

Primer Number	Primer Name	Sequence (5'-3')	Description
7	TC290-2	TGCTCTTCGCGCTGGCAGACATACTGTCCCAC	CmYLCV-F forward primer, type IIS site is highlighted in turquoise and overhang is underlined
24	TC219:GFPKIN-2	TCGTCTCCGTAATGTTAATGCTGCCTATACGGCAGTGAACCTG	gRNA1-R reverse primer, type IIS site is highlighted in turquoise, gRNA spacer sequence green and overhang underlined
25	TC220:GFPKIN-2	TCGTCTCATTACCATTAAACGTTTTAGAGCTAGAAATAGC	gRNA1-F reverse primer, type IIS site is highlighted in turquoise, gRNA spacer sequence green and overhang underlined
26	TC221:GFPKIN-2	TCGTCTCCATTGGTGTCCCTCTGCCTATACGGCAGTGAAC	gRNA2-R reverse primer, type IIS site is highlighted in turquoise, gRNA spacer sequence green and overhang underlined
27	TC222:GFPKIN-2	TCGTCTCACAATGTCTGCTGTTTTAGAGCTAGAAATAGC	gRNA2-F reverse primer, type IIS site is highlighted in turquoise, gRNA spacer sequence green and overhang underlined
12	TC222-2	TGCTCTTCTGACCTGCCTATACGGCAGTGAAC	35Sterm-R reverse primer, type IIS site is highlighted in turquoise and overhang is underlined
38	Bael-LHA-F1	CGCGTAGTCCTCGGTAGGCAAGCTTATTTAATTC	forward primer for the amplification of LHA flanked by 16bp of pMOD_C0000 for Gibson Assembly (blue)
39	Bael-LHA-R1	GATTAGTCTTAACCTTGTCTGCAGG	reverse primer for the amplification of LHA flanked by 10bp of pMOD_C0000 for Gibson Assembly (blue)
40	Bael-GFP-F2	AGACAAAGTTAAGACTAATCTTTTCTCTTTCTCA	forward primer for the amplification of GFP flanked by 10bp of pMOD_C0000 for Gibson Assembly (blue)
41	Bael-GFP-R2	GTTAATGTTATCAAAGCTCATCATGTTT	reverse primer for the amplification of GFP flanked by 10bp of pMOD_C0000 for Gibson Assembly (blue)
42	Bael-RHA-F3	TGAGCTTTGATAACATTAACTTACCATTAACTGTA	forward primer for the amplification of RHA flanked by 10bp of pMOD_C0000 for Gibson Assembly (blue)
43	Bael-RHA-R3	TGACTTGAAGTACACTCCGCTCTAGAATTAGTTG	reverse primer for the amplification of LHA flanked by 17bp of pMOD_C0000 for Gibson Assembly (blue)
173	17p-F	TCGAGCTCGGTACCCACCCTACTTAAAAACCCTTTCG	forward primer for the amplification of Manes.17G095200 flanked by 15bp of pTRANS_220d for In-fusion cloning (blue)
174	17p-R	ACGAATGGGATCCATGGTGACCTCTGGATTTCAGTACGAGG	reverse primer for the amplification of Manes.17G095200 flanked by 15bp of TAL20 product for In-fusion cloning (blue)
175	TAL20-F	ATGGATCCCATTCGTCCGCGCACGCCAAG	forward primer for the amplification of TAL20. First 15bp are included in Manes.17G095200 for In-fusion cloning (blue)
176	TAL20-R	GGCGCGCCGATCCCCCACTGAGGAAATAGCT	reverse primer for the amplification of TAL20 flanked by 15bp of pTRANS_220d for In-fusion cloning (blue)
171	SWEET10a_g-F	CTTCGCTACGCGCTTCTTCATTTACTTTATATGCTAACTACTGACAAATACTG	forward primer used for genotyping products of HDR (Figure 2)
10	SWEET10aPro-R	CATAACCAAGAACACGTTAATGGTAATGTTAATG	reverse primer used for genotyping products of HDR (Figure 2)
37	Actin-F	ACAGTGTCTGGATCGGAGGATC	forward primer for Actin RT-PCR, 28 cycles
KV38	Actin-R	GAAGCACTTCCTGTGGACGATG	reverse primer for Actin RT-PCR, 28 cycles
200	10a_qPCR-F	CTTACTGTGTACCTTATCTATGCCACAAAGAAG	forward primer for MeSWEET10a and MeSWEET10a-GFP RT-PCR, 30 cycles
201	10a_qPCR-R	GCCATGTGTAAGGAAAAGAGTTAAGATAGCG	reverse primer for MeSWEET10a RT-PCR, 30 cycles
172	GFP-R2	CCAGTGAAAAGTCTTCTCTTTACTGAATTCGGCCG	reverse primer for MeSWEET10a-GFP RT-PCR, 30 cycles
164	GFP-R	CGTTGTGGGAGTTGTAGTTGTA	reverse primer for MeSWEET10a-GFP genotyping lines 12 and 42 (Figure S2)