

# Parameter estimates from model Markov chains

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```
R version 3.5.3 (2019-03-11)
Platform: x86_64-pc-linux-gnu (64-bit)
Running under: Arch Linux

Matrix products: default
BLAS/LAPACK: /opt/intel/compilers_and_libraries_2019.1.144/linux/mkl/lib/intel64_lin/libmkl_gf_lp64.so

locale:
 [1] LC_CTYPE=en_US.UTF-8      LC_NUMERIC=C               LC_TIME=en_US.UTF-8
 [4] LC_COLLATE=en_US.UTF-8    LC_MONETARY=en_US.UTF-8    LC_MESSAGES=en_US.UTF-8
 [7] LC_PAPER=en_US.UTF-8      LC_NAME=C                  LC_ADDRESS=C
[10] LC_TELEPHONE=C            LC_MEASUREMENT=en_US.UTF-8 LC_IDENTIFICATION=C

attached base packages:
[1] compiler  stats      graphics  grDevices  utils      datasets  methods    base

other attached packages:
[1] showtext_0.6  showtextdb_2.0 sysfonts_0.8  gridExtra_2.3 ggplot2_3.1.0

loaded via a namespace (and not attached):
 [1] Rcpp_1.0.0      crayon_1.3.4    withr_2.1.2     grid_3.5.3      plyr_1.8.4
 [6] gtable_0.2.0    scales_1.0.0    pillar_1.3.1    rlang_0.3.1     lazyeval_0.2.1
[11] tools_3.5.3     munsell_0.5.0   pkgconfig_2.0.2 colorspace_1.4-0 tibble_2.0.1
```

In this document I calculate parameter estimates from the model Gibbs sampler Markov chains. Control and AI-treatment data sets were modeled separately, with a simple replication scheme without experiment-level covariates.

The model is

$$\begin{aligned}
\mathbf{y}_{i\cdot} &\sim t_{\nu_e, d} \left( \boldsymbol{\mu}_{j[i]\cdot}^{acc}; \boldsymbol{\Sigma}_e \right) \\
\boldsymbol{\mu}_{j\cdot}^{acc} &\sim N_d \left( \boldsymbol{\mu} + \mathbf{u}_{j\cdot} \boldsymbol{\Gamma}; \boldsymbol{\Sigma}_s \right) \\
\boldsymbol{\gamma}_{j\cdot} &\sim t_{\nu_g, d} \left( \mathbf{0}_d; \boldsymbol{\Sigma}_a \right) \\
\boldsymbol{\Sigma}_e^{-1} &\sim W_{d, \nu_0} \left( \left[ \sum_i \left( \mathbf{y}_{i\cdot} - \boldsymbol{\mu}_{j[i]\cdot}^{acc} \right) \left( \mathbf{y}_{i\cdot} - \boldsymbol{\mu}_{j[i]\cdot}^{acc} \right)^T \right]^{-1} \right) \\
\boldsymbol{\Sigma}_s^{-1} &\sim W_{d, \nu_0} \left( \left[ \sum_j \left( \boldsymbol{\mu}_{j\cdot}^{acc} - \boldsymbol{\mu} - \mathbf{u}_{j\cdot} \boldsymbol{\Gamma} \right) \left( \boldsymbol{\mu}_{j\cdot}^{acc} - \boldsymbol{\mu} - \mathbf{u}_{j\cdot} \boldsymbol{\Gamma} \right)^T \right]^{-1} \right) \\
\boldsymbol{\Sigma}_a^{-1} &\sim W_{d, \nu_0} \left( \left[ \boldsymbol{\Gamma}^T \boldsymbol{\Gamma} \right]^{-1} \right),
\end{aligned}$$

I define functions that calculate summaries of Markov chains.

```

> pmode <- cmpfun(function(vec){
+   dst <- density(vec, adjust = 2)
+   mxi <- which(dst$y == max(dst$y))
+   if(length(mxi) > 1){
+     warning("More than one mode in call to pmode(); picking randomly")
+     mxi <- sample(mxi, 1)
+   }
+   dst$x[mxi]
+ })
> HPDint <- cmpfun(function(vec, prob = 0.95){
+   nsamp <- length(vec)
+   if (nsamp <= 2) stop("vector must have length > 2")
+
+   vals <- sort(vec)
+   gap <- max(1, min(nsamp - 1, round(nsamp * prob)))
+   init <- 1:(nsamp - gap)
+   mInd <- which.min(vals[init + gap] - vals[init])
+   res <- c(vals[mInd], vals[mInd + gap])
+   names(res) <- c("lower", "upper")
+   return(res)
+ })

```

Next, I write a function that is similar to `quantile()`, but outputs the mode, lower and upper limits of the 95% and 50% HPD. The `outr` parameter is the probability for the outer margins, default is 95%.

```

> quantileLike <- cmpfun(function(vec, outr = 0.95){
+   if(outr <= 0.5) stop("Outer margin has to be >= 50%")
+   md <- pmode(vec)

```

```

+   hpd95 <- HPDint(vec, outr)
+   hpd50 <- HPDint(vec, 0.5)
+   res   <- c(hpd95[1], hpd50[1], md, hpd50[2], hpd95[2])
+
+   outNm <- paste(c("lower", "upper"), outr*100, sep = "")
+   names(res) <- c(outNm[1], "lower50", "mode", "upper50", outNm[2])
+   return(res)
+ })

```

I start with total accession means.

```

> trtNm <- c("Longest Root Length", "Total Root Length")
> d      <- 2
> Nacc   <- 119
> locDim <- d*Nacc
> nChn   <- 5
> chnLen <- 2000
> chn <- NULL
> addSamp <- cmpfun(function(i, vrNm, exNm, ncl){
+   inFlNm <- paste("chains/", vrNm, "_", exNm, "_IBS_3_3_", i, ".gbin", sep = "")
+   chn <-- rbind(chn,
+                 matrix(.C("GSLmatLoad",
+                           inFlNm, as.integer(chnLen), as.integer(ncl),
+                           out = double(chnLen*ncl))$out,
+                           nrow = chnLen, byrow = T)
+                 )
+   return(NULL)
+ })
> trash <- sapply(1:nChn, addSamp, "LN", "ctrl", locDim)
> accMnCtrlCHN <- chn
> accMnCtrl <- as.data.frame(t(apply(chn, 2, quantileLike)))
> accMnCtrl$trait <- rep(trtNm, times = Nacc)
> accMnCtrlS <- accMnCtrl[order(accMnCtrl[,6], accMnCtrl[,3]),]

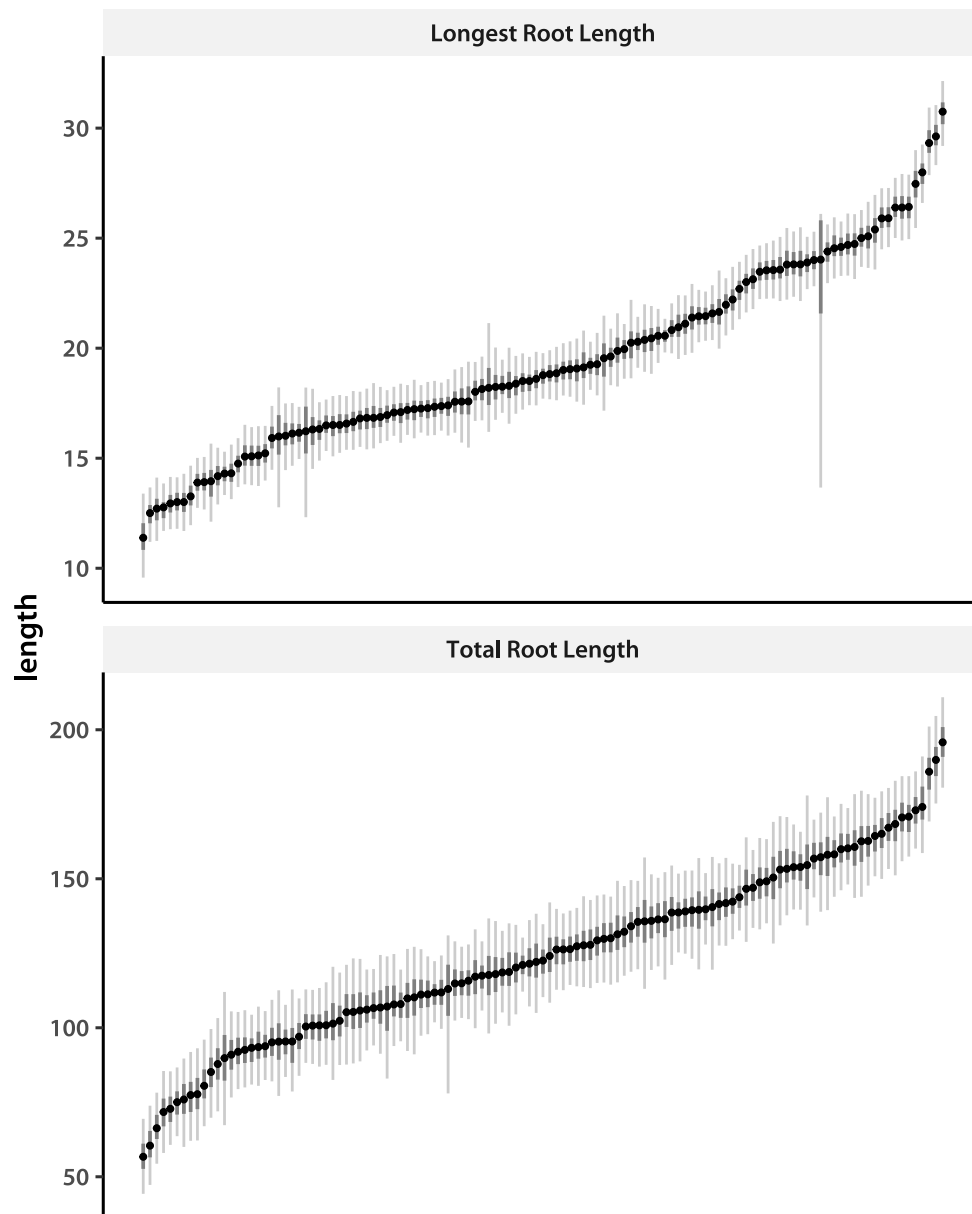
> pdfFlNm <- "lineMeansCtrlNEW.pdf"
> showtext_auto()
> ggplot(data=accMnCtrlS, aes(x=1:nrow(accMnCtrlS), y=mode)) +
+   geom_segment(aes(x=1:nrow(accMnCtrlS), y=lower95,
+                   xend=1:nrow(accMnCtrlS), yend=upper95),
+               color="grey80", size=0.75) +
+   geom_segment(aes(x=1:nrow(accMnCtrlS), y=lower50,
+                   xend=1:nrow(accMnCtrlS), yend=upper50),
+               color="grey50", size=1) +
+   geom_point() +
+   facet_wrap(~trait, scales="free", nrow=2) +
+   theme_classic(base_size=18, base_family="myriad") +

```

```

+   theme(axis.title.x=element_blank(), axis.text.x=element_blank(),
+         axis.ticks.x=element_blank(),
+         strip.background=element_rect(fill="grey95", linetype="blank")) +
+   labs(y="length")
> ggsave(pdfFlNam, width=8, height=10, units="in", device="pdf", useDingbats=F)
> cat("\\includegraphics{" , pdfFlNam, "}\\n\\n", sep="")

```



Process the GEBV estimates the same.

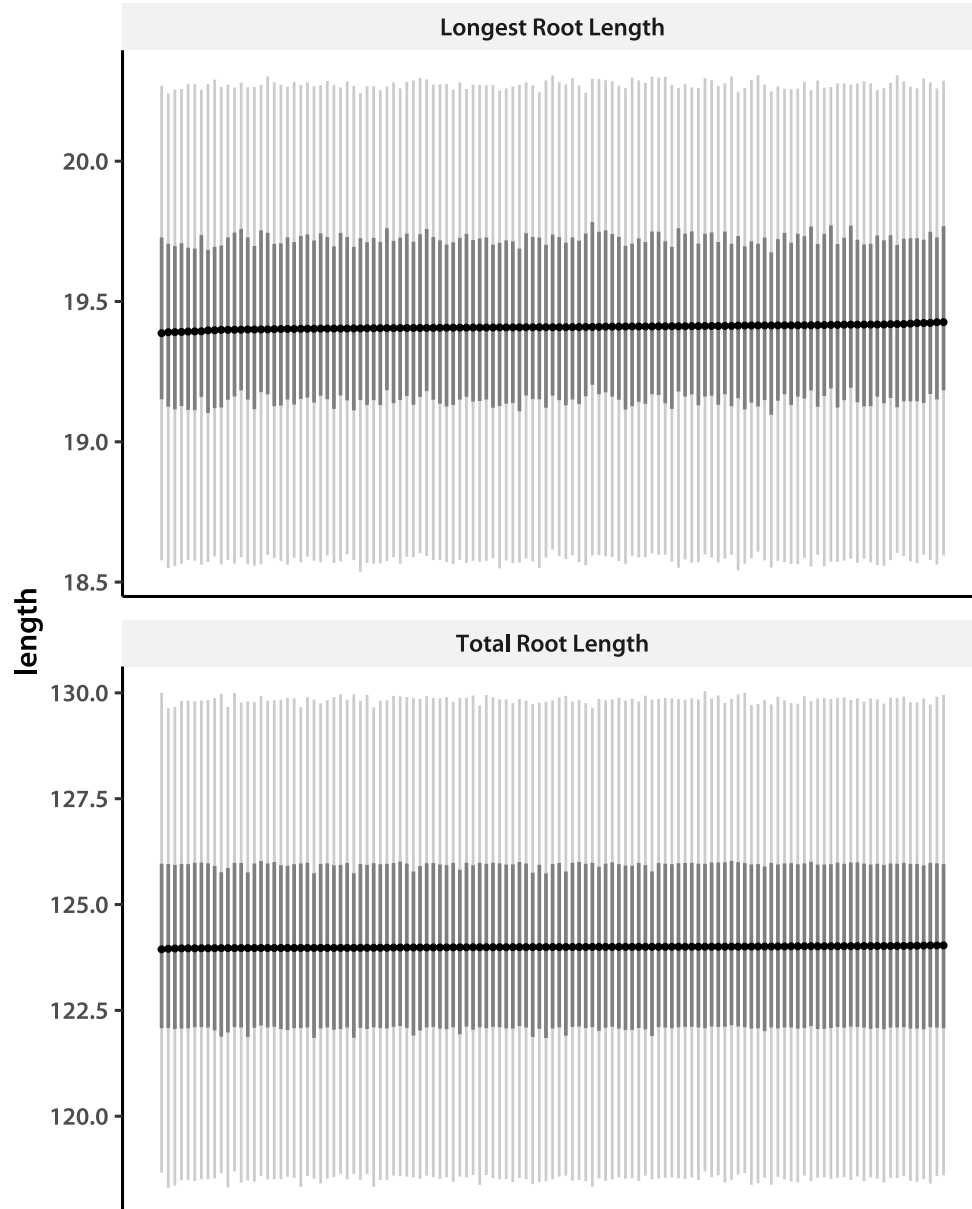
```

> chn      <- NULL
> trash    <- sapply(1:nChn, addSamp, "BV", "ctrl", locDim)

```

```
> chnGEBVctrl      <- chn
> accBVCtrl        <- as.data.frame(t(apply(chn, 2, quantileLike)))
> accBVCtrl$trait  <- rep(trtNam, times = Nacc)
> accBVCtrlS       <- accBVCtrl[order(accBVCtrl[,6], accBVCtrl[,3]),]

> pdfFlNam <- "gebvCtrlNEW.pdf"
> showtext_auto()
> ggplot(data=accBVCtrlS, aes(x=1:nrow(accBVCtrlS), y=mode)) +
+   geom_segment(aes(x=1:nrow(accBVCtrlS), y=lower95,
+     xend=1:nrow(accBVCtrlS), yend=upper95),
+     color="grey80", size=0.75) +
+   geom_segment(aes(x=1:nrow(accBVCtrlS), y=lower50,
+     xend=1:nrow(accBVCtrlS), yend=upper50),
+     color="grey50", size=1) +
+   geom_point() +
+   facet_wrap(~trait, scales="free", nrow=2) +
+   theme_classic(base_size=18, base_family="myriad") +
+   theme(axis.title.x=element_blank(), axis.text.x=element_blank(),
+     axis.ticks.x=element_blank(),
+     strip.background=element_rect(fill="grey95", linetype="blank")) +
+   labs(y="length")
> ggsave(pdfFlNam, width=8, height=10, units="in", device="pdf", useDingbats=F)
> cat("\\includegraphics{" , pdfFlNam, "}" , sep="")
```



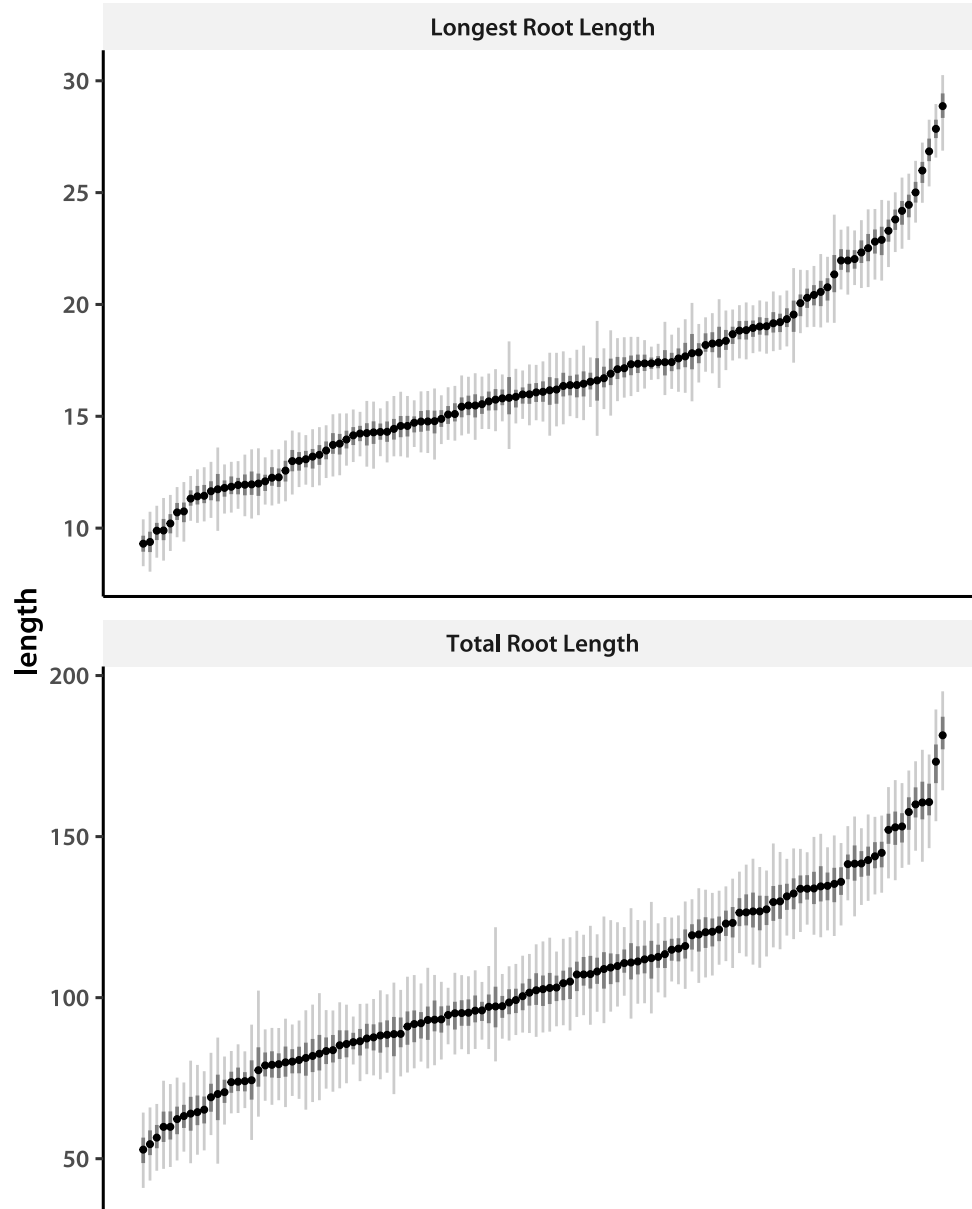
Clearly not much GEBV variation.

Now estimate the parameters from AI treatment data.

```
> chn <- NULL
> trash <- sapply(1:nChn, addSamp, "LN", "expt", locDim)
> accMnExpt <- as.data.frame(t(apply(chn, 2, quantileLike)))
> accMnExpt$trait <- rep(trtNam, times = Nacc)
> accMnExptS <- accMnExpt[order(accMnExpt[,6], accMnExpt[,3]),]

> pdfFlNam <- "lineMeansExptNEW.pdf"
> showtext_auto()
```

```
> ggplot(data=accMnExptS, aes(x=1:nrow(accMnExptS),y=mode)) +
+   geom_segment(aes(x=1:nrow(accMnExptS), y=lower95, xend=1:nrow(accMnExptS), yend=upper95)
+     color="grey80", size=0.75) +
+   geom_segment(aes(x=1:nrow(accMnExptS), y=lower50, xend=1:nrow(accMnExptS), yend=upper50)
+     color="grey50", size=1) +
+   geom_point() +
+   facet_wrap(~trait, scales="free", nrow=2) +
+   theme_classic(base_size=18, base_family="myriad") +
+   theme(axis.title.x=element_blank(), axis.text.x=element_blank(),
+     axis.ticks.x=element_blank(),
+     strip.background=element_rect(fill="grey95", linetype="blank")) +
+   labs(y="length")
> ggsave(pdfFlNam, width=8, height=10, units="in", device="pdf", useDingbats=F)
> cat("\\includegraphics{" , pdfFlNam, "}" , sep="")
```



Now look at the genetic correlation of treatment *vs* control.

```
> gCor <- cmpfun(function(i){
+   res <- cor(cbind(matrix(accMnCtrlCHN[i,], ncol=d, byrow=T),
+                       matrix(chn[i,], ncol=d, byrow=T)))
+   return(res[col(res)>row(res)]) # upper triangle
+ })
> corCHN <- sapply(1:nrow(chn), gCor)
> corHPD <- apply(corCHN, 1, quantileLike)
> corNames <- paste(rep(trtNam, 2), rep(c("CTL", "EXP"), each=d), sep=":")
```

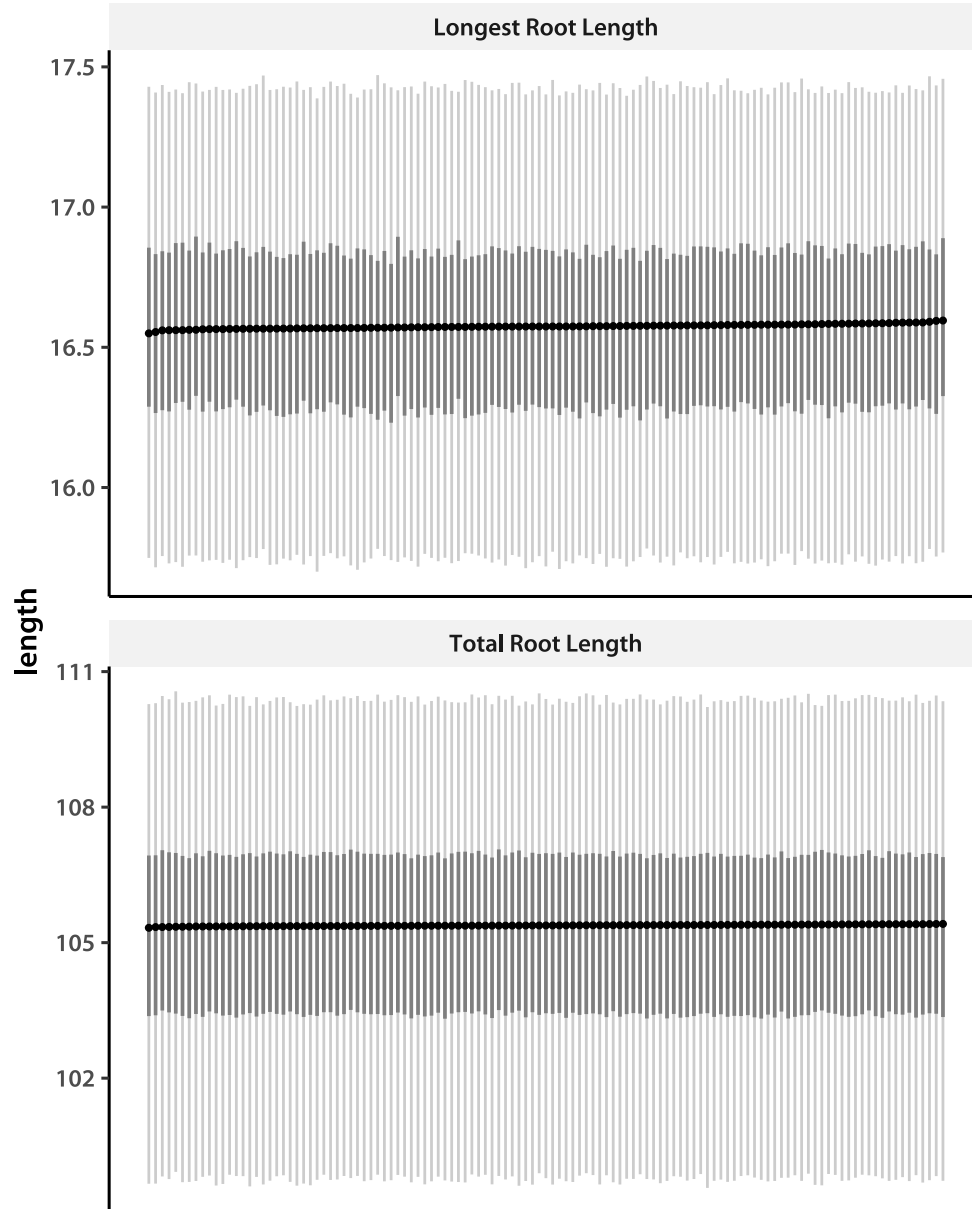
```
> corInd <- col(matrix(1, 2*d, 2*d))>row(matrix(1, 2*d, 2*d))
> colnames(corHPD) <- paste(matrix(corNames, 2*d, 2*d)[corInd],
+                               matrix(corNames, 2*d, 2*d, byrow=T)[corInd], sep="|")
> round(t(corHPD), 4)
```

	lower95	lower50	mode	upper50	upper95
Longest Root Length:CTL Total Root Length:CTL	0.5506	0.5762	0.5926	0.6044	0.6342
Longest Root Length:CTL Longest Root Length:EXP	0.7773	0.8040	0.8158	0.8270	0.8436
Total Root Length:CTL Longest Root Length:EXP	0.4511	0.4852	0.5024	0.5187	0.5505
Longest Root Length:CTL Total Root Length:EXP	0.5964	0.6245	0.6378	0.6526	0.6785
Total Root Length:CTL Total Root Length:EXP	0.7326	0.7569	0.7700	0.7839	0.8109
Longest Root Length:EXP Total Root Length:EXP	0.6439	0.6685	0.6822	0.6942	0.7173

And, finally, GEBV.

```
> chn <- NULL
> trash <- sapply(1:nChn, addSamp, "BV", "expt", locDim)
> chnGEBVexpt <- chn
> accBVExpt <- as.data.frame(t(apply(chn, 2, quantileLike)))
> accBVExpt$trait <- rep(trtNam, times = Nacc)
> accBVExptS <- accBVExpt[order(accBVExpt[,6], accBVExpt[,3]),]

> pdfFlNam <- "gebvExptNEW.pdf"
> showtext_auto()
> ggplot(data=accBVExptS, aes(x=1:nrow(accBVExptS),y=mode)) +
+   geom_segment(aes(x=1:nrow(accBVExptS), y=lower95,
+                               xend=1:nrow(accBVExptS), yend=upper95),
+               color="grey80", size=0.75) +
+   geom_segment(aes(x=1:nrow(accBVExptS), y=lower50,
+                               xend=1:nrow(accBVExptS), yend=upper50),
+               color="grey50", size=1) +
+   geom_point() +
+   facet_wrap(~trait, scales="free", nrow=2) +
+   theme_classic(base_size=18, base_family="myriad") +
+   theme(axis.title.x=element_blank(), axis.text.x=element_blank(),
+         axis.ticks.x=element_blank(),
+         strip.background=element_rect(fill="grey95", linetype="blank")) +
+   labs(y="length")
> ggsave(pdfFlNam, width=8, height=10, units="in", device="pdf", useDingbats=F)
> cat("\\\\includegraphics{" , pdfFlNam, "}\\n\\n", sep="")
```



The correlations are high, but not as high as they were in the previous experiment. I calculate the  $\log_{10}(\text{RRG})$  for each trait, and save point estimates.

```
> accNam <- scan(file="accList.tsv", what = character())
> rrgMode <- log10(accMnExpt$mode/accMnCtrl$mode)
> modeMat <- cbind(matrix(accMnExpt$mode, ncol = 2, byrow = T),
+                  matrix(accMnCtrl$mode, ncol = 2, byrow = T),
+                  matrix(rrgMode, ncol = 2, byrow = T))
> cat(
+   apply(
```

```
+      rbind(c("HDRA_ID", paste(trtNam, "trt", sep = "_"),
+      paste(trtNam, "ctrl", sep = "_"), paste(trtNam, "l10rrg", sep = "_")),
+      cbind(accNam, modeMat)), 1, paste, collapse = "\t"),
+      file = "accessionMeans.tsv", sep = "\n"
+    )
```

## 1 Heritability

I next estimate heritability, both broad-sense and marker-based narrow sense. Because I am using a Student- $t$  model for marker effects, I cannot use  $\Sigma_a$  directly, but have to calculate the variances from the GEBV estimates. In addition to broad-sense heritability, I am interested in the separate contribution of background effects. I first define the functions I need.

```
> makeVar <- cmpfun(function(vec){
+   apply(matrix(vec, ncol=d, byrow=T), 2, var)
+ })
> get.hsqr <- cmpfun(function(vec){
+   vec[1:d]/rowSums(matrix(vec, nrow = d))
+ })
> get.Hsq <- cmpfun(function(vec){
+   (vec[1:d] + vec[(d+1):(2*d)])/rowSums(matrix(vec, nrow = d))
+ })
> get.bck <- cmpfun(function(vec){
+   (vec[(d+1):(2*d)])/rowSums(matrix(vec, nrow = d))
+ })
```

The function `get.hsqr` calculates the marker heritability, which is a kind of narrow-sense heritability. It is

$$h^2 = \frac{\sigma_{\text{GEBV}}^2}{\sigma_{\text{GEBV}}^2 + \sigma_s^2 + \sigma_e^2}$$

for each treatment. The `get.Hsq` function calculates the broad-sense (among-accession) heritability:

$$H^2 = \frac{\sigma_{\text{GEBV}}^2 + \sigma_s^2}{\sigma_{\text{GEBV}}^2 + \sigma_s^2 + \sigma_e^2}$$

The `get.bck` function calculates the fraction of total variance contributed by the background effects alone:

$$FVE_s = \frac{\sigma_s^2}{\sigma_{\text{GEBV}}^2 + \sigma_s^2 + \sigma_e^2}$$

I read in covariance matrix chains, starting with the controls.

```
> diagInd <- diag(matrix(1:(d^2), ncol = d, byrow = T))
> chn1 <- matrix(.C("GSLmatLoad",
```

```
+      "chains/SgS_ctrl_IBS_3_3_1.gbin",
+      as.integer(chnLen), as.integer(d^2), out = double(chnLen*d^2))$out,
+      nrow = chnLen, byrow = T)[,diagInd]
> chn1 <- cbind(chn1,
+      matrix(.C("GSLmatLoad",
+      "chains/SgE_ctrl_IBS_3_3_1.gbin",
+      as.integer(chnLen), as.integer(d^2), out = double(chnLen*d^2))$out,
+      nrow = chnLen, byrow = T)[,diagInd])
> chn2 <- matrix(.C("GSLmatLoad",
+      "chains/SgS_ctrl_IBS_3_3_2.gbin",
+      as.integer(chnLen), as.integer(d^2), out = double(chnLen*d^2))$out,
+      nrow = chnLen, byrow = T)[,diagInd]
> chn2 <- cbind(chn2,
+      matrix(.C("GSLmatLoad",
+      "chains/SgE_ctrl_IBS_3_3_2.gbin",
+      as.integer(chnLen), as.integer(d^2), out = double(chnLen*d^2))$out,
+      nrow = chnLen, byrow = T)[,diagInd])
> chn3 <- matrix(.C("GSLmatLoad",
+      "chains/SgS_ctrl_IBS_3_3_3.gbin",
+      as.integer(chnLen), as.integer(d^2), out = double(chnLen*d^2))$out,
+      nrow = chnLen, byrow = T)[,diagInd]
> chn3 <- cbind(chn3,
+      matrix(.C("GSLmatLoad",
+      "chains/SgE_ctrl_IBS_3_3_3.gbin",
+      as.integer(chnLen), as.integer(d^2), out = double(chnLen*d^2))$out,
+      nrow = chnLen, byrow = T)[,diagInd])
> chn4 <- matrix(.C("GSLmatLoad",
+      "chains/SgS_ctrl_IBS_3_3_4.gbin",
+      as.integer(chnLen), as.integer(d^2), out = double(chnLen*d^2))$out,
+      nrow = chnLen, byrow = T)[,diagInd]
> chn4 <- cbind(chn4,
+      matrix(.C("GSLmatLoad",
+      "chains/SgE_ctrl_IBS_3_3_4.gbin",
+      as.integer(chnLen), as.integer(d^2), out = double(chnLen*d^2))$out,
+      nrow = chnLen, byrow = T)[,diagInd])
> chn5 <- matrix(.C("GSLmatLoad",
+      "chains/SgS_ctrl_IBS_3_3_5.gbin",
+      as.integer(chnLen), as.integer(d^2), out = double(chnLen*d^2))$out,
+      nrow = chnLen, byrow = T)[,diagInd]
> chn5 <- cbind(chn5,
+      matrix(.C("GSLmatLoad",
+      "chains/SgE_ctrl_IBS_3_3_5.gbin",
+      as.integer(chnLen), as.integer(d^2), out = double(chnLen*d^2))$out,
+      nrow = chnLen, byrow = T)[,diagInd])
> sigChnCtrl <- rbind(chn1, chn2, chn3, chn4, chn5)
```

```
> sigChnCtrl <- cbind(t(apply(chnGEBVctrl, 1, makeVar)), sigChnCtrl)
> chnhsqCtrl <- t(apply(sigChnCtrl, 1, get.hsq))
> chnHsqCtrl <- t(apply(sigChnCtrl, 1, get.Hsq))
> chnBckCtrl <- t(apply(sigChnCtrl, 1, get.bck))
```

Add the data from treated accessions.

```
> chn1 <- matrix(.C("GSLmatLoad",
+   "chains/SgS_expt_IBS_3_3_1.gbin",
+   as.integer(chnLen), as.integer(d^2), out = double(chnLen*d^2))$out,
+   nrow = chnLen, byrow = T)[,diagInd]
> chn1 <- cbind(chn1,
+   matrix(.C("GSLmatLoad",
+   "chains/SgE_expt_IBS_3_3_1.gbin",
+   as.integer(chnLen), as.integer(d^2), out = double(chnLen*d^2))$out,
+   nrow = chnLen, byrow = T)[,diagInd])
> chn2 <- matrix(.C("GSLmatLoad",
+   "chains/SgS_expt_IBS_3_3_2.gbin",
+   as.integer(chnLen), as.integer(d^2), out = double(chnLen*d^2))$out,
+   nrow = chnLen, byrow = T)[,diagInd]
> chn2 <- cbind(chn2,
+   matrix(.C("GSLmatLoad",
+   "chains/SgE_expt_IBS_3_3_2.gbin",
+   as.integer(chnLen), as.integer(d^2), out = double(chnLen*d^2))$out,
+   nrow = chnLen, byrow = T)[,diagInd])
> chn3 <- matrix(.C("GSLmatLoad",
+   "chains/SgS_expt_IBS_3_3_3.gbin",
+   as.integer(chnLen), as.integer(d^2), out = double(chnLen*d^2))$out,
+   nrow = chnLen, byrow = T)[,diagInd]
> chn3 <- cbind(chn3,
+   matrix(.C("GSLmatLoad",
+   "chains/SgE_expt_IBS_3_3_3.gbin",
+   as.integer(chnLen), as.integer(d^2), out = double(chnLen*d^2))$out,
+   nrow = chnLen, byrow = T)[,diagInd])
> chn4 <- matrix(.C("GSLmatLoad",
+   "chains/SgS_expt_IBS_3_3_4.gbin",
+   as.integer(chnLen), as.integer(d^2), out = double(chnLen*d^2))$out,
+   nrow = chnLen, byrow = T)[,diagInd]
> chn4 <- cbind(chn4,
+   matrix(.C("GSLmatLoad",
+   "chains/SgE_expt_IBS_3_3_4.gbin",
+   as.integer(chnLen), as.integer(d^2), out = double(chnLen*d^2))$out,
+   nrow = chnLen, byrow = T)[,diagInd])
> chn5 <- matrix(.C("GSLmatLoad",
+   "chains/SgS_expt_IBS_3_3_5.gbin",
+   as.integer(chnLen), as.integer(d^2), out = double(chnLen*d^2))$out,
```

```

+       nrow = chnLen, byrow = T)[,diagInd]
> chn5 <- cbind(chn5,
+       matrix(.C("GSLmatLoad",
+       "chains/SgE_expt_IBS_3_3_5.gbin",
+       as.integer(chnLen), as.integer(d^2), out = double(chnLen*d^2))$out,
+       nrow = chnLen, byrow = T)[,diagInd])
> sigChnExpt <- rbind(chn1, chn2, chn3, chn4, chn5)
> sigChnExpt <- cbind(t(apply(chnGEBVexpt, 1, makeVar)), sigChnExpt)
> chnhsqExpt <- t(apply(sigChnExpt, 1, get.hsq))
> chnHsqExpt <- t(apply(sigChnExpt, 1, get.Hsq))
> chnBckExpt <- t(apply(sigChnExpt, 1, get.bck))
> hsqHPD <- as.data.frame(rbind(t(apply(chnhsqCtrl, 2, quantileLike)),
+       t(apply(chnHsqCtrl, 2, quantileLike)),
+       t(apply(chnhsqExpt, 2, quantileLike)),
+       t(apply(chnHsqExpt, 2, quantileLike))))
> hsqHPD$trait <- rep(rep(trtNam, times=2), times=d)
> hsqHPD$heritability <- rep(rep(c("GEBV", "Broad"), each=d), times=2)
> hsqHPD$treatment <- rep(c("Control", "Treated"), each=2*d)
> fveHPD <- as.data.frame(rbind(t(apply(chnhsqCtrl, 2, quantileLike)),
+       t(apply(chnBckCtrl, 2, quantileLike)),
+       t(apply(chnhsqExpt, 2, quantileLike)),
+       t(apply(chnBckExpt, 2, quantileLike))))
> fveHPD$trait <- rep(rep(trtNam, times=2), times=d)
> fveHPD$variance <- factor(rep(rep(c("Marker", "Background"), each=d), times=2),
+       levels=c("Marker", "Background"))
> fveHPD$treatment <- rep(c("Control", "Treated"), each=2*d)

```

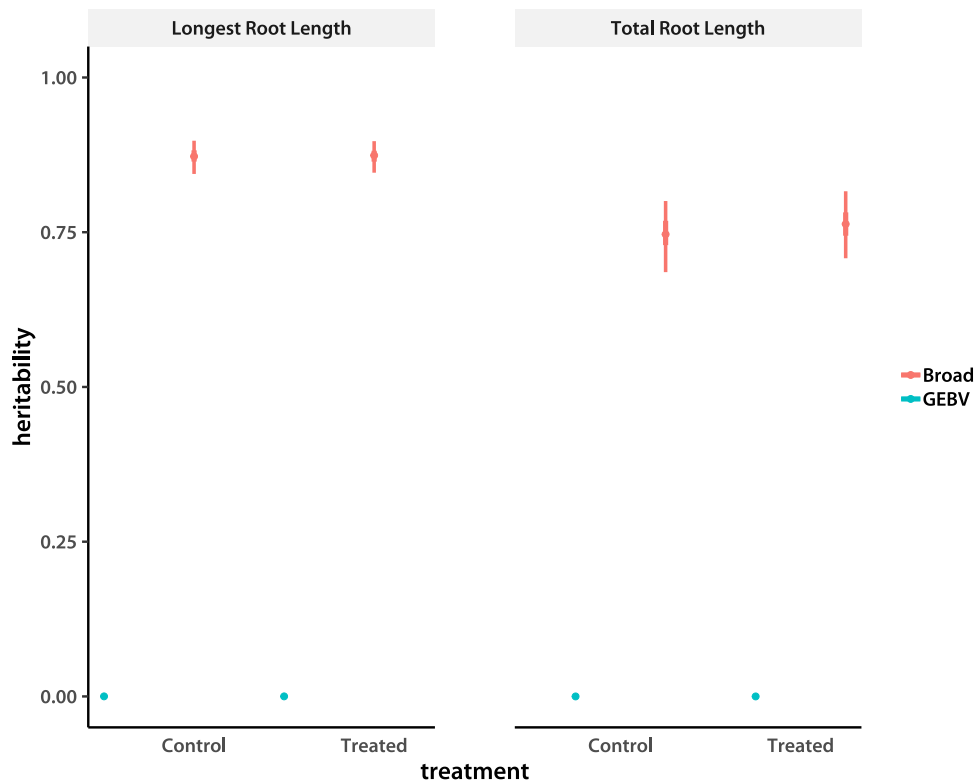
Now plot both kinds of heritabilities on the same panel.

```

> pdfFlNam <- "heritabilityNEW.pdf"
> showtext_auto()
> ggplot(data=hsqHPD, aes(x=1:nrow(hsqHPD), y=mode, color=heritability)) +
+   geom_segment(aes(x=1:nrow(hsqHPD), y=lower95, xend=1:nrow(hsqHPD),
+   yend=upper95, color=heritability),
+   size=1.2) +
+   geom_segment(aes(x=1:nrow(hsqHPD), y=lower50, xend=1:nrow(hsqHPD),
+   yend=upper50, color=heritability),
+   size=1.75) +
+   geom_point(size=2) +
+   facet_wrap(~trait, ncol=2) +
+   theme_classic(base_size=18, base_family="myriad") +
+   theme(legend.title=element_blank(),
+   strip.background=element_rect(fill="grey95", linetype="blank")) +
+   scale_x_continuous(breaks=c(3,7), labels=c("Control", "Treated")) +
+   theme(panel.spacing.x=unit(4.0, "lines"), axis.ticks.x=element_blank()) +
+   ylim(c(0,1)) +

```

```
+ labs(y="heritability", x="treatment")
> ggsave(pdfFlNam, width=10, height=8, units="in", device="pdf", useDingbats=F)
> cat("\\includegraphics{" , pdfFlNam, "}\\n\\n", sep="")
```



Finally plot fractions of variance explained on the same panel.

```
> pdfFlNam <- "fveNEW.pdf"
> showtext_auto()
> ggplot(data=fveHPD, aes(x=1:nrow(fveHPD), y=mode, color=variance)) +
+   geom_segment(aes(x=1:nrow(fveHPD), y=lower95, xend=1:nrow(fveHPD),
+     yend=upper95, color=variance),
+     size=1.2) +
+   geom_segment(aes(x=1:nrow(fveHPD), y=lower50, xend=1:nrow(fveHPD),
+     yend=upper50, color=variance),
+     size=1.75) +
+   geom_point(size=2) +
+   facet_wrap(~trait, ncol=2) +
+   theme_classic(base_size=18, base_family="myriad") +
+   theme(legend.title=element_blank(),
+     strip.background=element_rect(fill="grey95", linetype="blank")) +
+   scale_x_continuous(breaks=c(3,7), labels=c("Control", "Treated")) +
+   theme(panel.spacing.x=unit(4.0, "lines"), axis.ticks.x=element_blank()) +
+   ylim(c(0,1)) +
```

```
+ labs(y="fraction of variance", x="treatment")
> ggsave(pdfFlNam, width=10, height=8, units="in", device="pdf", useDingbats=F)
> cat("\\includegraphics{" , pdfFlNam, "}\n\n", sep="")
```

