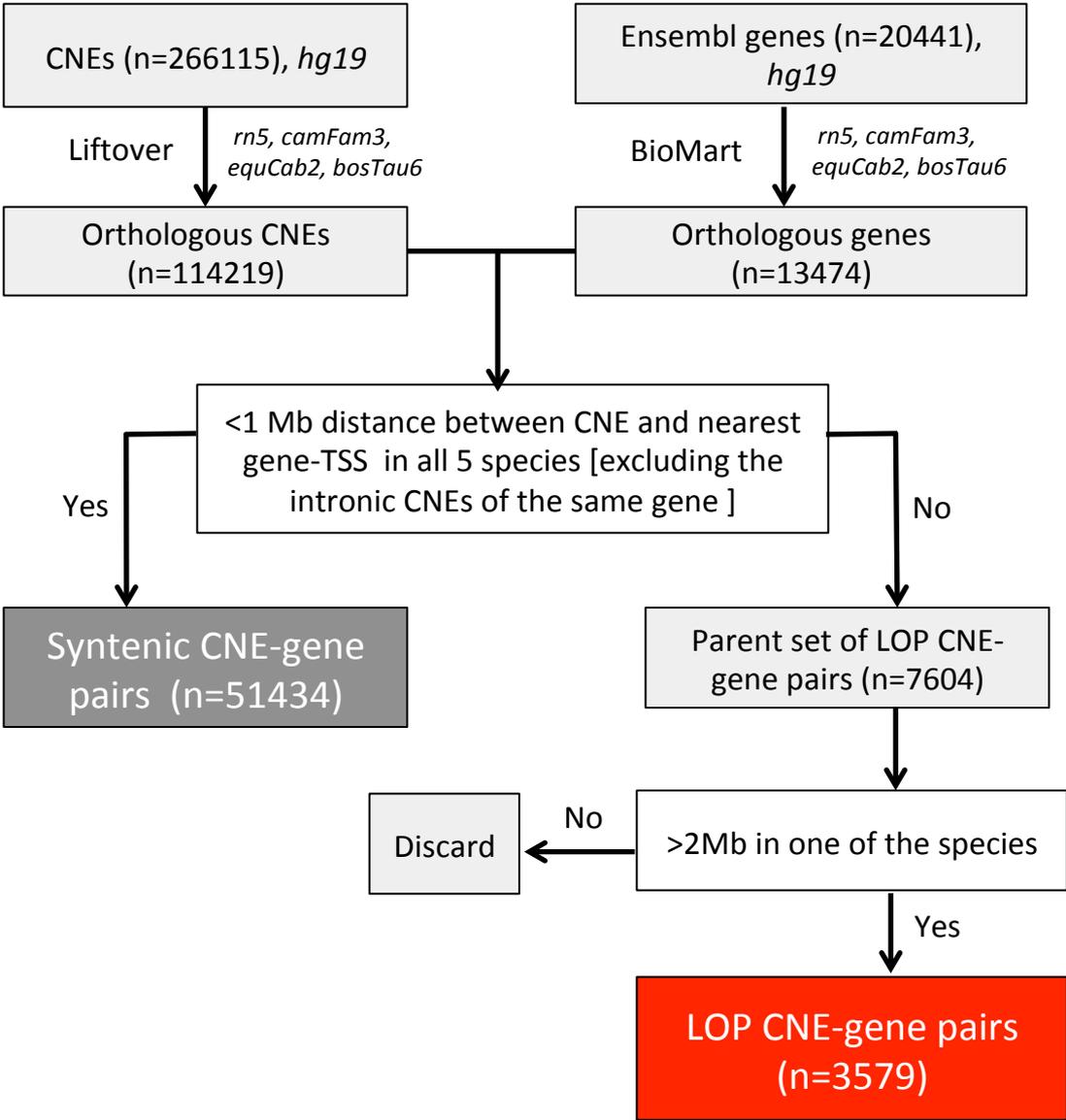


Figure S2

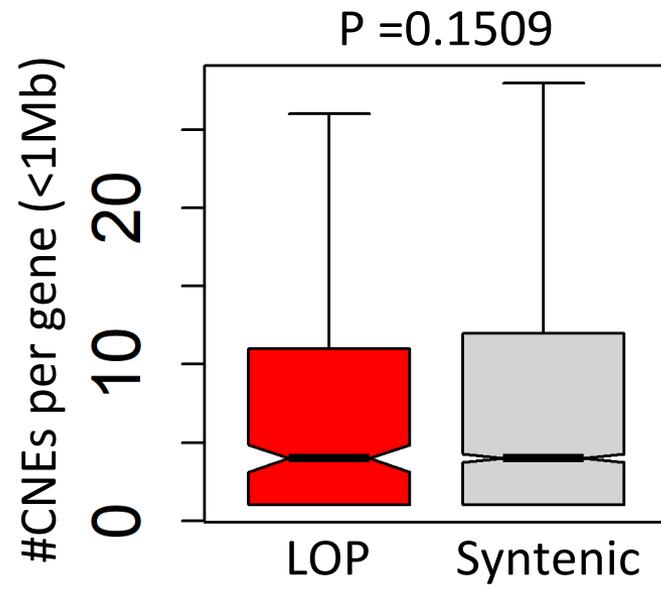
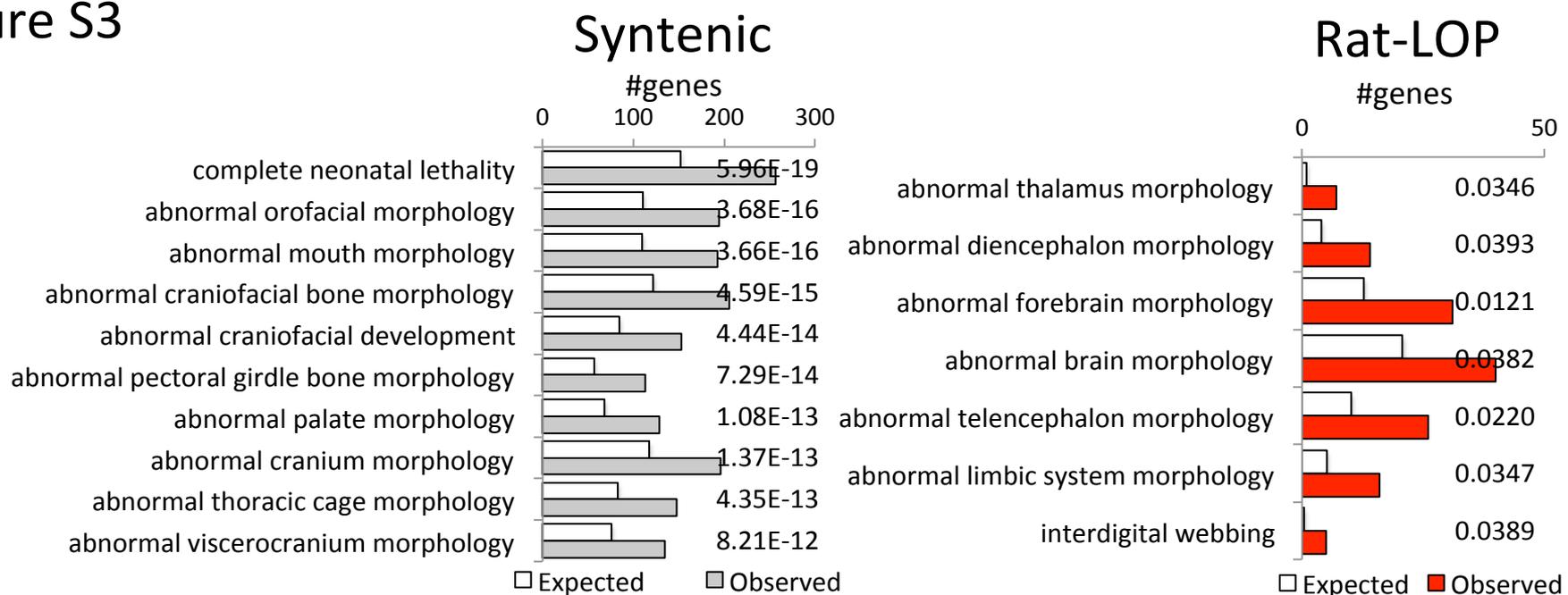


Figure S3

**A**



**B**

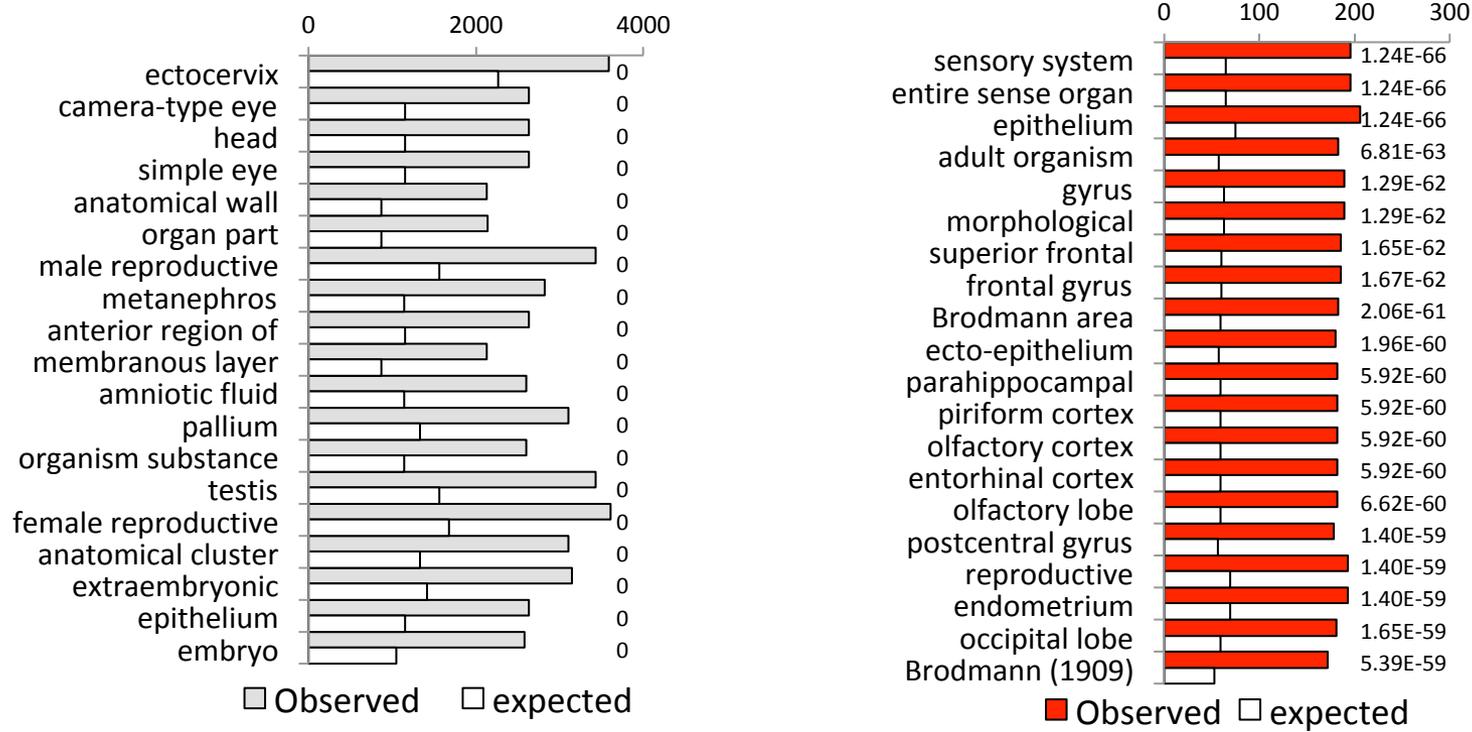


Figure S4

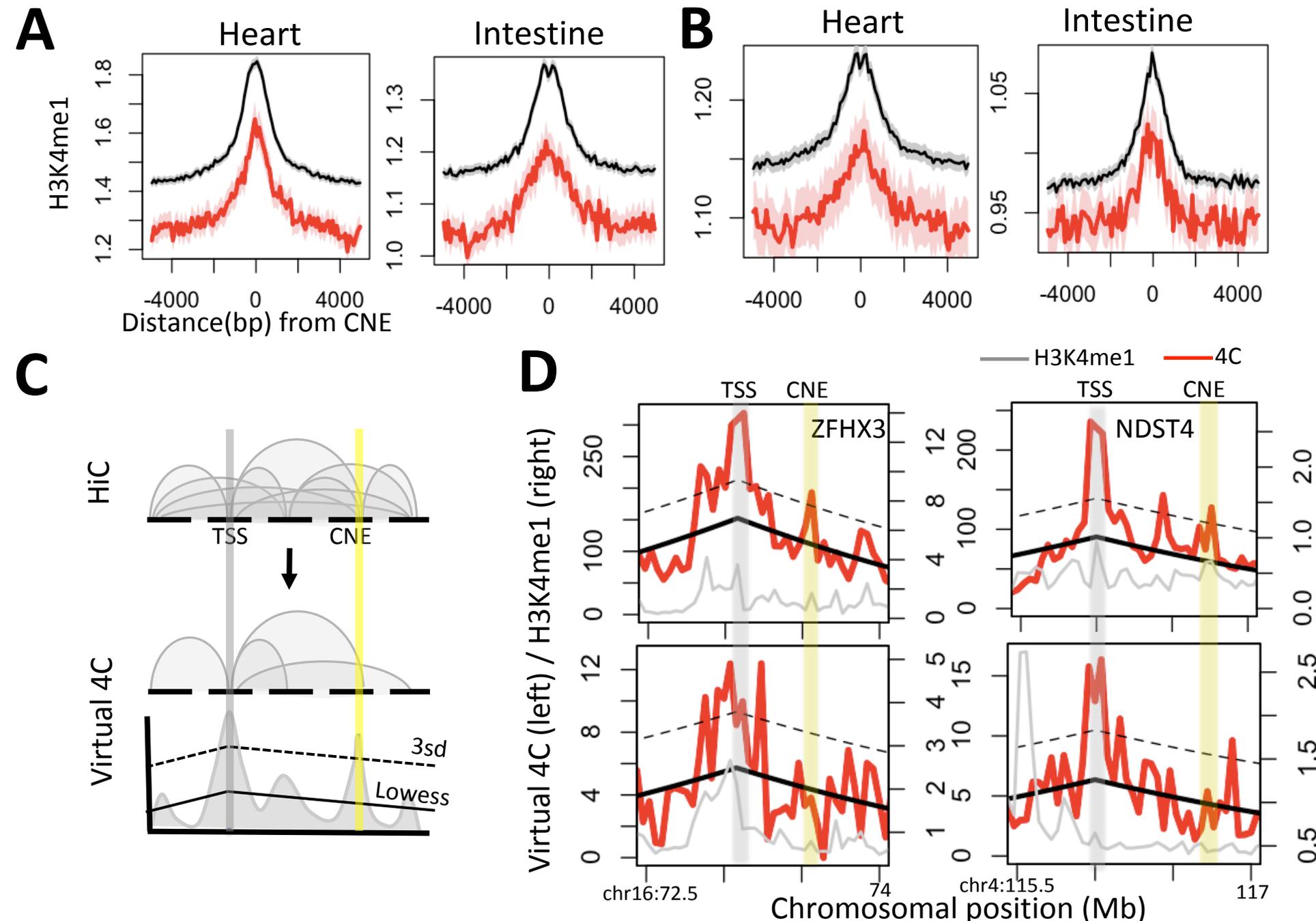
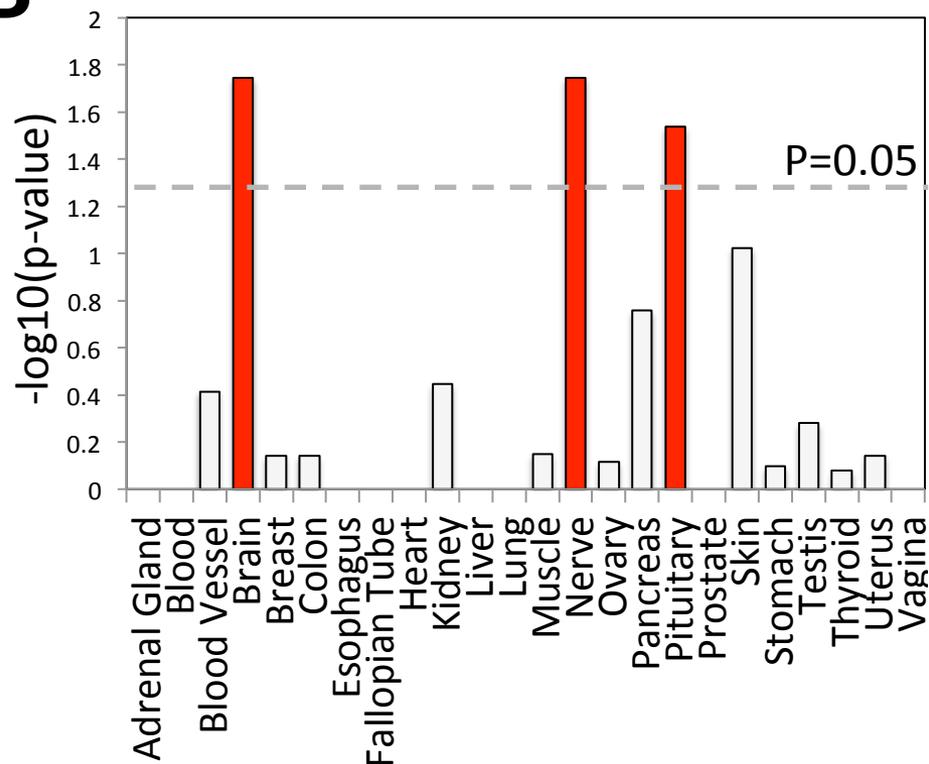


Figure S5

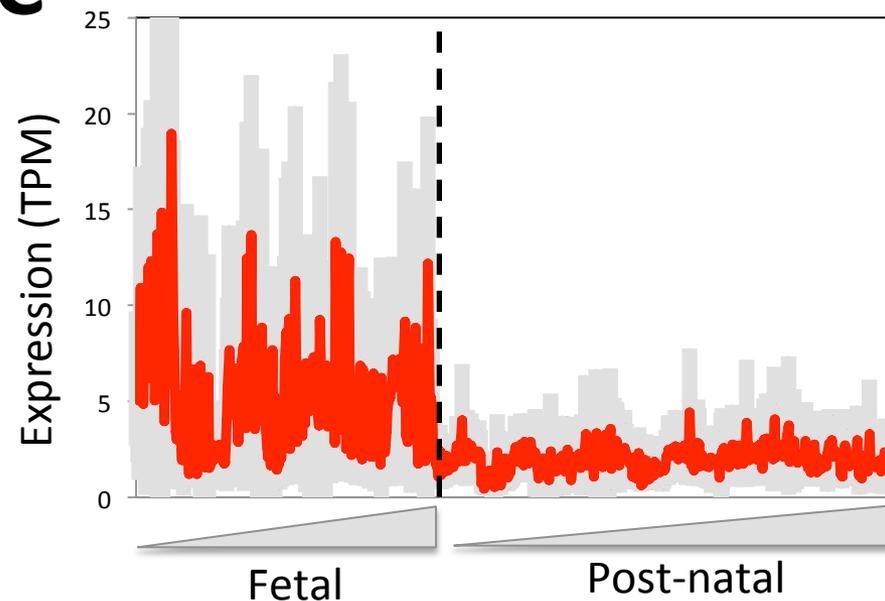
**A**

Motif	E-value	TF match
mdGCAAATTAaww	0.00025	mix_a,RAX,NOTO,HESX1,GSX2,LHX2,ESX1,LBX2,Dlx1,MIXL1,MNX1,Arid3b,POU2F2,Pou2f3,POU1F1,POU3F2,POU3F1,PRRX1,SHOX,UNCX,Shox2,RAX2,Prrx2,LMX1B,ISX,LHX9,MSX2,PAX4,POU3F3,LBX1,Msx3,Nobox,DLX6,Dlx3,VSX2,NKX6_2,VSX1,MSX1,LMX1A,EN1,Dlx4,Lhx4,POU3F4,Dlx2,POU5F1B,NKX6_1,BSX,PDX1,POU5F1,Lhx3,POU2F1,Arid3a,Nkx2_5
datGaTATAATGwa	0.00038	NKX6_2
wwCATTATGwh	0.0019	NKX6_2,NR3C2,POU6F2,POU2F1,POU3F3
wrCAATAAaAw	0.0043	ZNF410,LMX1B,POU2F1,POU3F3
hrmATGCAAat	0.0032	POU5F1,POU2F1,POU5F1B,POU3F4,POU3F3,Ddit3_Cebpa,POU2F2,POU1F1,POU3F1,POU3F2,Pou5f1_Sox2,Pou2f3

**B**



**C**



# Figure S6

## A

1<sup>st</sup> tertile (0-62Kb), #81

GO term	P-value
<b>regulation of neuron differentiation</b>	7.59E-05
phosphatidylinositol dephosphorylation	1.46E-04
<b>regulation of glial cell differentiation</b>	2.13E-04
<b>regulation of nervous system dev</b>	2.51E-04
<b>regulation of neurogenesis</b>	3.63E-04
<b>negative reg of neuron projection dev</b>	4.18E-04
phospholipid dephosphorylation	4.40E-04
long-chain fatty acid metabolic process	5.06E-04
<b>neuron differentiation</b>	5.28E-04
<b>negative regulation of axon regeneration</b>	5.96E-04

2<sup>nd</sup> tertile (62-200Kb), #81

GO term	P-value
cardiac cell fate commitment	1.96E-05
muscle structure development	8.79E-05
<b>central nervous system neuron diff</b>	1.92E-04
muscle cell differentiation	3.25E-04
<b>regulation of neurogenesis</b>	4.07E-04
specification of animal organ identity	4.57E-04
myoblast differentiation	4.87E-04
cell fate specification	5.31E-04
<b>cranial suture morphogenesis</b>	7.63E-04
<b>central nervous system development</b>	8.95E-04

3<sup>rd</sup> tertile (200-1000Kb), #82

GO term	P-value
<b>positive regulation of synapse assembly</b>	1.24E-05
<b>regulation of synapse assembly</b>	4.06E-05
<b>synapse assembly</b>	4.52E-05
<b>regulation of grooming behavior</b>	1.85E-04
<b>regulation of synapse organization</b>	2.22E-04
tricarboxylic acid cycle	2.67E-04
peptidyl-tyrosine phosphorylation	2.68E-04
peptidyl-tyrosine modification	2.77E-04
type II pneumocyte differentiation	3.87E-04
citrate metabolic process	3.94E-04

## B

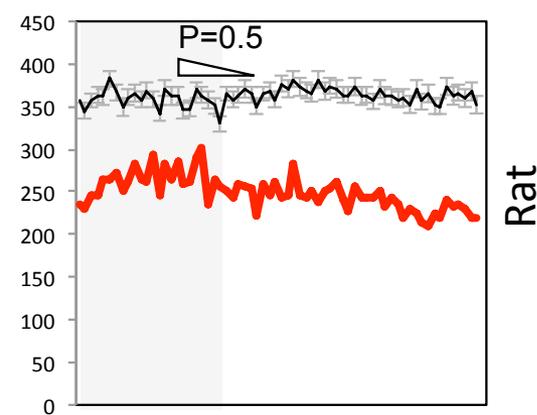
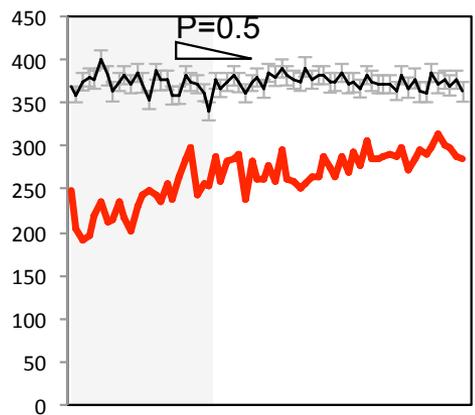
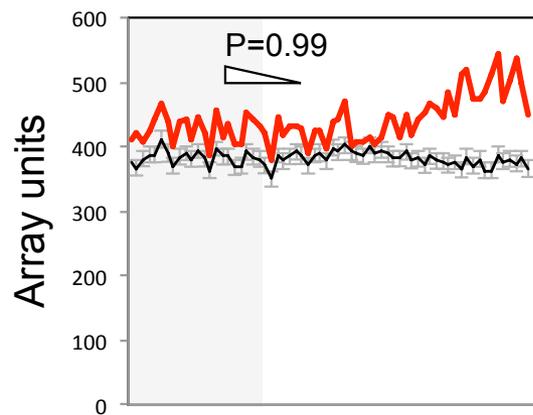
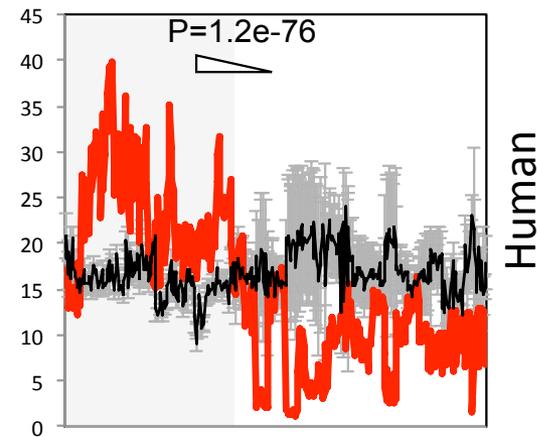
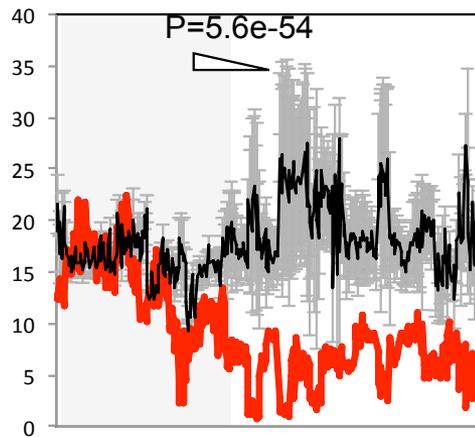
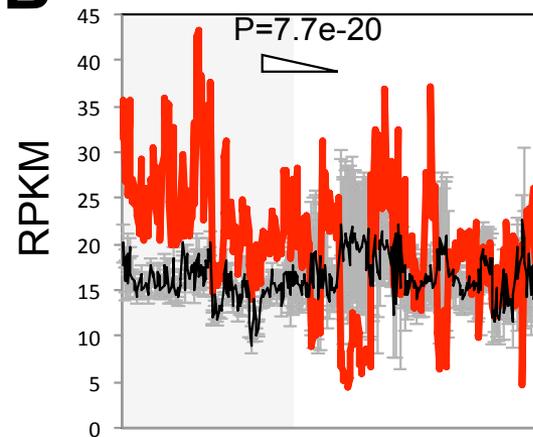
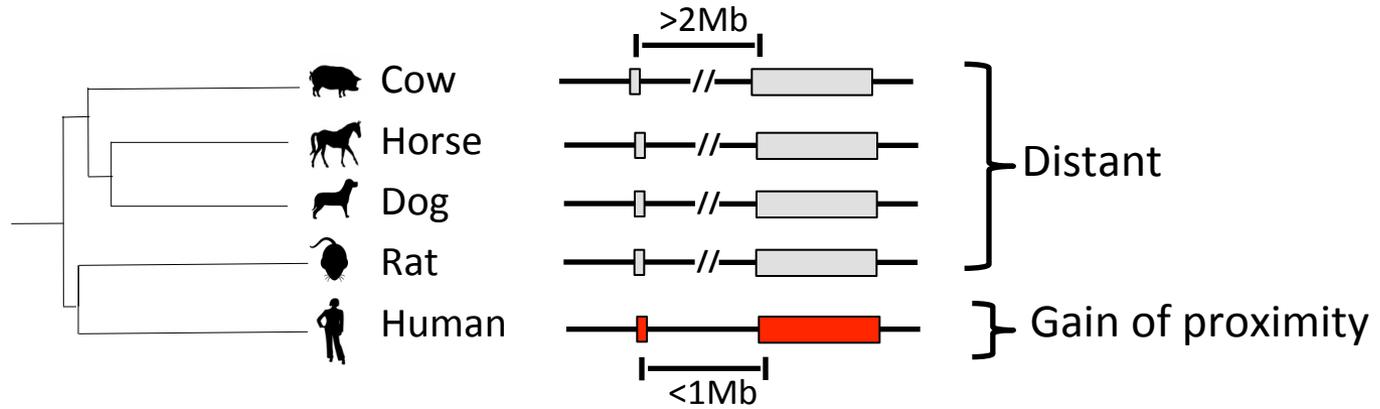


Figure S7



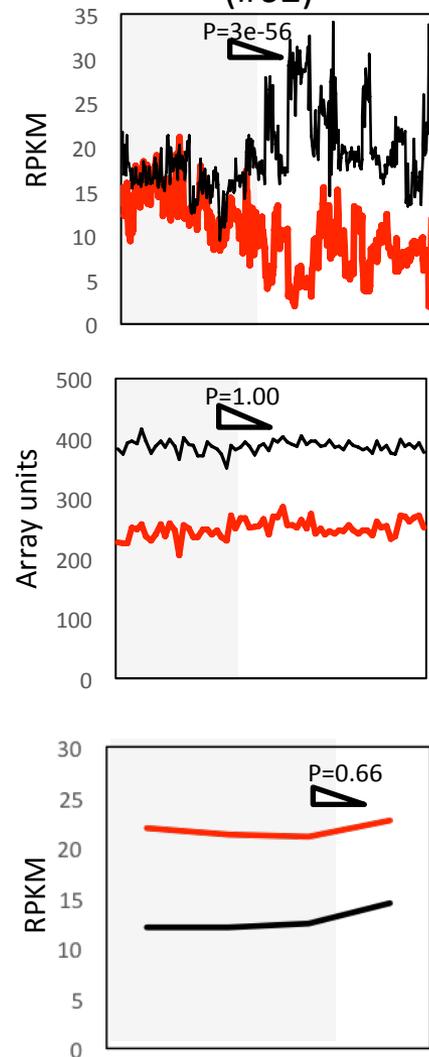
Genes	GO Term Name	Expected	Observed	Obs %	Fold change	FDR
388	G-protein coupled receptor signaling pathway	22.86409	41	10.5%	1.793205	1

# Figure S8

## A Rodent-LOP-2 [vs. syn in primates] (#832)

GO term	P-value	FDR
negative regulation of cell migration	1.74E-05	6.92E-02
negative regulation of cell motility	3.92E-05	6.92E-02
regulation of cell motility	4.85E-05	6.92E-02
regulation of cell migration	5.13E-05	6.92E-02
response to hormone	6.06E-05	6.92E-02
regulation of stress fiber assembly	8.21E-05	6.92E-02
Reg. of actin filament polymerization	8.45E-05	6.92E-02
Reg. of actin filament bundle assembly	8.88E-05	6.92E-02
actin filament polymerization	1.05E-04	7.28E-02
Reg. of cellular component movement	1.39E-04	7.85E-02

## B Rodent-LOP-1 [vs syn in other 4] (#62)



## Rodent-LOP-2 [vs. syn in primates] (#832)

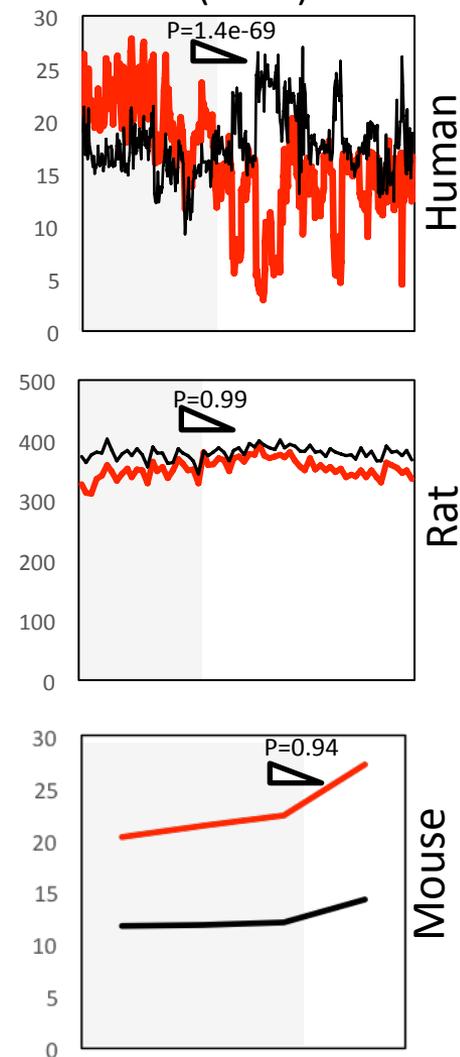


Figure S9

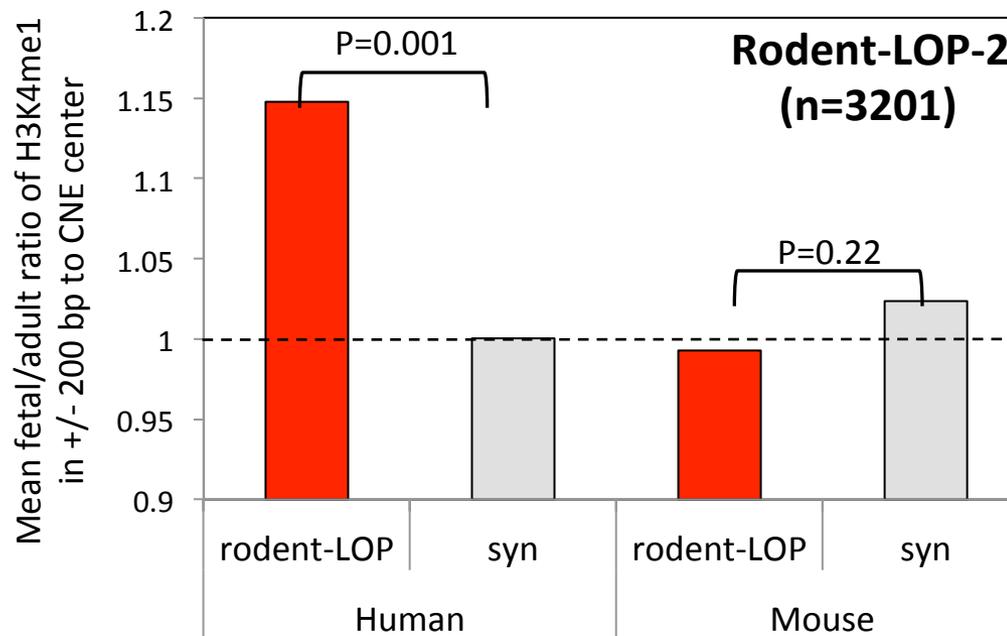
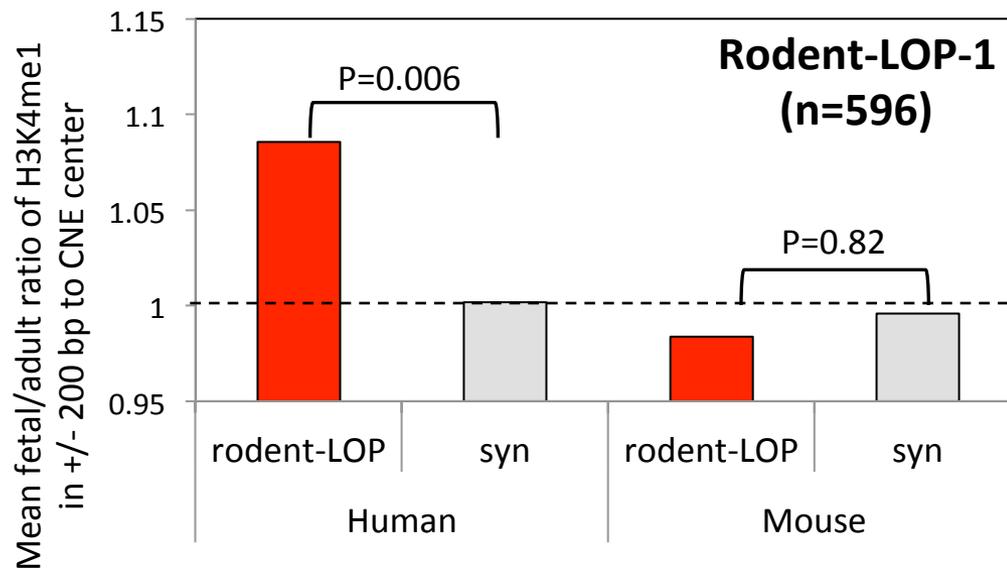


Figure S10

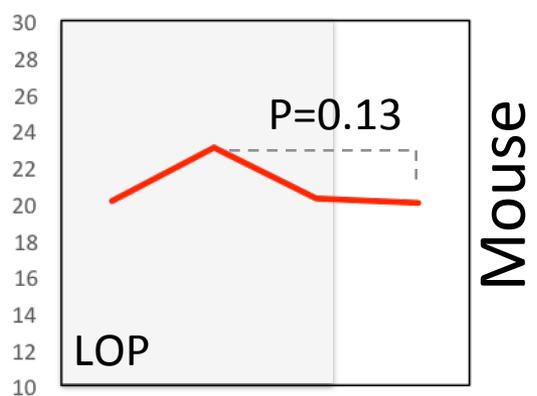
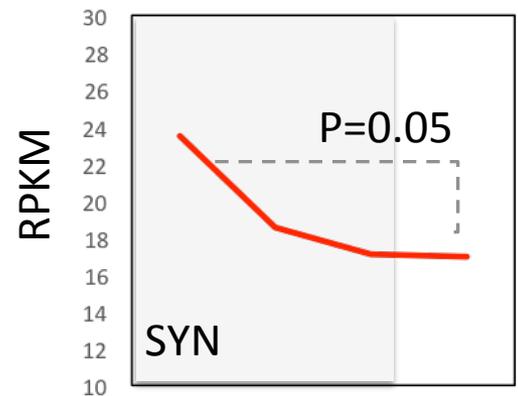
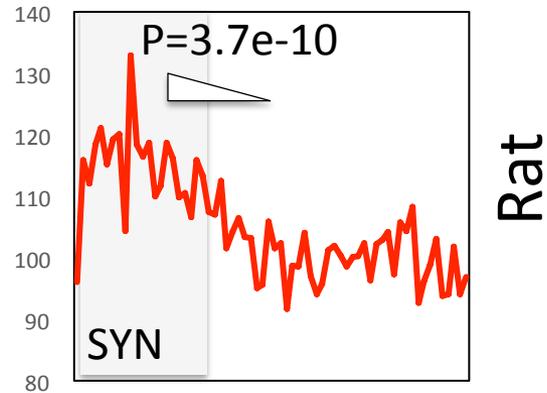
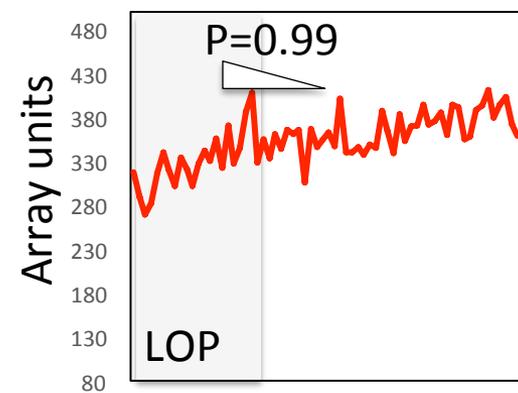
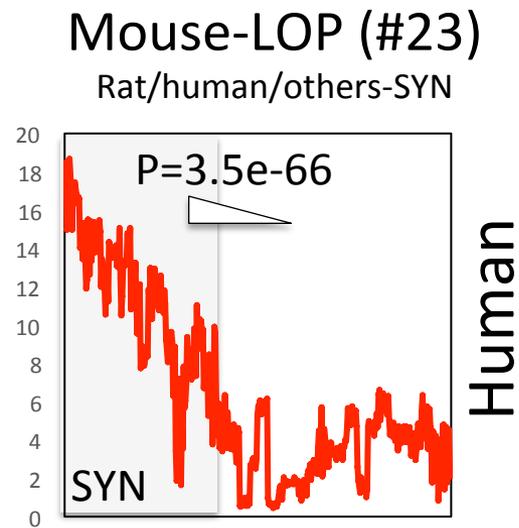
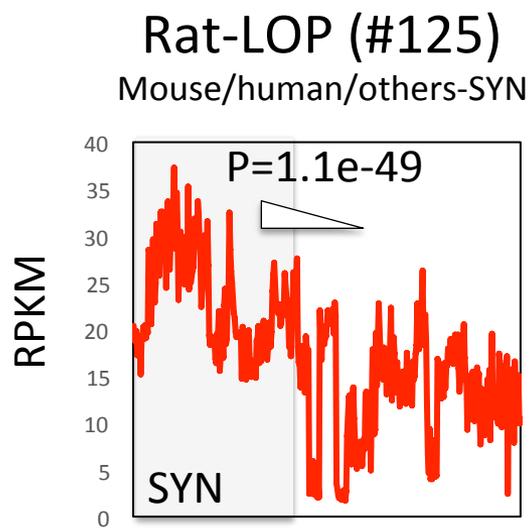
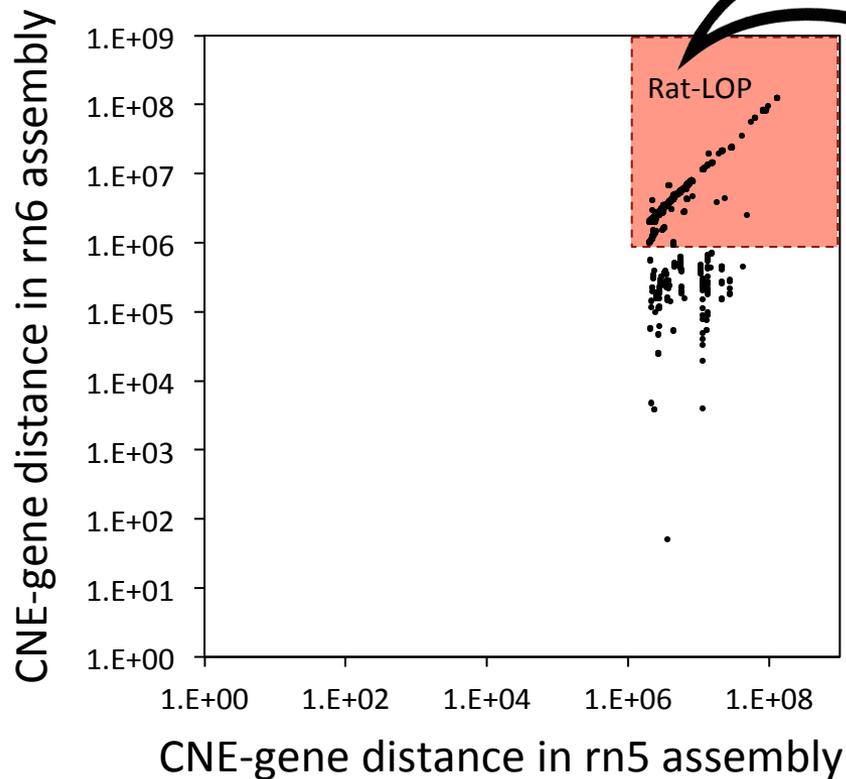


Figure S11

**A**



**B**

