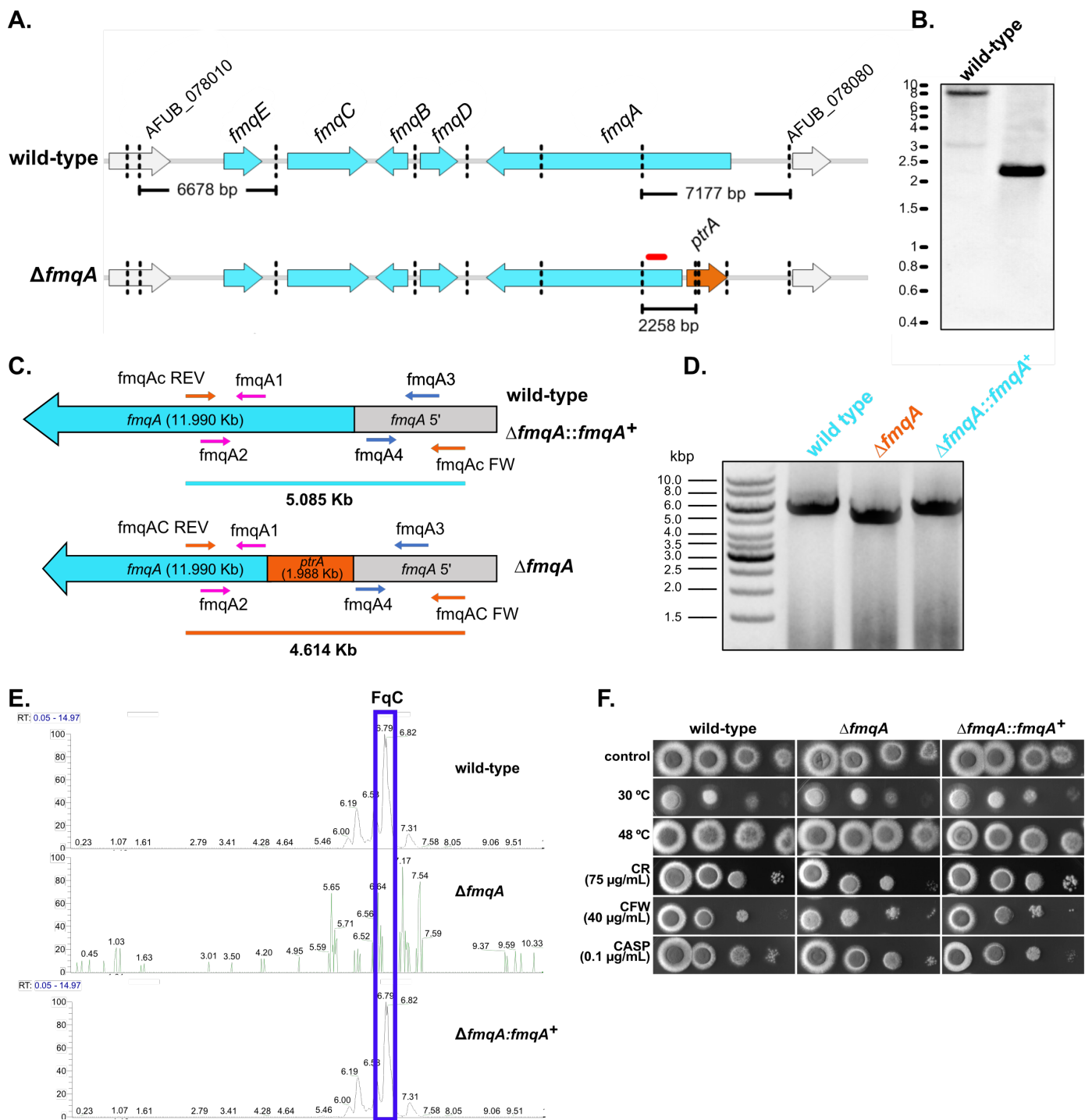
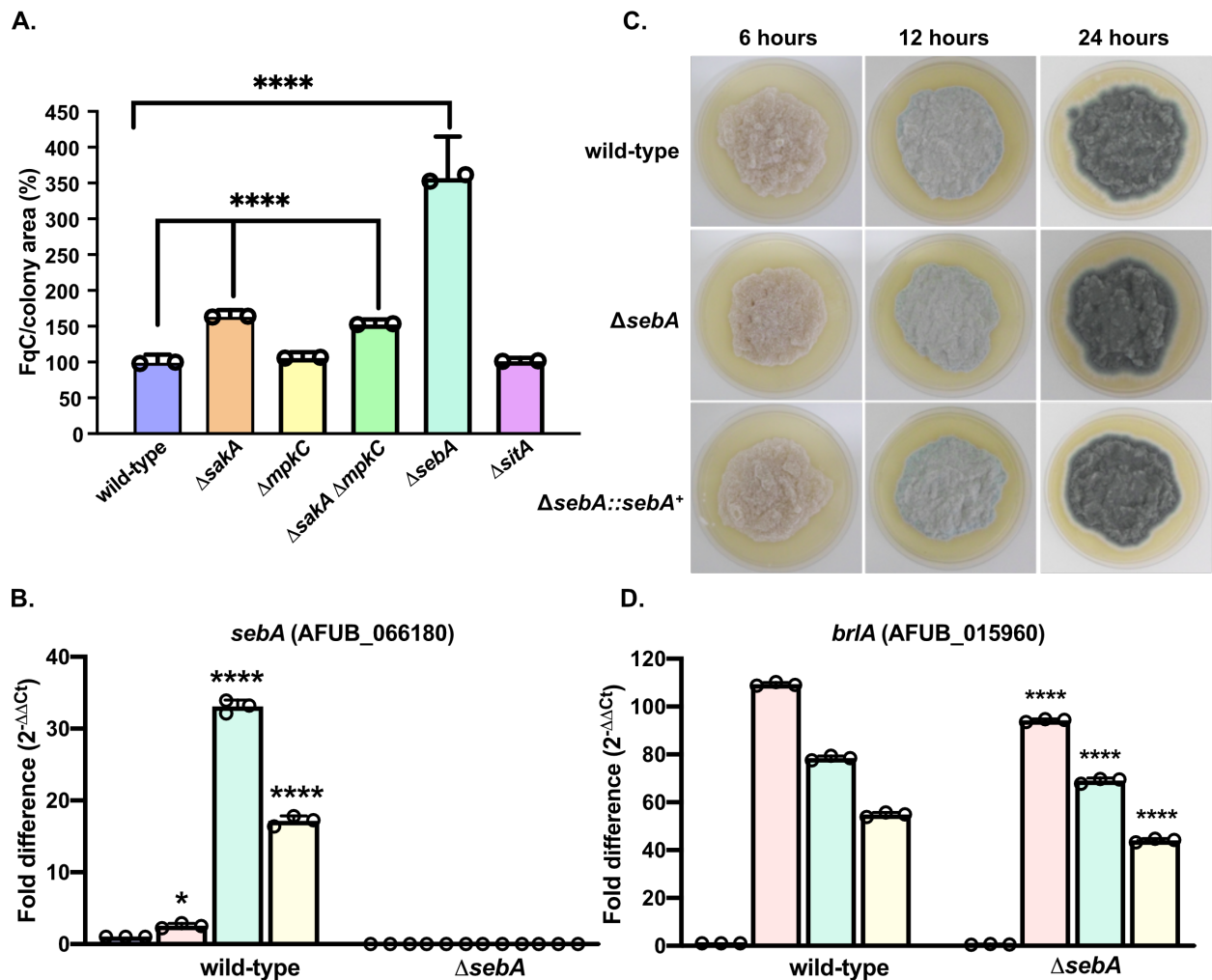


**Figure S1 Graphical representation of the mutated loci for the tagged *A. fumigatus* mutants.** (A) Radial growth of the tagged *mpkA::3×HA fmqA::GFP*, *fmqC::GFP* or *fmqD::GFP* and  $\Delta mpkA fmqC::GFP$  strains.  $1 \times 10^5$  conidia of each strain were inoculated on solid YG medium at the indicated temperature and in the presence of cell wall stressors congo red (CR) and calcofluor white (CFW) at the indicated concentrations. Plates were incubated for 72 hours at 37°C (or the indicated temperature) and photographed. (B) Asexual differentiation of the tagged mutants employed in this study. With the exception of  $\Delta mpkA$  background, for which delayed conidiation is reported previously, no defects in conidiation rate were observed for the other tagged strains. (C) The *mpkA::3×HA::ptrA* cassette was amplified and used to transform each *fmqA-C-D::GFP* recipient strain. (C) The pyrithiamine-resistant transformants were selected and validated by PCR with primers *mpkA* 5F and linker 3×HA REV to confirm the *mpkA* locus replacement by the amplification of the 5.954 kb band in the double-tagged strains. (D) The  $\Delta mpkA$  deletion cassette was amplified from the genomic DNA of the  $\Delta mpkA$  strain using primers *mpkA* 5F and *mpkA* 3' REV and transformed into TFYL50.1 strain to generate the  $\Delta mpkA fmqC::GFP$  mutant. (F) The *mpkA* deletion cassette contains the *ptrA* gene as a selectable marker and the gene replacement in this strain mutants was confirmed by PCR using the primers *mpkA* 600 ups and *mpkA* 3' REV, which can be used to discriminate the *mpkA* deletion from the wild-type *mpkA* locus by the amplification of the 5.6 kb or 5.2 kb band, respectively.



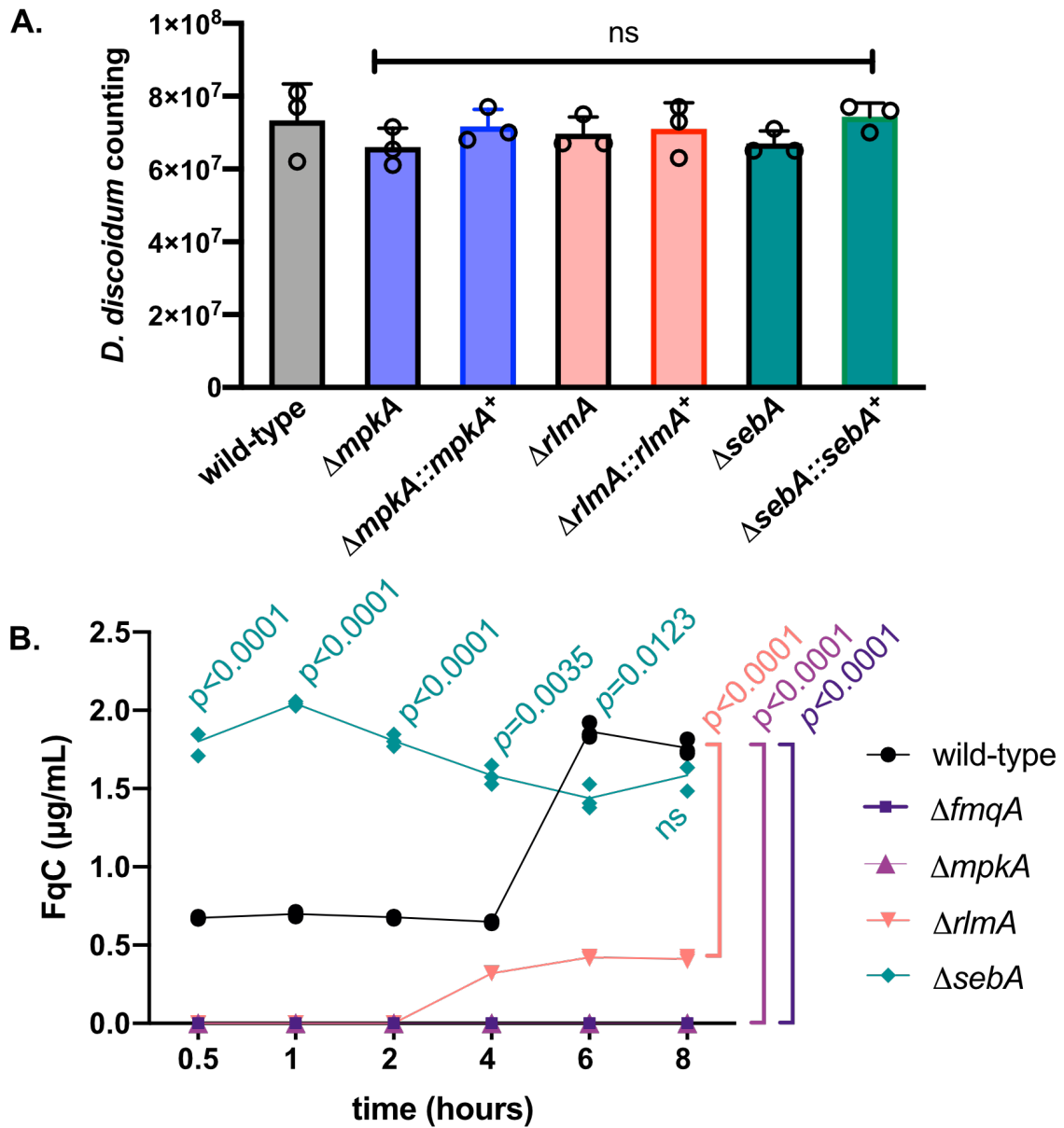
**Figure S2 Graphical representation of the mutated *fmqA* loci and FqC analysis.** (A) Graphical representation of the mutated *fmqA* locus and respective Southern blot. The genes of the Fq biosynthetic cluster are shown in sky-blue. The cassette used for *fmqA* gene disruption is the orange labeled pyrithiamine (*ptrA*). The probe used for Southern blot analyses is marked as red stripes. (B) The DNA ladder used is indicated (Hyperladder 1Kb, Bioline). The Southern blot was performed by digesting genomic DNA with *Xba*I (dashed bands). (C-D) To complement the  $\Delta fmqA$  strain a co-transformation strategy was employed using the 2495 bp gel-purified hygromycin resistant cassette (*gpdA(p)::hph*; not shown) and the *fmqA* coding sequence, which was amplified using the *fmqAc* FW and *fmqAc* REV oligonucleotides. Complementation was validated by PCR using the primers *fmqAc* FW and *fmqAc* REV which yield a band of 5.085 bp after reconstruction of the wild-type *fmqA* locus in the deletion strain. (E) Chromatograms indicating that FqC production is abolished in the  $\Delta fmqA$  strain and fully restored in the complemented  $\Delta fmqA::fmqA^+$  strain. FqC was quantified in the mutant strains by HPLC-MS/MS. (F)  $\Delta fmqA$  mutant presents wild-type levels of susceptibility to cell wall-disturbing compounds. The indicated number of conidia was inoculated onto solid YG plates that were supplemented with the following different cell wall perturbing agents: congo red (CR), calcofluor white (CFW) and caspofungin (CASP) and incubated at 37°C (or the indicated temperature) for 24 hours.



**Figure S3 Selected results for the quantification of FqC in a collection of mutant strains and expression levels of *sebA* and *brlA* during the asexual differentiation.**

(A) Quantification of FqC in mutant strains from the screen of an arbitrary collection of selected *A. fumigatus* mutant strains carrying deletions on kinases, phosphatases and transcription factors involved in different signaling pathways. FqC was quantified by HPLC-MS/MS,  $n=2 \pm SD$ . One-way ANOVA with Tukey's multiple comparison test relative to wild-type was performed. \*\*\*\* $p<0.0001$ . (B) Induction of asexual differentiation of the wild-type,  $\Delta sebA$  and the complemented strain shows no conidiation defects. The experiment was performed at 37°C and images obtained at the indicated time points. (C-D) Gene expression measured by RT-qPCR of *sebA* and *brlA* in the wild-type and  $\Delta sebA$  strains subjected to synchronized asexual differentiation shown in B. Control

indicates the hyphal state (submerged culture).  $n=3 \pm \text{SD}$ . One-way ANOVA with Sidak's multiple comparison test relative to wild-type at the same time point of differentiation was performed. \*\*\*\* $p<0.0001$ ; \* $p=0.0348$ .



**Figure S4 Survival of *D. discoideum* after confrontation with the mutant strains and quantification of FqC release in the culture medium.** (A) Viability of *D. discoideum* cells determined 24 h after the confrontation with the wild-type,  $\Delta mpkA$ ,  $\Delta rlmA$ , and  $\Delta sebA$  conidia. *A. fumigatus* conidia were coincubated with *D. discoideum* at a MOI of 10:1. After 24 h at 22°C, viable amoebae cells were counted using a hemocytometer and plotted in the graph. One-way ANOVA with Dunnett's multiple comparison test was performed in comparison to the control condition ( $n=3 \pm SD$ ). (B) FqC accumulated in *A. fumigatus* conidia is released into the culture medium.  $2 \times 10^8$  conidia of each strain were harvested from plates grown for five days (37°C) and immediately inoculated into 20 mL of liquid

YG. Conidia suspension was incubated at 37°C (80 rpm). Samples (1 mL) were collected at the intervals shown in the graphs starting after 30 min of incubation. The sample volume was centrifuged at maximum speed, filtered (0.22 µm) and subjected to HPLC-MS/MS analysis and quantification, as described in the materials and methods section ( $n=3 \pm \text{SD}$ ). Two-way ANOVA with Tukey's multiple comparisons test was performed.  $p$ -values shown at each time point refer to the comparison to the wild-type strain. ns = non-significant.